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Genetic characterization of antimicrobial resistance in coagulase-negative Staphylococci from bovine mastitis

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Approved by the Vetsuisse Faculty as inaugural dissertation on proposal from
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I dedicate this thesis to my parents who unremittingly supported me during my years of study.

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SUMMARY

Vetsuisse Faculty, University of Bern, 2012

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Genetic characterization of antimicrobial resistance in coagulase-negative staphylococci from bovine mastitis milk

Summary

Coagulase-negative staphylococci (CNS) (n = 417) were isolated from bovine milk and identified by MALDI-TOF MS. Nineteen different species were identified, and *S. xylosus*, *S. chromogenes*, and *S. haemolyticus* were the most abundant species. Resistance to oxacillin (47.0% of the isolates), fusidic acid (33.8%), tiamulin (31.9%), penicillin (23.3%), tetracycline (15.8%), streptomycin (9.6%), erythromycin (7.0%), sulfonamides (5%), trimethoprim (4.3%), clindamycin (3.4%), kanamycin (2.4%) and gentamicin (2.4%) was detected. Resistance to oxacillin was attributed to the *mecA* gene in 9.7% of the oxacillin-resistant isolates. The *mecA* gene was detected in *S. fleurettii*, *S. epidermidis*, *S. haemolyticus*, and *S. xylosus*. Resistance to tetracycline was attributed to the presence of *tet(K)* and *tet(L)*, penicillin resistance to *blaZ*, streptomycin resistance to *str* and *ant(6)-Ia*, erythromycin resistance to *erm(C)*, *erm(B)*, and *msr*. In total, 15.1% of the CNS isolates were multidrug-resistant (i.e., exhibiting resistance to two or more antimicrobials). The remaining CNS isolates were susceptible to antimicrobials commonly used in mastitis treatment. Methicillin-resistant CNS isolates were diverse, as determined by *mecA* gene sequence analysis, *SCCmec* typing, and PFGE. Because this study revealed the presence of multidrug-resistant CNS in a heterogeneous CNS population, we recommend antibiogram analysis of CNS in persistent infections prior to treatment with antimicrobials.

Keywords: methicillin-resistance, CNS, genotyping, antibiotic resistance, *SCCmec*

OVERVIEW AND AIM OF THE THESIS

Mastitis is one of the most important diseases in bovine medicine. It is defined as an infection of the mammary gland and occurs in all mammals. Bovine mastitis has many different causes, and is classified roughly in clinical and subclinical mastitis. Mastitis normally leads to a decrease of the milk yield in cows and therefore to severe economic losses for the farmers.

In modern dairy cow herds, major mastitis pathogens such as *S. agalactiae*, *S. dysgalactiae*, *S. aureus*, and *E. coli* are managed and reduced quite well. However, minor pathogens such as coagulase-negative staphylococci (CNS) or *C. bovis* are increasingly causing mastitis. Many studies showed that CNS are the microorganisms most commonly isolated from bovine milk in many countries, and that they are an important cause of mastitis.

CNS are a group of aerobic living, gram-positive, coagulase-negative bacteria and belong to the cocci genus. Most of the CNS are opportunistic skin commensals, in cows they are often found on udder skin, teat skin and on the teat apices from where they can enter the udder and cause infections.

Antibiotic treatment of the mammary gland is a frequent practice in bovine industry. Not only in cases of acute mastitis but also preventive for dry cow therapy. In Switzerland, β -lactam antimicrobials (including penicillin and cephalosporins), aminoglycosides (gentamicin and neomycin), and macrolides (spiramycin) are commonly used to treat mastitis. Under this selection pressure bacteria such as CNS have the ability to adapt and acquire antibiotic resistance genes.

The aim of our study is to identify the different CNS species in milk from cows with different types of mastitis (clinical and subclinical), to characterize their antimicrobial resistance mechanisms, and to determine whether specific methicillin-resistant and multidrug-resistant CNS clones are common in dairy cows.

Further, this project will allow implementing a new and cheap method for the rapid identification of CNS at species level in the veterinary diagnostic. The results will also give an overview of the prevalence and the phenotypic and genotypic antibiotic resistance profile of CNS isolated from mastitis milk in Switzerland.

THESIS

**Genetic characterization of antimicrobial resistance in coagulase-negative
Staphylococci from bovine mastitis**

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Key words: methicillin-resistant, CNS, genotyping, antibiotic resistance, *SCCmec*

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1.) ABSTRACT

Coagulase-negative staphylococci (CNS) (n = 417) were isolated from bovine milk and identified by MALDI-TOF MS. Nineteen different species were identified, and *S. xylosus*, *S. chromogenes*, *S. haemolyticus*, and *S. sciuri* were the most abundant species. Resistance to oxacillin (47.0% of the isolates), fusidic acid (33.8%), tiamulin (31.9%), penicillin (23.3%), tetracycline (15.8%), streptomycin (9.6%), erythromycin (7.0%), sulfonamides (5%), trimethoprim (4.3%), clindamycin (3.4%), kanamycin (2.4%) and gentamicin (2.4%) was detected. Resistance to oxacillin was attributed to the *mecA* gene in 9.7% of the oxacillin-resistant isolates. The remaining oxacillin-resistant CNS did not contain the *mecC* gene or *mecAI* promoter mutations. The *mecA* gene was detected in *S. fleurettii*, *S. epidermidis*, *S. haemolyticus*, and *S. xylosus*. Resistance to tetracycline was attributed to the presence of *tet(K)* and *tet(L)*, penicillin resistance to *blaZ*, streptomycin resistance to *str* and *ant(6)-Ia*, erythromycin resistance to *erm(C)*, *erm(B)*, and *msr*. Resistance to tiamulin and fusidic acid could not be attributed to an acquired resistance gene. In total, 15.1% of the CNS isolates were multidrug-resistant (i.e., exhibiting resistance to two or more antimicrobials). The remaining CNS isolates were susceptible to antimicrobials commonly used in mastitis treatment. Methicillin-resistant CNS isolates were diverse, as determined by *mecA* gene sequence analysis, staphylococcal cassette chromosome *mec* (SCC*mec*) typing, and pulsed-field gel electrophoresis (PFGE). Arginine catabolic mobile element (ACME) types 1 and 3 were detected in both methicillin-resistant and susceptible *S. epidermidis* and were associated with sequence types ST59 and ST111. Because this study revealed the presence of multidrug-resistant CNS in a heterogeneous CNS population, we recommend antibiogram analysis of CNS in persistent infections prior to treatment with antimicrobials.

2.) INTRODUCTION

Coagulase-negative staphylococci (CNS) are the microorganisms most commonly isolated from bovine milk in many countries, and they are an important cause of mastitis (Rajala-Schultz et al., 2009; Pyörälä and Taponen, 2009; Piessens et al., 2011; De Vliegher et al., 2012). CNS are opportunistic pathogens that are usually diagnosed as a group without species identifications. They cause subclinical intramammary infections that result in an increase in the somatic cell count and reduced milk quality, leading to economic losses (Pyörälä and Taponen, 2009). Because simple subclinical CNS infections can be self-limiting, they are usually not treated with antibiotics. However, CNS often appear with other major pathogens such as *Staphylococcus aureus*, *Streptococcus* species or coliform bacteria. In these cases and in persistent CNS infections, the cows undergo antimicrobial treatment. Currently, β -lactam antimicrobials (including penicillin and cephalosporins), aminoglycosides (gentamicin and neomycin), and macrolides (spiramycin) are commonly used to treat mastitis in Switzerland (Büttner et al., 2011). Resistance to these antibiotics has been increasingly reported in CNS associated with bovine mastitis (Walther and Perreten, 2007; Sawant et al., 2009; Sampimon et al., 2011). CNS may also harbor antimicrobial resistance elements and pathogenicity islands, such as the staphylococcal cassette chromosome (SCC*mec*) element (Wielders et al., 2001; Barbier et al., 2010; Tsubakishita et al., 2010) and the arginine catabolic mobile element (ACME) (Diep et al., 2006; Diep et al., 2008; Miragaia et al., 2009) that can be transferred to *S. aureus*. ACMEs are genomic islands in *S. epidermidis* that have been associated with host colonization, fitness and pathogenicity. ACME mobility is associated with recombinase genes present on the SCC*mec* elements (Goering et al., 2007; Diep et al., 2008). SCC*mec* contains the *mec* genes like *mecA* and *mecC* (*mecA*_{LGA251}), which encode alternative penicillin binding proteins (PBP 2a) and confer resistance to all β -lactam antimicrobials (García-Álvarez et al., 2011; Ito et al., 2012). In *S. sciuri*, the *mecA* gene homologue *mecA1* is a native gene that is not part of the *mec* gene complex (Couto et al., 1996; Wu et al., 1998; Couto et al., 2000; Wu et al., 2001; Tsubakishita et al., 2010). Most *S. sciuri* isolates are susceptible to β -lactam antimicrobials. However, alterations in the promoter regions of *mecA1* up-regulate *mecA1* expression and methicillin resistance (Wu et al., 2001; Couto et al., 2003; Wu et al., 2005). Methicillin-resistant staphylococci are often also resistant to other classes of drugs such as the aminoglycosides and the macrolides (Woodford 2005). Nevertheless, little is known about the molecular mechanisms of the antimicrobial resistance (Lüthje and Schwarz, 2006) or the genetic backgrounds of multidrug-resistant CNS strains in bovine milk.

We have identified the different CNS species in milk from cows with clinical and subclinical bovine mastitis, characterized their antimicrobial resistance mechanisms, and determined whether specific methicillin-resistant and multidrug-resistant CNS clones are common in dairy cows.

3.) MATERIALS AND METHODS

Origin of the milk samples

CNS ($n = 417$) were isolated from milk ($n = 370$) obtained from cows diagnosed with clinical ($n = 115$) and subclinical ($n = 255$) mastitis and control samples ($n = 47$) in Switzerland. Control samples were collected from cows that had suffered from mastitis previously and had been treated; the control milk samples contained $< 150,000$ cells/ml. The 417 isolates came from 363 different cows and from two different mammary quarters of seven cows. The 363 cows originated from 195 different farms (n_f) in the cantons of Berne ($n_f = 91$), Jura ($n_f = 56$), Fribourg ($n_f = 26$), Vaud ($n_f = 8$), Lucerne ($n_f = 5$), Valais ($n_f = 4$), Solothurn ($n_f = 3$), Aargau ($n_f = 1$) and Thurgau ($n_f = 1$). In 47 cases, two different CNS strains were found in the same milk sample.

Isolation and identification of CNS

Milk samples were centrifuged at $590\times g$ for 10 min at room temperature. The milk pellets were cultivated on tryptone soy agar containing 5% defibrinated sheep blood (TSA-SB, Becton, Dickinson and Company, Franklin Lakes, NJ) and incubated at 37°C for 18 to 24 hours. Staphylococci were selected based on colony morphology, gram-positive staining of cocci and catalase production and were subcultured on TSA-SB.

The isolates were identified by Matrix-Assisted-Laser-Desorption/Ionization-Time-Of-Flight-Mass-Spectrometry (MALDI-TOF MS) analysis using the ethanol-formic acid extraction method for better resolution (Application Note, MT-80, Bruker) (Microflex LT, Bruker Daltonics GmbH, Bremen). Species identification was considered valid when the matching score with reference spectra of the MALDI Biotyper v3.0 database (Bruker) was ≥ 2 , according to the criteria proposed by the manufacturer. Isolates whose measured spectra had score < 2.0 were further identified by DNA sequencing of the 16S rDNA (Kuhnert et al., 1996). The CNS strains were stored at -80°C in trypticase soy medium containing 30% glycerin (Becton, Dickinson and Company, Franklin Lakes, NJ).

DNA extraction and amplification

To obtain total DNA, cells were incubated in 100 µl TE buffer containing 0.1 mg/ml lysostaphin for 15 min at 37°C, and then, 450 µl lysis buffer (0.1 M Tris-HCl pH 8.5, 0.05% Tween 20, 0.24 mg/ml proteinase K) was added and incubated at 60°C for 45 min. The DNA was then denatured at 95°C for 15-min.

PCRs were performed with HOT FIREPol® DNA Polymerase (Solis BioDyne, Tartu, Estonia), and the primers and conditions are listed in Table 1.

Antimicrobial resistance tests

The CNS isolates were tested for antimicrobial susceptibility with the broth microdilution technique (Clinical and Laboratory Standards Institute 2009) using Sensititre susceptibility plates NLEUST (Trek Diagnostics Systems, East Grinstead, West Sussex; MCS diagnostics BV, JL Swalmen, The Netherlands) that contained the following 19 antimicrobials: chloramphenicol, ciprofloxacin, clindamycin, dalfopristin-quinupristin, erythromycin, fusidic acid, gentamicin, kanamycin, linezolid, mupirocin, oxacillin, penicillin, rifampicin, streptomycin, sulfamethoxazole, tetracycline, tiamulin, trimethoprim, and vancomycin. The resistance breakpoints were those proposed for CNS in the EUCAST guidelines (www.eucast.org) (Table 2). The production of β-lactamase was tested on nitrocefin dry slides (Becton, Dickinson and Company, Franklin Lakes, NJ) using colonies grown on Mueller Hinton agar for 18 h at 37°C with 0.05 µg penicillin per ml to induce β-lactamase production (Schnellmann et al., 2006). The antimicrobial resistance genes were detected by a custom-made microarray (AMR+ve-2 array tubes, Alere technologies GmbH, Jena, Germany) (Perreten et al., 2005). The microarray results were analyzed using the IconoClust program (Alere GmbH), and the data were interpreted visually.

Characterization of the *mec* genes and SCC*mec* elements

All isolates displaying a MIC for oxacillin above the resistance breakpoint (MIC > 0.25 µg/ml), which suggests the presence of an alternative PBP (CLSI, EUCAST), were additionally tested by PCR for the *mecA*, *mecA1* and *mecC* genes (García-Álvarez et al., 2011; Ito et al., 2012) using the primers listed in Table 1. The complete nucleotide sequences of the *mecA* genes were obtained by PCR amplification with the *mecA*-F7 and *mecA*-R7 primers (Table 1). Sequencing was performed on an ABI PRISM 3100 genetic analyzer (Applied Biosystems, Foster City, CA). The SCC*mec* types were determined by the Kondo method (Kondo et al., 2007).

Analysis of the *mecA1* promoter region in *S. sciuri*

S. sciuri isolates carrying a *mecA1* homologue (n = 37) were analyzed for a point mutation (Wu et al., 2001) in the promoter region by restriction analysis of PCR products amplified with primers *mecAscK1-F* and *mecAsc-R* (Table 1). The 335 bp PCR product was tested for *PsiI* cleavage using the manufacturer's suggested conditions (New England BioLabs, Beverly, MA). *PsiI* recognizes the mutated promoter sequences TATAAT but not the wild-type sequence TATATT.

Genotyping of methicillin-resistant CNS

Methicillin-resistant, *mecA*-positive CNS isolates and multiresistant, *mecA*-negative *S. epidermidis* isolates were genotyped by pulsed-field gel electrophoresis (PFGE). Analysis of *SmaI*-digested chromosomal DNA was performed as described previously (Schnellmann et al., 2006). Digested DNA was separated by gel electrophoresis in a contour-clamped homogenous electric field DRIII device (Bio-Rad Laboratories, Inc., Richmond, CA) with a ramped pulse time of 5 to 40 s at 6 V/cm for 21 h at 12°C. The lambda ladder PFG marker (New England BioLabs) was used as a size reference. The digital PFGE pattern images were analyzed with the BioNumerics software (Applied Maths, Kortrijk, Belgium), and the PFGE profiles were defined by the DNA banding patterns and criteria of Tenover et al. (Tenover et al., 1995).

Multilocus sequence typing (MLST)

All *S. epidermidis* isolates (n=15) were examined by multilocus sequence typing (MLST) which is based on the sequencing of internal fragments of seven housekeeping genes (Thomas et al., 2007). Allele and sequence type (ST) numbers were assigned according to the *S. epidermidis* MLST database (<http://sepidermidis.mlst.net/>).

Detection of *ica* and ACME

The *S. epidermidis* isolates (n=15) were tested by PCR for the biofilm operon *ica* (Gu et al., 2005) and the arginine catabolic mobile element (ACME). The presence and type of ACME was determined using the primer pairs AIPS.27 and AIPS.28 for *arcA* and AIPS.45 and AIPS.46 for *opp3* gene clusters (Diep et al., 2008).

Statistical analysis

Antimicrobial resistance phenotypes (Table 2) were compared using the Fisher's exact test. This test is useful when the sample size is small (zero in some cells), and the test evaluates the hypothesis that the 2 column percentages in a 2x2 table are equal. Statistical analysis was

performed with the statistical software NCSS 2007 (www.ncss.com). The overall level of statistical significance was set to $P < 0.05$.

4.) RESULTS

Incidence and identification of CNS

In total, 97.8% of the CNS isolates ($n = 408$) were clearly identified at the species level by MALDI-TOF MS analysis. The most frequent CNS species were *S. xylosus*, *S. chromogenes*, *S. sciuri*, and *S. haemolyticus* (Table 3). The remaining 2.2% were identified by 16S rDNA analysis as *S. chromogenes* ($n=1$), *S. saprophyticus* ($n=1$) or new *Staphylococcus* species ($n=7$) (Table 3). Neither clinical nor subclinical mastitis could be correlated with the presence of individual bacterial species. Similar species were identified in control milk samples with low somatic cell counts. Of the 417 CNS isolated, 268 isolates (64.3%) were the only species present in the milk from which they originated. Eighteen CNS isolates were co-purified with *S. aureus* (4.3%), while 83 isolates were present with *Streptococcus* species (19.9%). Altogether, 48 isolates (11.5%) were coincident in milk with at least one other bacterium (e.g., *A. pyogenes*, *E. coli*, *C. bovis* or a mix of more than 3 different bacteria).

Analysis of antimicrobial resistance phenotypes and genotypes

Oxacillin resistance, which is the indicator of *mec* gene-mediated methicillin resistance, was the most frequent resistance phenotype (47.0% of isolates), followed by resistance to fusidic acid (34.1%), tiamulin (31.9%), penicillin (23.3%), tetracycline (15.8%), streptomycin (9.6%), and erythromycin (7.0%) (Table 2). Resistance to two or more antibiotics was observed in 15.1% of the CNS isolates. Multiresistant isolates were found in milk from clinical ($n = 21$) and subclinical ($n = 34$) mastitis cases and control milk ($n = 8$). Oxacillin resistance was significantly more frequent in clinical mastitis isolates (56.5%) than in subclinical mastitis isolates (43.9%), whereas sulfamethoxazole resistance was significantly more frequent in clinical mastitis isolates (11.3%) than in control milk. No significant difference in resistance between isolates was observed for the other antimicrobials tested (Table 2).

Oxacillin resistance was attributed to the *mecA* gene present in 9.7% ($n=19$) of the oxacillin resistant isolates ($n = 196$). The *mecA* gene was detected in *S. fleurettii* (11/12), *S. epidermidis* (6/15), *S. haemolyticus* (1/37), and *S. xylosus* (1/155) isolates. The *mecA* or *mecC* gene was not detected in the other 177 oxacillin-resistant isolates (90.3%) (Table 4). These isolates

exhibited an oxacillin MIC of 0.5 or 1.0 µg/ml, which is just above the clinical resistance breakpoint.

All *S. sciuri* isolates (n = 37) contained the *mecA1* gene, and exhibited low-level resistance to oxacillin (MIC between 0.5 and 1.0 µg/ml). The low oxacillin resistance was due to the absence of the T-A mutation in the -10 promoter sequence (Wu et al., 2001), as demonstrated by *PsiI* restriction analysis.

Resistance to other antimicrobials was attributed to the beta-lactamase gene *blaZ*, the tetracycline efflux genes *tet(L)* and *tet(K)*, the streptomycin adenylyltransferase and nucleotidyltransferase genes *ant(6)-Ia* and *str*, the chloramphenicol acetyltransferase genes *cat_{pC221}* and *cat_{pC223}*, the gentamicin acetyltransferase gene *aac(6')-Ie*, the kanamycin-neomycin phosphotransferase genes *aph(2')-Ia* and *aph(3')-III*, the macrolide and lincosamide 23S rRNA methylase genes *erm(B)* and *erm(C)*, the macrolide efflux gene *msr*, the lincosamide nucleotidyltransferase gene *lnu(A)* and the trimethoprim-resistant dihydrofolate reductase genes *dfr(A)*, *dfr(D)*, *dfr(G)*, and *dfr(K)* (Table 4). In a few strains, resistance to erythromycin, clindamycin, streptomycin, gentamicin, chloramphenicol, and trimethoprim could not be explained by the presence of any of the tested genes, suggesting new antimicrobial resistance mechanisms in CNS (Table 4). Resistance to fusidic acid was not due to the known fusidic acid resistance genes *fus(B)*, and *fus(C)* suggesting the appearance of new resistance genes or mutations in the elongation factor G *fus(A)* (Farrell et al., 2011). Similarly, no known tiamulin resistance genes (*vga* or *lsa*) were detected in the tiamulin-resistant strains. Resistance to sulfonamides was not further characterized.

Multiple combinations of these genes were found in 4.8% of the CNS isolates. The most frequent resistance genes detected in combination were those conferring resistance to oxacillin, tetracycline, penicillin, streptomycin, gentamicin, kanamycin, erythromycin and clindamycin (Table 5). The presence of several genes in one isolate was linked to the presence of *mecA* in *S. epidermidis* (n = 6), *S. sciuri* (n = 3), *S. haemolyticus* (n = 1), and *S. fleurettii* (n = 1). The other *mecA* positive isolates (*S. fleurettii* (n = 10), and *S. xylosus* (n = 1)) contained only the *mecA* gene. CNS isolates lacking *mecA*, but containing several other resistance genes were classified as *S. chromogenes* (n = 3), *S. epidermidis* (n = 3), *S. haemolyticus* (n = 1), and *S. warneri* (n = 2) (Table 5). All CNS isolates containing several resistance genes were obtained from cows presenting with subclinical or clinical mastitis (Table 5). Although one *S. xylosus* isolate contained a *mecA* gene, multidrug resistance was never observed in *S. xylosus*, the most frequently detected CNS in our study.

Genotyping of methicillin-resistant CNS

CNS strains containing the *mecA* gene (n = 19) were further analyzed for *mecA* sequences, clonality, SCC*mec* and ACME types. Two different *mecA* genes with slight sequence differences were detected in *S. epidermidis* isolates (Fig.1). Four different *mecA* genes that slightly differed from each other were found in *S. fleurettii* (Fig. 1). The *mecA* genes detected in *S. haemolyticus* and *S. xylosus* also differed slightly from each other and from those in *S. epidermidis* and *S. fleurettii* (Fig.1).

PFGE analysis of methicillin-resistant *S. epidermidis* (n=6) and *S. fleurettii* (n=11) showed that strains of the same species were not clonally related, except for four *S. fleurettii* from four different farms that showed two similar PFGE patterns (Figure 1). Methicillin-resistant *S. epidermidis* belonged to ST59 (n = 2), ST55, ST89, and the new strains ST452 and ST454, while *S. epidermidis* strains lacking the *mecA* gene (MSSE), and displaying a multidrug-resistance profile belonged to ST111 (n = 4), ST184 (n = 1), ST293 (n = 1) and to the new strain ST453 (n = 1). MSSE also showed a different PFGE profile (data not shown).

Several different SCC*mec* elements were detected among the methicillin-resistant CNS isolates (Fig. 1). *S. epidermidis* strains contained SCC*mec* IV (n = 4), SCC*mec* V (n = 1), and one non-typeable SCC*mec* related to types IV and VI. *S. haemolyticus* (n = 1) and *S. xylosus* (n = 1) both contained a non-typeable SCC*mec*. The SCC*mec* elements of *S. fleurettii* strains (n = 11) could not be assigned to a known SCC*mec* element. These SCC*mec* elements contained the known *mec* gene complex A, but lacked a known *ccr* element. The *S. haemolyticus* and the *S. xylosus* strains were not typeable (Fig. 1). Two methicillin-resistant *S. epidermidis* strains, ST59-SCC*mec* IV and ST454-SCC*mec* V, contained a type 2 and type 3 ACME, respectively. ACME type 1 was detected in four MSSE ST111 and in one MSSE ST456. None of the *S. epidermidis* isolates carried the biofilm-formation operon *ica* (Fig. 1).

5.) DISCUSSION

Many diverse CNS species have been identified in bovine milk, and MALDI-TOF MS is a reliable and rapid method to identify CNS species (Loonen et al., 2012). We observed that a short ethanol-formic acid extraction is necessary for accurate identification. CNS in milk were frequently detected as single bacterial species, suggesting that these species were the infectious agents. However, the presence of these CNS species was not correlated with a clinical mastitis diagnosis (Table 3). The most frequently occurring species in this study were *S. xylosus*, *S. chromogenes*, *S. sciuri*, and *S. haemolyticus*, as reported in other studies (Waller

et al., 2011; Piessens et al., 2011; Supré et al., 2011). Although *S. xylosus* is not known to cause mastitis, it was detected in 35.9% of the milk samples in our study and, as a single species in 22.8% of those samples, emphasizing the previous conclusions that *S. xylosus* is an underestimated pathogenic CNS in bovine mastitis (Supré et al., 2011). Additionally, two-thirds of the *S. xylosus* isolates were resistant to oxacillin but lacked a known *mec* gene. The absence of a *mecA* gene was also observed in other oxacillin-resistant CNS isolates (Table 4). For these oxacillin-resistant isolates, *mec*-independent mechanisms, which may not be related to an acquired resistance gene, explain the decreased susceptibility to oxacillin with MIC in the range of 0.5 – 2.0 µg/ml. Oxacillin resistance in *S. sciuri* may also depend on *mecA1* gene overexpression. Alterations to the promoter region of *mecA1* are necessary for high level *mecA1* expression and oxacillin resistance in *S. sciuri* (Wu et al., 2001; Couto et al., 2003; Wu et al., 2005). In our study, none of the *S. sciuri* strains contained the -10 promoter mutation that is associated with oxacillin resistance (Wu et al., 2001). However, the MICs for these isolates were between 0.5 and 1.0 µg/ml, which are values above the CLSI and the EUCAST resistance breakpoints. The clinical and therapeutic relevance of decreased susceptibility to oxacillin remains to be clarified. The oxacillin breakpoint may be set to low to properly gauge resistance in CNS from bovine mastitis cases (Fessler et al., 2010), and detection of acquired *mec* genes may be necessary for correct interpretation of the antibiogram.

The *mecA* gene was detected in *S. epidermidis*, *S. fleurettii*, *S. haemolyticus* and *S. xylosus*. Three different *mecA* genes that differ from each other by only a few base pairs were found in the methicillin-resistant *S. fleurettii* and *S. epidermidis* isolates, suggesting the independent acquisition of SCC*mec* elements in these species. This conclusion is supported by the observation that the different SCC*mec* elements were detected in individual *S. epidermidis* isolates. All but one *S. fleurettii* isolate contained a *mecA* gene associated with a non-typeable SCC*mec* element. Genetic diversity was confirmed by PFGE, which showed that with the exception of two pairs of *S. fleurettii* with similar PFGE profiles, all other methicillin-resistant CNS isolates had different PFGE profiles. Despite different PFGE profiles, *S. epidermidis* isolates belonging to the ST111 and ST59 groups were predominant in bovine mastitis cases, suggesting that a specific clonal lineage of *S. epidermidis* has adapted to the udder environment (Piessens et al., 2012). Half of the *S. epidermidis* isolates contained an ACME operon, which may be involved in host adaption in humans (Miragaia et al., 2009). ACMEs were mainly observed in the *S. epidermidis* ST 59 or ST 111 groups, suggesting that it may also play a role in host adaption in cows. Additionally, *S. epidermidis* was the predominant CNS species among those that contained multiple antimicrobial resistance genes. Multiple

resistance genes were also found in *S. sciuri*, *S. chromogenes*, *S. haemolyticus*, and *S. fleurettii*, and these genes were frequently associated with the presence of the *mecA* gene. Genes conferring resistance to clinically relevant antimicrobials such as the penicillins, macrolides, lincosamides, and aminoglycosides were also detected. In total, 15.1% of the isolates studied were resistant to more than two antimicrobials, and some strains were virtually resistant to all antimicrobials authorized for the treatment of mastitis. The remaining CNS isolates were susceptible to antimicrobials commonly used in mastitis treatment.

Our study demonstrated that CNS species in milk from cows experiencing mastitis are generally susceptible to the antimicrobials commonly used for treatment. However, CNS have the potential to acquire resistance genes, leading to therapeutic failures. Some multidrug-resistant isolates, especially *S. epidermidis*, *S. chromogenes* and *S. haemolyticus*, are present in bovine mastitis milk and may resist antimicrobial treatment. An antibiogram is therefore recommended for targeted therapy, and chronically infected cows should be culled from the herd.

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7.) REFERENCES

- Barbier, F., E. Ruppe, D. Hernandez, D. Lebeaux, P. Francois, B. Felix, A. Desprez, A. Maiga, P. L. Woerther, K. Gaillard, C. Jeanrot, M. Wolff, J. Schrenzel, A. Andremont, and R. Ruimy. 2010. Methicillin-resistant coagulase-negative *staphylococci* in the community: high homology of SCCmec IVa between *Staphylococcus epidermidis* and major clones of methicillin-resistant *Staphylococcus aureus*. *J. Infect. Dis.* 202: 270-281.
- Büttner, S., O. Flechtner, C. Müntener, and G. Overesch. Bericht über den Vertrieb von Antibiotika in der Veterinärmedizin und das Antibiotikaresistenzmonitoring bei Nutztieren in der Schweiz (ARCH-VET 2010). 2011. Federal Veterinary Office and Swissmedic, Bern, Switzerland. www.swissmedic.ch/archvet-d.asp.
- Clinical and Laboratory Standards Institute. 2009. Methods for dilution antimicrobial susceptibility tests for bacteria that grow aerobically, 8th ed., vol. 29, no. 2. Approved standard M07-A8. Wayne, PA: Clinical and Laboratory Standards Institute.
- Couto, I., H. de Lencastre, E. Severina, W. Kloos, J. A. Webster, R. J. Hubner, I. S. Sanches, and A. Tomasz. 1996. Ubiquitous presence of a *mecA* homologue in natural isolates of *Staphylococcus sciuri*. *Microb. Drug Resist.* 2: 377-391.
- Couto, I., I. S. Sanches, R. Sá-Leão, and H. de Lencastre. 2000. Molecular characterization of *Staphylococcus sciuri* strains isolated from humans. *J. Clin. Microbiol.* 38: 1136-1143.
- Couto, I., S. W. Wu, A. Tomasz, and H. de Lencastre. 2003. Development of methicillin resistance in clinical isolates of *Staphylococcus sciuri* by transcriptional activation of the *mecA* homologue native to s. *J. Bacteriol.* 185: 645-653.
- De Vlieghe, S., L. K. Fox, S. Piepers, S. McDougall, and H. W. Barkema. 2012. Invited review: Mastitis in dairy heifers: nature of the disease, potential impact, prevention, and control. *J. Dairy Sci.* 95: 1025-1040.
- Diep, B. A., S. R. Gill, R. F. Chang, T. H. Phan, J. H. Chen, M. G. Davidson, F. Lin, J. Lin, H. A. Carleton, E. F. Mongodin, G. F. Sensabaugh, and F. Perdreau-Remington. 2006. Complete genome sequence of USA300, an epidemic clone of community-acquired methicillin-resistant *Staphylococcus aureus*. *Lancet* 367: 731-739.
- Diep, B. A., G. G. Stone, L. Basuino, C. J. Graber, A. Miller, S. A. des Etages, A. Jones, A. M. Palazzolo-Ballance, F. Perdreau-Remington, G. F. Sensabaugh, F. R. Deleo, and H. F. Chambers. 2008. The arginine catabolic mobile element and staphylococcal chromosomal cassette *mec* linkage: convergence of virulence and resistance in the USA300 clone of methicillin-resistant *Staphylococcus aureus*. *J. Infect. Dis.* 197: 1523-1530.
- Farrell, D. J., M. Castanheira, and I. Chopra. 2011. Characterization of global patterns and the genetics of fusidic acid resistance. *Clin. Infect. Dis.* 52 (S7): 487-492.
- Fessler, A. T., C. Billerbeck, K. Kadlec, and S. Schwarz. 2010. Identification and characterization of methicillin-resistant coagulase-negative staphylococci from bovine mastitis. *J. Antimicrob. Chemother.* 65: 1576-1582.

- García-Álvarez, L., M. T. Holden, H. Lindsay, C. R. Webb, D. F. Brown, M. D. Curran, E. Walpole, K. Brooks, D. J. Pickard, C. Teale, J. Parkhill, S. D. Bentley, G. F. Edwards, E. K. Girvan, A. M. Kearns, B. Pichon, R. L. Hill, A. R. Larsen, R. L. Skov, S. J. Peacock, D. J. Maskell, and M. A. Holmes. 2011. Meticillin-resistant *Staphylococcus aureus* with a novel *mecA* homologue in human and bovine populations in the UK and Denmark: a descriptive study. *Lancet. Infect. Dis.* 11: 595-603.
- Goering, R. V., L. K. McDougal, G. E. Fosheim, K. K. Bonnstetter, D. J. Wolter, and F. C. Tenover. 2007. Epidemiologic distribution of the arginine catabolic mobile element among selected methicillin-resistant and methicillin-susceptible *Staphylococcus aureus* isolates. *J. Clin. Microbiol.* 45: 1981-1984.
- Gu, J., H. Li, M. Li, C. Vuong, M. Otto, Y. Wen, and Q. Gao. 2005. Bacterial insertion sequence IS256 as a potential molecular marker to discriminate invasive strains from commensal strains of *Staphylococcus epidermidis*. *J. Hosp. Infect.* 61: 342-348.
- Ito, T., K. Hiramatsu, A. Tomasz, H. de Lencastre, V. Perreten, M. T. Holden, D. C. Coleman, R. Goering, P. M. Giffard, R. L. Skov, K. Zhang, H. Westh, F. O'Brien, F. C. Tenover, D. C. Oliveira, S. Boyle-Vavra, F. Laurent, A. M. Kearns, B. Kreiswirth, K. S. Ko, H. Grundmann, J. E. Sollid, J. F. John, Jr., R. Daum, B. Soderquist, and G. Buist. 2012. Guidelines for Reporting Novel *mecA* Gene Homologues. *Antimicrob. Agents Chemother.*
- Kondo, Y., T. Ito, X. X. Ma, S. Watanabe, B. N. Kreiswirth, J. Etienne, and K. Hiramatsu. 2007. Combination of multiplex PCRs for Staphylococcal Cassette Chromosome *mec* type assignment: rapid identification system for *mec*, *ccr*, and major differences in junkyard regions. *Antimicrob. Agents Chemother.* 51: 264-274.
- Kuhnert, P., S. Capaul, J. Nicolet, and J. Frey. 1996. Phylogenetic positions of *Clostridium chauvoei* and *Clostridium septicum* based on 16S rRNA gene sequences. *Int. J. Syst. Bacteriol.* 46: 1174-1176.
- Loonen, A. J., A. R. Jansz, J. N. Bergland, M. Valkenburg, P. F. Wolffs, and A. J. van den Brule. 2012. Comparative study using phenotypic, genotypic, and proteomics methods for identification of coagulase-negative *staphylococci*. *J. Clin. Microbiol.* 50: 1437-1439.
- Lüthje, P. and S. Schwarz. 2006. Antimicrobial resistance of coagulase-negative staphylococci from bovine subclinical mastitis with particular reference to macrolide-lincosamide resistance phenotypes and genotypes. *J. Antimicrob. Chemother.* 57: 966-969.
- Miragaia, M., H. de Lencastre, F. Perdreau-Remington, H. F. Chambers, J. Higashi, P. M. Sullam, J. Lin, K. I. Wong, K. A. King, M. Otto, G. F. Sensabaugh, and B. A. Diep. 2009. Genetic diversity of arginine catabolic mobile element in *Staphylococcus epidermidis*. *PLoS ONE* 4: e7722.
- Perreten, V., L. Vorlet-Fawer, P. Slickers, R. Ehricht, P. Kuhnert, and J. Frey. 2005. Microarray-based detection of 90 antibiotic resistance genes of gram-positive bacteria. *J. Clin. Microbiol.* 43: 2291-2302.
- Piessens, V., S. De Vlieghe, B. Verbist, G. Braem, A. Van Nuffel, L. De Vuyst, M. Heyndrickx, and E. Van Coillie. 2012. Intra-species diversity and epidemiology varies among coagulase-negative *Staphylococcus species* causing bovine intramammary infections. *Vet. Microbiol.* 155: 62-71.

- Piessens, V., E. Van Coillie, B. Verbist, K. Supre, G. Braem, A. Van Nuffel, L. De Vuyst, M. Heyndrickx, and S. De Vliegher. 2011. Distribution of coagulase-negative *Staphylococcus* species from milk and environment of dairy cows differs between herds. *J. Dairy Sci.* 94: 2933-2944.
- Pyörälä, S. and S. Taponen. 2009. Coagulase-negative staphylococci-emerging mastitis pathogens. *Vet. Microbiol.* 134: 3-8.
- Rajala-Schultz, P. J., A. H. Torres, F. J. Degraes, W. A. Gebreyes, and P. Patchanee. 2009. Antimicrobial resistance and genotypic characterization of coagulase-negative staphylococci over the dry period. *Vet. Microbiol.* 134: 55-64.
- Sampimon, O. C., T. J. Lam, D. J. Mevius, Y. H. Schukken, and R. N. Zadoks. 2011. Antimicrobial susceptibility of coagulase-negative staphylococci isolated from bovine milk samples. *Vet. Microbiol.* 150: 173-179.
- Sawant, A. A., B. E. Gillespie, and S. P. Oliver. 2009. Antimicrobial susceptibility of coagulase-negative *Staphylococcus* species isolated from bovine milk. *Vet. Microbiol.* 134: 73-81.
- Schnellmann, C., V. Gerber, A. Rossano, V. Jaquier, Y. Panchaud, M. G. Doherr, A. Thomann, R. Straub, and V. Perreten. 2006. Presence of new *mecA* and *mph(C)* variants conferring antibiotic resistance in *Staphylococcus* spp. isolated from the skin of horses before and after clinic admission. *J. Clin. Microbiol.* 44: 4444-4454.
- Supré, K., F. Haesebrouck, R. N. Zadoks, M. Vaneechoutte, S. Piepers, and S. De Vliegher. 2011. Some coagulase-negative *Staphylococcus* species affect udder health more than others. *J. Dairy Sci.* 94: 2329-2340.
- Tenover, F. C., R. D. Arbeit, R. V. Goering, P. A. Mickelsen, B. E. Murray, D. H. Persing, and B. Swaminathan. 1995. Interpreting chromosomal DNA restriction patterns produced by pulsed-field gel electrophoresis: criteria for bacterial strain typing. *J. Clin. Microbiol.* 33: 2233-2239.
- Thomas, J. C., M. R. Vargas, M. Miragaia, S. J. Peacock, G. L. Archer, and M. C. Enright. 2007. Improved multilocus sequence typing scheme for *Staphylococcus epidermidis*. *J. Clin. Microbiol.* 45: 616-619.
- Tsubakishita, S., K. Kuwahara-Arai, T. Sasaki, and K. Hiramatsu. 2010. Origin and molecular evolution of the determinant of methicillin resistance in staphylococci. *Antimicrob. Agents. Chemother.* 54: 4352-4359.
- Waller, K. P., A. Aspan, A. Nyman, Y. Persson, and U. G. Andersson. 2011. CNS species and antimicrobial resistance in clinical and subclinical bovine mastitis. *Vet. Microbiol.* 152: 112-116.
- Walther, C. and V. Perreten. 2007. Methicillin-resistant *Staphylococcus epidermidis* in organic milk production. *J. Dairy Sci.* 90: 5351.
- Wielders, C. L., M. R. Vriens, S. Brisse, L. A. Graaf-Miltenburg, A. Troelstra, A. Fleer, F. J. Schmitz, J. Verhoef, and A. C. Fluit. 2001. In-vivo transfer of *mecA* DNA to *Staphylococcus aureus* [corrected]. *Lancet* 357: 1674-1675.

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- Woodford, N. 2005. Biological counterstrike: antibiotic resistance mechanisms of Gram-positive cocci. *Clin. Microbiol. Infect.* 11: 2-21.
- Wu, S., H. de Lencastre, and A. Tomasz. 1998. Genetic organization of the *mecA* region in methicillin-susceptible and methicillin-resistant strains of *Staphylococcus sciuri*. *J. Bacteriol.* 180: 236-242.
- Wu, S. W., H. de Lencastre, and A. Tomasz. 2001. Recruitment of the *mecA* gene homologue of *Staphylococcus sciuri* into a resistance determinant and expression of the resistant phenotype in *Staphylococcus aureus*. *J. Bacteriol.* 183: 2417-2424.
- Wu, S. W., H. D. Lencastre, and A. Tomasz. 2005. Expression of high-level methicillin resistance in *Staphylococcus aureus* from the *Staphylococcus sciuri mecA* homologue: role of mutation(s) in the genetic background and in the coding region of *mecA*. *Microb. Drug. Resist.* 11: 215-224.

8.) TABLES AND FIGURE

Table 1. Primer list

Target genes	Primer names and oligonucleotides	Size of PCR fragment	Annealing -temp.
<i>mecA</i> , <i>mecA1</i> , <i>mecA2</i>	mecAu-F: 5'-AAAAGATAAATCTTGGGGTG mecAu-R: 5'-CCTTGTTTCATYTTGAGTTC	525 bp	51°C
<i>mecA</i>	mecA-1: 5'-AAAATCGATGGTAAAGGTTGGC mecA-2: 5'-AGTTCTGCAGTACCGGATTTGC	533 bp	54°C
<i>mecA1</i>	mecA1-sc-F: 5'-ATTAATCATCGCCATCGTGA mecA1-sc-R: 5'-TTTGTATCTTGATTCATATTTGAACA	663 bp	52°C
<i>mecC</i>	mecA-LA251-F: 5'-CAGCCAGATTCATTTGTACC mecA-LA251-R: 5'-AACATCGTACGATGGGGTAC	486 bp	54°C
<i>mecA1</i> Promoter	mecAscK1-F: 5'-CATATATATATTTATACGCTCATC mecAsc-R: 5'-TTCAATGGCATCAATTGTTTC	335 bp	50°C
<i>mecA</i> ^{a)} (full-length gene)	mecA-F7: 5'-GATAACACCTGCTACAC mecA-R7: 5'-AAGGGAGAAGTAACAGC	2194 bp	51°C

a) Primers annealing external to *mecA* for amplification and sequencing of the full-length gene.

Table 2. Distribution of antimicrobial resistance phenotypes in CNS

Antimicrobial substance	Abr.	Breakpoints (µg/ml)	Resistance phenotypes							
			Total		clinical mastitis		subclinical mastitis		control milk	
			(n _{total} = 417)		(n _{total} = 115)		(n _{total} = 255)		(n _{total} = 47)	
			No.	%	No.	%	No.	%	No.	%
Oxacillin	OXA	R > 0.25	196	47.0	65 ^a	56.5	112 ^a	43.9	19	40.4
Fusidic acid	FUS	R > 1.0	141	33.8	44	38.3	81	31.8	16	34.0
Tiamulin	TIA	R > 2.0	133	31.9	34	29.6	85	33.3	14	29.8
Penicillin	PEN	R > 0.125	97	23.3	26	22.6	63	24.7	8	17.0
Tetracycline	TET	R > 2.0	66	15.8	18	15.7	37	14.5	11	23.4
Streptomycin	STR	R > 16	40	9.6	9	7.8	29	11.4	2	4.3
Erythromycin	ERY	R > 2.0	29	7.0	7	6.1	18	7.1	4	8.5
Sulfamethoxazole	SMX	R > 128	21	5.0	13 ^b	11.3	8	3.1	0 ^b	0.0
Trimethoprim	TMP	R > 4.0	18	4.3	6	5.2	11	4.3	1	2.1
Clindamycin	CLI	R > 0.5	14	3.4	3	2.6	10	3.9	1	2.1
Chloramphenicol	CHL	R > 8.0	13	3.1	4	3.5	8	3.1	1	2.1
Gentamicin	GEN	R > 1.0	10	2.4	5	4.4	5	2.0	0	0.0
Kanamycin	KAN	R > 8.0	10	2.4	5	4.4	5	2.0	0	0.0
Quinupristin-Dalfopristin	Q-D	R > 2.0	0	0.0	0	0.0	0	0.0	0	0.0
Rifampicin	RIF	R > 0.5	0	0.0	0	0.0	0	0.0	0	0.0
Ciprofloxacin	CIP	R > 1.0	0	0.0	0	0.0	0	0.0	0	0.0
Mupirocin	MUP	R > 256	0	0.0	0	0.0	0	0.0	0	0.0

a) Denotes a significant difference ($P = 0.0324$) in the number of oxacillin-resistant isolates from clinical and subclinical mastitis cases, as determined by Fisher's exact test.

b) Denotes a significant difference ($P=0.0208$) in the number of sulfamethoxazole-resistant isolates from clinical mastitis cases and control milk, as determined by Fisher's exact test.

Table 3. Incidence of coagulase-negative staphylococci (CNS) and distribution of the different CNS strains in clinical and subclinical mastitis milk and control milk¹.

Coagulase-negative staphylococci	Total		clinical mastitis milk		subclinical mastitis milk		control milk ¹	
	No.	%	No.	%	No.	%	No.	%
Total strains	417	100.0	115	100.0	255	100.0	47	100.0
<i>S. xylosus</i>	150	36.0	43	37.4	95	37.2	12	25.5
<i>S. chromogenes</i>	70	16.8	20	17.4	40	15.7	10	21.3
<i>S. sciuri</i>	37	8.9	10	8.7	25	9.8	2	4.3
<i>S. haemolyticus</i>	35	8.4	8	6.9	23	9.0	4	8.5
<i>S. devriesei</i>	18	4.3	7	6.0	11	4.3	0	0.0
<i>S. warneri</i>	17	4.1	4	3.5	6	2.3	7	14.8
<i>S. simulans</i>	16	3.8	1	0.9	13	5.1	2	4.3
<i>S. epidermidis</i>	15	3.6	4	3.5	10	3.9	1	2.1
<i>S. fleurettii</i>	12	2.9	6	5.2	4	1.6	2	4.3
<i>S. succinus</i>	9	2.2	0	0.0	7	2.8	2	4.3
<i>S. vitulinus</i>	9	2.2	5	4.4	4	1.6	0	0.0
<i>S. hyicus</i>	6	1.4	2	1.7	4	1.6	0	0.0
<i>S. equorum</i>	6	1.4	1	0.9	2	0.8	3	6.3
<i>S. saprophyticus</i>	2	0.5	1	0.9	1	0.4	0	0.0
<i>S. auricularis</i>	2	0.5	1	0.9	1	0.4	0	0.0
<i>S. capitis</i>	2	0.5	0	0.0	2	0.8	0	0.0
<i>S. cohnii</i>	2	0.5	0	0.0	0	0.0	2	4.3
<i>S. hominis</i>	1	0.2	0	0.0	1	0.4	0	0.0
<i>S. lentus</i>	1	0.2	0	0.0	1	0.4	0	0.0
<i>Staphylococcus sp.</i>	7	1.6	2	1.7	5	1.9	0	0.0

¹ control milk; milk taken from cows after mastitis treatment.

Table 4. Distribution of antimicrobial resistance and antimicrobial resistance genes in CNS from bovine milk

Antimicrobial substances	Percentage of phenotypical resistance (%)	Resistance genes	Proportion of resistance genes in resistant isolates		Number of strains (n)
			No.	%	
Oxacillin ^a	47.0	<i>mecA</i>	19	9.7	<i>S. fleurettii</i> (11), <i>S. epidermidis</i> (6), <i>S. haemolyticus</i> (1), <i>S. xylosus</i> (1)
		<i>mecAI</i>	37	18.9	<i>S. sciuri</i> (37)
		unknown	140	71.4	<i>S. xylosus</i> (102), <i>S. chromogenes</i> (11), <i>S. vitulinus</i> (8), <i>S. succinus</i> (5), <i>S. warneri</i> (3), <i>S. cohnii</i> (2), <i>S. saprophyticus</i> (2), <i>S. devriesei</i> (1), <i>S. equorum</i> (1), <i>S. hyicus</i> (1), <i>S. lentus</i> (1), <i>S. simulans</i> (1), <i>Staphylococcus</i> sp. (2)
Penicillin ^b	23.3	<i>blaZ</i>	88	90.7	<i>S. chromogenes</i> (37), <i>S. devriesei</i> (11), <i>S. haemolyticus</i> (11), <i>S. xylosus</i> (9), <i>S. epidermidis</i> (9), <i>S. warneri</i> (3), <i>S. cohnii</i> (2), <i>S. saprophyticus</i> (2), <i>S. auricularis</i> (1), <i>S. capitis</i> (1), <i>S. fleurettii</i> (1), <i>S. hominis</i> (1)
		no β-lactamase ^c	8	8.3	<i>S. fleurettii</i> (8)
		unknown	1	1.0	<i>S. xylosus</i> (1) ^d
Tetracycline	15.8	<i>tet(K)</i>	62	95.4	<i>S. xylosus</i> (33), <i>S. warneri</i> (7), <i>S. epidermidis</i> (6), <i>S. sciuri</i> (4), <i>S. chromogenes</i> (3), <i>S. simulans</i> (3), <i>S. fleurettii</i> (2), <i>S. vitulinus</i> (2), <i>S. haemolyticus</i> (1), <i>Staphylococcus</i> sp. (1)
		<i>tet(L)</i>	3	4.6	<i>S. chromogenes</i> (3)
Streptomycin	9.6	<i>str</i>	36	90.0	<i>S. chromogenes</i> (12), <i>S. epidermidis</i> (7), <i>S. sciuri</i> (5), <i>S. haemolyticus</i> (4), <i>S. devriesei</i> (3), <i>S. warneri</i> (2), <i>S. simulans</i> (1), <i>S. vitulinus</i> (1), <i>S. fleurettii</i> (1)
		<i>ant(6)-Ia</i>	3	7.5	<i>S. epidermidis</i> (2), <i>S. haemolyticus</i> (1)
		unknown	1	2.5	<i>S. haemolyticus</i> (1)
Erythromycin	7.0	<i>erm(B)</i>	2	7.4	<i>S. chromogenes</i> (1), <i>S. fleurettii</i> (1)
		<i>erm(C)</i>	11	40.8	<i>S. epidermidis</i> (6), <i>S. haemolyticus</i> (4), <i>S. warneri</i> (1)
		<i>msr</i>	6	22.2	<i>S. xylosus</i> (3), <i>S. epidermidis</i> (2), <i>S. hominis</i> (1)
Clindamycin	3.4	unknown	8	29.6	<i>S. equorum</i> (3), <i>S. xylosus</i> (1), <i>S. cohnii</i> (1), <i>S. fleurettii</i> (1), <i>Staphylococcus</i> sp. (2)
		<i>erm(B)</i>	2	14.3	<i>S. chromogenes</i> (1), <i>S. sciuri</i> (1)
		<i>erm(C)</i>	8	57.2	<i>S. haemolyticus</i> (4), <i>S. epidermidis</i> (4)
		<i>lnu(A)</i>	1	7.1	<i>S. xylosus</i> (1)
		unknown	3	21.4	<i>S. xylosus</i> (1), <i>S. fleurettii</i> (1), <i>S. lentus</i> (1)
Chloramphenicol	3.1	<i>cat_{pC221}</i>	7	53.8	<i>S. epidermidis</i> (3), <i>S. xylosus</i> (2), <i>S. chromogenes</i> (1), <i>S. sciuri</i> (1)
		<i>cat_{pC223}</i>	4	30.8	<i>S. cohnii</i> (1), <i>S. haemolyticus</i> (1), <i>S. simulans</i> (1), <i>S. xylosus</i> (1)
		unknown	2	15.4	<i>S. haemolyticus</i> (1), <i>S. simulans</i> (1)
Kanamycin	2.4	<i>aac(6')-Ie – aph(2')-Ia</i>	7	70.0	<i>S. epidermidis</i> (4), <i>S. sciuri</i> (1), <i>S. chromogenes</i> (1), <i>S. fleurettii</i> (1)
		<i>aph(3')-III</i>	3	30.0	<i>S. haemolyticus</i> (2), <i>S. epidermidis</i> (1)
Gentamicin	2.4	<i>aac(6')-Ie – aph(2')-Ia</i>	7	70.0	<i>S. epidermidis</i> (4), <i>S. sciuri</i> (1), <i>S. chromogenes</i> (1), <i>S. fleurettii</i> (1)
		unknown	3	30.0	<i>S. haemolyticus</i> (2), <i>S. xylosus</i> (1)
Trimethoprim	1.2	<i>dfr(A)</i>	1	20.0	<i>S. epidermidis</i> (1)
		<i>dfr(D)</i>	2	40.0	<i>S. sciuri</i> (1), <i>S. fleurettii</i> (1)
		<i>dfr(G)</i>	1	20.0	<i>S. vitulinus</i> (1)
		<i>dfr(K)</i>	1	20.0	<i>S. chromogenes</i> (1)

Legend Table 4.

Antimicrobial resistance genes and their functions:

aac(6')-Ie – *aph(2')-Ia*, gentamicin, kanamycin and neomycin acetyltransferase; *aph(3')-III*, kanamycin and neomycin phosphotransferase; *ant(6)-Ia*, streptomycin adenylyltransferase; *blaZ*, β -lactamase; *cat_{pC221}* and *cat_{pC223}*, chloramphenicol acetyltransferases; *dfr(A)*, *dfr(D)*, *dfr(G)*, *dfr(K)*, trimethoprim-resistant dihydrofolate reductases; *erm(B)* and *erm(C)*, macrolide, lincosamide and streptogramin B 23S rRNA methylase; *lnu(A)*, lincosamide nucleotidyltransferase; *mecA* and *mecA1*, penicillin-binding proteins; *str*, streptomycin nucleotidyltransferase; and *tet(K)* and *tet(L)*, tetracycline efflux proteins.

- a) Oxacillin, indicator antimicrobial for the presence of an alternative PBP 2a encoded by *mec* genes.
- b) Penicillin used for the prediction of a β -lactamase.
- c) Do not produce β -lactamase, but contained *mecA*.
- d) β -lactamase-positive.

Table 5. Occurrence of multiple antimicrobial resistance genes in CNS isolated from the milk of 20 different cows suffering from bovine mastitis.

Isolate	CNS species	Mastitis	Resistances
<i>mecA</i> positive:			
M1529/10	<i>S. epidermidis</i>	subclinical	<i>erm</i> (C): CLI, ERY; <i>tet</i> (K): TET; <i>str</i> : STR; <i>bla</i> Z: PEN; <i>aac</i> (6')-Ie – <i>aph</i> (2')-Ia: GEN-KAN; <i>dfr</i> (A): TMP; <i>mecA</i> : OXA
M1186/10	<i>S. epidermidis</i>	clinical	<i>erm</i> (C): CLI, ERY; <i>tet</i> (K): TET; <i>ant</i> (6)-Ia: STR; <i>bla</i> Z: PEN; <i>aac</i> (6')-Ie – <i>aph</i> (2')-Ia: GEN-KAN; <i>mecA</i> : OXA
M4460/09	<i>S. fleuretii</i>	clinical	<i>erm</i> (B): CLI, ERY; <i>str</i> : STR; <i>bla</i> Z: PEN; <i>aac</i> (6')-Ie – <i>aph</i> (2')-Ia: GEN-KAN; <i>dfr</i> (D): TMP; <i>mecA</i> : OXA
M1570/10	<i>S. haemolyticus</i>	clinical	<i>erm</i> (C): CLI, ERY; <i>tet</i> (K): TET; <i>bla</i> Z: PEN; <i>aph</i> (3')-III: KAN; <i>mecA</i> : OXA
M744/10	<i>S. epidermidis</i>	subclinical	<i>str</i> : STR; <i>cat</i> _{pC221} : CHL; <i>aac</i> (6')-Ie – <i>aph</i> (2')-Ia: GEN-KAN; <i>erm</i> (C): CLI, ERY; <i>mecA</i> : OXA
M1383/10	<i>S. epidermidis</i>	subclinical	<i>erm</i> (C): CLI, ERY; <i>bla</i> Z: PEN; <i>aac</i> (6')-Ie – <i>aph</i> (2')-Ia: GEN-KAN; <i>mecA</i> : OXA
M1965/10	<i>S. sciuri</i>	clinical	<i>tet</i> (K): TET; <i>str</i> : STR; <i>aac</i> (6')-Ie – <i>aph</i> (2')-Ia: GEN-KAN; <i>dfr</i> (D): TMP; <i>mecA</i> : OXA
M8/10	<i>S. epidermidis</i>	clinical	<i>ant</i> (6)-Ia: STR; <i>bla</i> Z: PEN; <i>cat</i> _{pC221} : CHL; <i>aph</i> (3')-III: KAN; <i>erm</i> (C): CLI, ERY; <i>mecA</i> : OXA
M1201/10	<i>S. sciuri</i>	subclinical	<i>tet</i> (K): TET; <i>str</i> : STR; <i>cat</i> _{pC221} : CHL; <i>mecA</i> : OXA
M703/10	<i>S. epidermidis</i>	subclinical	<i>bla</i> Z: PEN; <i>msr</i> : ERY; <i>mecA</i> : OXA
M3901/09	<i>S. sciuri</i>	clinical	<i>tet</i> (K): TET; <i>str</i> : STR; <i>mecA</i> : OXA
<i>mecA</i> negative:			
M425/10	<i>S. chromogenes</i>	subclinical	<i>erm</i> (B): CLI, ERY; <i>tet</i> (L): TET; <i>bla</i> Z: PEN; <i>cat</i> _{pC221} : CHL; <i>dfr</i> (K): TMP
M47/10	<i>S. chromogenes</i>	subclinical	<i>tet</i> (L): TET; <i>str</i> : STR; <i>bla</i> Z: PEN; <i>aac</i> (6')-Ie – <i>aph</i> (2')-Ia: GEN-KAN
M1256/10	<i>S. warneri</i>	subclinical	<i>str</i> : STR; <i>bla</i> Z: PEN; <i>erm</i> (C): CLI, ERY
M4233-1/09	<i>S. epidermidis</i>	subclinical	<i>erm</i> (C): CLI-ERY; <i>tet</i> (K): TET; <i>str</i> : STR
M4298-1/09	<i>S. epidermidis</i>	subclinical	<i>tet</i> (K): TET; <i>str</i> : STR; <i>bla</i> Z: PEN; <i>cat</i> _{pC221} : CHL
M46/10	<i>S. chromogenes</i>	subclinical	<i>tet</i> (K): TET; <i>str</i> : STR; <i>bla</i> Z: PEN
M523/10	<i>S. epidermidis</i>	clinical	<i>tet</i> (K): TET; <i>str</i> : STR; <i>bla</i> Z: PEN
M619-2/10	<i>S. haemolyticus</i>	subclinical	<i>ant</i> (6)-Ia: STR; <i>bla</i> Z: PEN; <i>aph</i> (3')-III: KAN
M1094-1/10	<i>S. warneri</i>	subclinical	<i>tet</i> (K): TET; <i>str</i> : STR; <i>bla</i> Z: PEN

Antimicrobial resistance genes and their functions: *aac*(6')-Ie – *aph*(2')-Ia, gentamicin, kanamycin and neomycin acetyltransferase; *aph*(3')-III, kanamycin phosphotransferase; *ant*(6)-Ia, streptomycin adenylyltransferase; *bla*Z, β -lactamase; *cat*_{pC221} and *cat*_{pC223}, chloramphenicol acetyltransferases; *dfr*(A), *dfr*(D) and *dfr*(K), trimethoprim-resistant dihydrofolate reductases; *erm*(B) and *erm*(C), macrolide, lincosamide and streptogramin B 23S rRNA methylase; *msr*, macrolide efflux protein; *mecA* and *mecA*I, penicillin-binding proteins; *str*, streptomycin nucleotidyltransferase; and *tet*(K) and *tet*(L), tetracycline efflux proteins.

CLI, clindamycin; TET, tetracycline; STR, streptomycin; PEN, penicillin; KAN, kanamycin; GEN, gentamicin; TMP, trimethoprim; ERY, erythromycin; OXA, oxacillin; CHL, chloramphenicol

Figure 1: PFGE profile, SCCmec, ST and ACME types of 19 methicillin-resistant coagulase-negative staphylococci

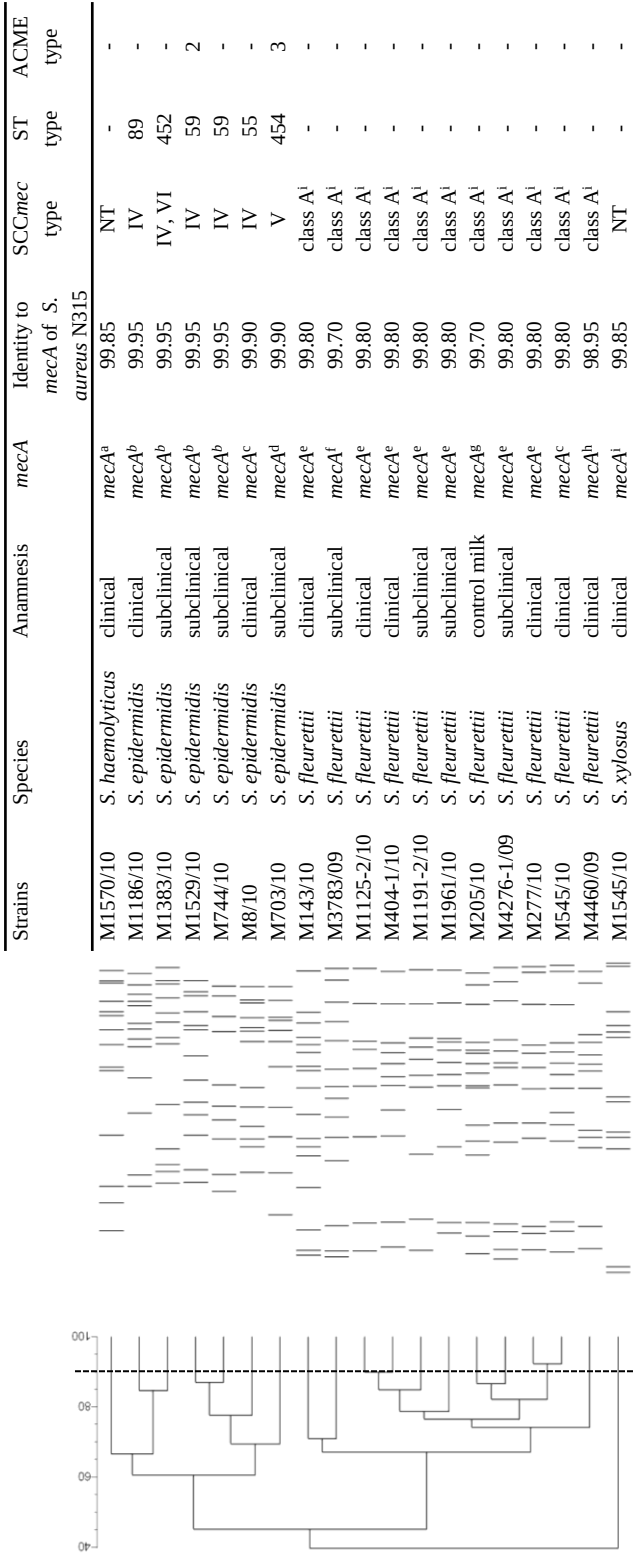


Figure 1. Genetic background and properties of methicillin-resistant coagulase-negative staphylococci (CNS) from bovine mastitis milk from Switzerland. The phylogenetic tree was constructed from pulsed-field gel electrophoresis (PFGE) patterns of 19 methicillin-resistant CNS isolates. Cluster analysis was generated by Bionumerics 6.6; Applied Maths, Kortijk, Belgium. Comparison settings: Dice, UPGMA, optimization 1.5%, position tolerance 1.5%. The dotted line indicates the cut-off value of $\geq 90\%$ determining clonality between the isolates, according to the criteria of Tenover et al. 1995.

- a) *mecA* of *S. haemolyticus* M1570/10 (EMBL Accession no HE978799)
- b) *mecA* identical to *mecA* of *S. epidermidis* RP26A (EMBL Accession no CP000029.1)
- c) *mecA* of *S. epidermidis* M8/10 (EMBL Accession no HE978797)
- d) *mecA* of *S. epidermidis* M703/10 (EMBL Accession no HE978798)
- e) *mecA* identical to *mecA* of *S. fleurettii* M143/10 (EMBL Accession no HE978795)
- f) *mecA* of *S. fleurettii* M3783/09 (EMBL Accession no HE978796)
- g) *mecA* of *S. fleurettii* M205/10 (EMBL Accession no HE978794)
- h) *mecA* of *S. fleurettii* M4460/09 (EMBL Accession no HE861945)
- i) *mecA* of *S. xylosus* M1545/10 (EMBL Accession no HE978800)
- j) class A; no *ccr* gene detected

NT, not typeable.

Antibiotic resistance in coagulase–negative staphylococci from bovine mastitis

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Antibiotic resistance in coagulase–negative staphylococci from bovine mastitis

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Coagulase–negative staphylococci (CoNS) are regarded as important opportunistic mastitis pathogens in cows which may persist after therapy. The aim of the project is to investigate which CoNS strains are present in bovine mastitis milk and if methicillin-resistant CoNS play a role in bovine mastitis.

448 CoNS were isolated from milk of cows suffering from subclinical or clinical mastitis. The isolates were identified at species level using MALDI-TOF. Species with unclear identification were submitted to Vitek 2 and DNA sequence analysis of 16S rDNA and hsp60 marker. Antibiotic susceptibility was tested by broth dilution method. MICs were interpreted using resistance breakpoints from EUCAST. The antibiotic resistance genes were detected using a microarray.

The 448 CoNS isolates were identified as *S. xylosus* (39.2%), *S. chromogenes* (16.3%), *S. sciuri* (8.9%), *S. haemolyticus* (8.5%), *S. devriesii* (4.5%), *S. warneri* (4.0%), *S. simulans* (3.8%), *S. epidermidis* (3.4%), *S. fleuretti* (2.2%), *S. succinus* (2.2%), *S. vitulinus* (2.0%) and 8 other species (each <1%). The most frequent resistances were those to oxacillin (47.1%), fusidic acid (34.2%), tiamulin (33.3%), penicillin (22.8%), tetracycline (15.4%), streptomycin (9.2%), erythromycin (7.4%), kanamycin (2.5%) and gentamicin (2.5%). Resistance to oxacillin was attributed to the presence of the *mecA* gene in 32.2% of the oxacillin-resistant isolates. The mechanism of resistance for the remaining oxacillin-resistant isolates remained unknown. The *mecA* gene was detected in *S. xylosus*, *S. fleuretti*, *S. epidermidis* and *S. sciuri*. Resistance to tetracycline was attributed to *tet(K)* (96%) and *tet(L)* (4%), penicillin resistance to *blaZ* (90%) and/or *mecA* (32.2%), streptomycin resistance to *str* (90%) and *ant(6)-Ia* (10%), erythromycin resistance to *erm(C)* (36.4%), *erm(B)* (6.0%), *msr* (21.2%), *mph(C)* (18.2%) and unknown genes (12.1%), whereas resistance to tiamulin and fusidic acid could not be attributed to an acquired resistance gene.

Methicillin-resistant CoNS are present in bovine mastitis milk in Switzerland. These isolates are resistant to all beta-lactam antibiotics (including cephalosporins) which are widely used in

the treatment of mastitis. The strains were also resistant, but in a lower proportion, to other classes of drugs (macrolides, aminoglycosides) that are also used for the treatment of mastitis. Exact identification of the staphylococcus species followed by an antibiogram is recommended prior therapy.

Acknowledgements

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Antibiotic resistance in coagulase-negative staphylococci from bovine mastitis

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Introduction

Coagulase-negative staphylococci (CNS) are regarded as important opportunistic mastitis pathogens in cows which cause an increase of the somatic cell count (SCC) and light intramammary infections (IMI). After therapy CNS may persist because of the decreased susceptibility to antibiotics. The aim of the project was to investigate which CNS strains are present in bovine mastitis milk and if methicillin-resistant CNS as well as other multidrug-resistant CNS are associated with bovine mastitis.

Table 1: Antimicrobial resistance profile of CNS from bovine mastitis

Antimicrobial substances	Phenotypical Resistance ¹ (%)	Resistance genes	Resistance		Number of strains
			No.	%	
Oxacillin	47.0	<i>mecA</i>	18	9.2	<i>S. fleurettii</i> (11), <i>S. epidermidis</i> (6), <i>S. haemolyticus</i> (1)
		<i>mecA1</i>	37	19.0	<i>S. sciuri</i> (37)
		unknown	140	71.8	<i>S. xylosus</i> (103), <i>S. chromogenes</i> (11), <i>S. vitulinus</i> (8), <i>S. succinus</i> (5), <i>S. warneri</i> (3), <i>S. cohnii</i> (2), <i>S. saprophyticus</i> (2), <i>S. devriesei</i> (1); <i>S. equorum</i> (1), <i>S. hyicus</i> (1), <i>S. lentus</i> (1), <i>S. sciuri</i> (1); <i>S. simulans</i> (1)
Penicillin	23.3	<i>blaZ</i>	81	83.5	<i>S. chromogenes</i> (36), <i>S. devriesei</i> (11), <i>S. haemolyticus</i> (10), <i>S. xylosus</i> (9), <i>S. epidermidis</i> (4), <i>S. warneri</i> (3), <i>S. cohnii</i> (2), <i>S. saprophyticus</i> (2), <i>S. fleurettii</i> (1), <i>S. auricularis</i> (1), <i>S. capitis</i> (1), <i>S. hominis</i> (1)
		<i>blaZ</i> , <i>mecA</i>	5	5.2	<i>S. epidermidis</i> (4), <i>S. haemolyticus</i> (1)
		<i>mecA</i>	9	9.3	<i>S. fleurettii</i> (8), <i>S. sciuri</i> (1)
		unknown	2	2.0	<i>S. epidermidis</i> (1), <i>S. xylosus</i> (1)
Tetracycline	14.9	<i>tet(K)</i>	62	95.4	<i>S. xylosus</i> (34), <i>S. warneri</i> (7), <i>S. epidermidis</i> (6), <i>S. sciuri</i> (4), <i>S. chromogenes</i> (3), <i>S. simulans</i> (3), <i>S. fleurettii</i> (2), <i>S. vitulinus</i> (2), <i>S. haemolyticus</i> (1)
		<i>tet(L)</i>	3	4.6	<i>S. chromogenes</i> (3)
		<i>str</i>	36	90.0	<i>S. chromogenes</i> (12), <i>S. epidermidis</i> (7), <i>S. sciuri</i> (5), <i>S. haemolyticus</i> (4), <i>S. devriesei</i> (3), <i>S. warneri</i> (2), <i>S. simulans</i> (1), <i>S. vitulinus</i> (1), <i>S. fleurettii</i> (1)
Streptomycin	9.6	<i>ant(6)-Ia</i>	3	7.5	<i>S. epidermidis</i> (2), <i>S. haemolyticus</i> (1)
		unknown	1	2.5	<i>S. haemolyticus</i> (1)
Erythromycin	6.7	<i>erm(B)</i>	2	7.4	<i>S. chromogenes</i> (1), <i>S. fleurettii</i> (1)
		<i>erm(C)</i>	11	40.8	<i>S. epidermidis</i> (6), <i>S. haemolyticus</i> (4), <i>S. warneri</i> (1)
		<i>msr</i>	6	22.2	<i>S. xylosus</i> (3), <i>S. epidermidis</i> (2), <i>S. hominis</i> (1)
		<i>mph(C)</i>	6	22.2	<i>S. equorum</i> (3), <i>S. xylosus</i> (3)
Clindamycin	3.4	unknown	2	7.4	<i>S. cohnii</i> (1), <i>S. fleurettii</i> (1)
		<i>erm(B)</i>	2	14.3	<i>S. chromogenes</i> (1), <i>S. sciuri</i> (1)
		<i>erm(C)</i>	8	57.1	<i>S. haemolyticus</i> (4), <i>S. epidermidis</i> (4)
		<i>lnu(A)</i>	1	7.1	<i>S. xylosus</i> (1)
Chloramphenicol	3.1	unknown	3	21.4	<i>S. xylosus</i> (1), <i>S. fleurettii</i> (1), <i>S. lentus</i> (1)
		<i>catpC221</i>	7	53.8	<i>S. epidermidis</i> (3), <i>S. xylosus</i> (2), <i>S. chromogenes</i> (1), <i>S. sciuri</i> (1)
		<i>catpC223</i>	4	30.8	<i>S. cohnii</i> (1), <i>S. haemolyticus</i> (1), <i>S. simulans</i> (1), <i>S. xylosus</i> (1)
		unknown	2	15.4	<i>S. haemolyticus</i> (1), <i>S. simulans</i> (1)
Kanamycin	2.4	<i>aac(6)-Ie-aph(2')-Ia</i>	7	70.0	<i>S. epidermidis</i> (4), <i>S. sciuri</i> (1), <i>S. chromogenes</i> (1), <i>S. fleurettii</i> (1)
		<i>aph(3')-III</i>	3	30.0	<i>S. haemolyticus</i> (2), <i>S. epidermidis</i> (1)
Gentamicin	2.4	<i>aac(6)-Ie-aph(2')-Ia</i>	7	70.0	<i>S. epidermidis</i> (4), <i>S. sciuri</i> (1), <i>S. chromogenes</i> (1), <i>S. fleurettii</i> (1)
		unknown	3	30.0	<i>S. haemolyticus</i> (2), <i>S. xylosus</i> (1)
Trimethoprim	1.4	<i>dfp(A)</i>	1	16.7	<i>S. epidermidis</i> (1)
		<i>dfp(D)</i>	2	33.3	<i>S. sciuri</i> (1), <i>S. fleurettii</i> (1)
		<i>dfp(G)</i>	1	16.7	<i>S. vitulinus</i> (1)
		<i>dfp(K)</i>	1	16.7	<i>S. chromogenes</i> (1)
		unknown	1	16.6	<i>S. xylosus</i> (1)

¹ MIC determining the resistance breakpoints were those from EUCAST (www.eucast.org).

Material and Methods

Sample collection: 417 CNS isolates from 363 different dairy cows in Switzerland.

Identification of the CNS: MALDI-TOF MS (Bruker), Vitek 2 (Biomérieux) and DNA sequencing analysis of 16S rDNA and *hsp60* marker.

Antibiotic susceptibility test: broth microdilution method [Sensititre susceptibility plates NLEUST (Trek diagnostics)], 19 different antimicrobial substances.

Antibiotic resistance genes detection: microarray AMR+ve-2 (Identibac, Alere Tech) and PCR.

Results I: Identification of CNS isolates

The 417 CNS isolates from bovine mastitis milk were identified as *S. xylosus* (37.2%), *S. chromogenes* (16.6%), *S. sciuri* (9.6%), *S. haemolyticus* (8.9%), *S. devriesei* (4.3%), *S. warneri* (4.1%), *S. simulans* (3.8%), *S. epidermidis* (3.6%), *S. fleuretti* (2.4%), *S. succinus* (2.2%), *S. vitulinus* (2.2%), and 8 other species (each <1%)

Fig.1: Microarray results of *S. epidermidis* strain M1186/10 from bovine mastitis

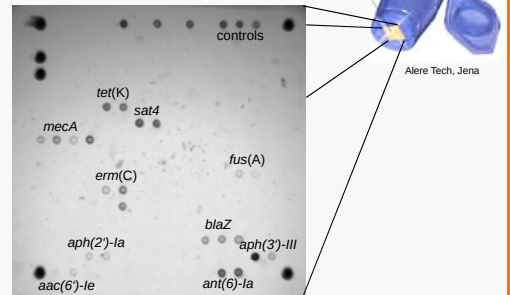


Table 2: Multiresistance profiles of CNS from bovine mastitis

Isolate	CNS species	Anamnesis	Resistances
mecA positive:			
M1529/10	<i>S. epidermidis</i>	subclinical	<i>erm(C)</i> : CLU, ERY; <i>tet(K)</i> : TET; <i>str</i> : STR; <i>blaZ</i> : PEN; <i>aac(6)-Ie-aph(2')-Ia</i> : GEN-KAN; <i>dfp(A)</i> : TMP; <i>mecA</i> : OXA
M1186/10	<i>S. epidermidis</i>	clinical	<i>erm(C)</i> : CLU, ERY; <i>tet(K)</i> : TET; <i>ant(6)-Ia</i> : STR; <i>blaZ</i> : PEN; <i>aac(6)-Ie-aph(2')-Ia</i> : GEN-KAN; <i>mecA</i> : OXA
M4460/10	<i>S. fleurettii</i>	clinical	<i>erm(B)</i> : CLU, ERY; <i>str</i> : STR; <i>blaZ</i> : PEN; <i>aac(6)-Ie-aph(2')-Ia</i> : GEN-KAN; <i>dfp(D)</i> : TMP; <i>mecA</i> : OXA
M1570/10	<i>S. haemolyticus</i>	clinical	<i>erm(C)</i> : CLU, ERY; <i>tet(K)</i> : TET; <i>blaZ</i> : PEN; <i>aph(3')-III</i> : KAN; <i>mecA</i> : OXA
M744/10	<i>S. epidermidis</i>	subclinical	<i>str</i> : STR; <i>catpC221</i> : CHL; <i>aac(6)-Ie-aph(2')-Ia</i> : GEN-KAN; <i>erm(C)</i> : ERY; <i>mecA</i> : OXA
M1383/10	<i>S. epidermidis</i>	subclinical	<i>erm(C)</i> : CLU, ERY; <i>blaZ</i> : PEN; <i>aac(6)-Ie-aph(2')-Ia</i> : GEN-KAN; <i>mecA</i> : OXA
M1965/10	<i>S. sciuri</i>	clinical	<i>tet(K)</i> : TET; <i>str</i> : STR; <i>aac(6)-Ie-aph(2')-Ia</i> : GEN-KAN; <i>dfp(D)</i> : TMP; <i>mecA1</i> : OXA
M8/10	<i>S. epidermidis</i>	clinical	<i>ant(6)-Ia</i> : STR; <i>catpC221</i> : CHL; <i>aph(3')-III</i> : KAN; <i>erm(C)</i> : ERY; <i>mecA</i> : OXA
M1201/10	<i>S. sciuri</i>	subclinical	<i>tet(K)</i> : TET; <i>str</i> : STR; <i>catpC221</i> : CHL; <i>mecA1</i> : OXA
M703/10	<i>S. epidermidis</i>	subclinical	<i>blaZ</i> : PEN; <i>msr</i> : ERY; <i>mecA</i> : OXA
M3901/09	<i>S. sciuri</i>	clinical	<i>tet(K)</i> : TET; <i>str</i> : STR; <i>mecA1</i> : OXA
M545/10	<i>S. fleurettii</i>	clinical	<i>tet(K)</i> : TET; <i>mecA</i> : OXA, PEN
mecA negative:			
M425/10	<i>S. chromogenes</i>	subclinical	<i>erm(B)</i> : CLU, ERY; <i>tet(L)</i> : TET; <i>blaZ</i> : PEN; <i>catpC221</i> : CHL; <i>dfp(K)</i> : TMP
M47/10	<i>S. chromogenes</i>	subclinical	<i>tet(L)</i> : TET; <i>str</i> : STR; <i>blaZ</i> : PEN; <i>aac(6)-Ie-aph(2')-Ia</i> : GEN-KAN
M1256/10	<i>S. warneri</i>	subclinical	<i>str</i> : STR; <i>blaZ</i> : PEN; <i>erm(C)</i> : ERY
M4233-1/09	<i>S. epidermidis</i>	subclinical	<i>erm(C)</i> : CLU, ERY; <i>tet(K)</i> : TET; <i>str</i> : STR
M4298-1/09	<i>S. epidermidis</i>	subclinical	<i>tet(K)</i> : TET; <i>str</i> : STR; <i>blaZ</i> : PEN; <i>catpC221</i> : CHL
M46/10	<i>S. chromogenes</i>	subclinical	<i>tet(K)</i> : TET; <i>str</i> : STR; <i>blaZ</i> : PEN
M523/10	<i>S. epidermidis</i>	clinical	<i>tet(K)</i> : TET; <i>str</i> : STR; <i>blaZ</i> : PEN
M619-2/10	<i>S. haemolyticus</i>	subclinical	<i>ant(6)-Ia</i> : STR; <i>blaZ</i> : PEN; <i>aph(3')-III</i> : KAN
M1094-1/10	<i>S. warneri</i>	subclinical	<i>tet(K)</i> : TET; <i>str</i> : STR; <i>blaZ</i> : PEN

Results II: Antimicrobial resistance in CNS

The distribution of antimicrobial resistance in CNS is shown in Table 1.

5% of the CNS exhibited resistance to three or more different antibiotics. Multiresistance was most frequently found in *S. epidermidis* strains (Table 2 and Figure 1).

Resistance to oxacillin was attributed to the presence of the *mecA* genes (*mecA*, *mecA1*) in 28.2% of the oxacillin-resistant isolates. The remaining 71.8% displayed resistance to oxacillin and did not contain *mecA*. For these isolates, the MIC of oxacillin was 0.5 µg/ml which is just above the EUCAST resistance breakpoint (> 0.25 µg/ml) (Figure 2).

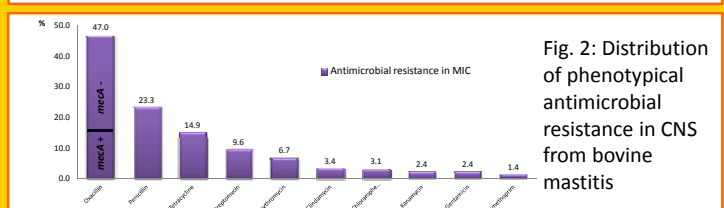


Fig. 2: Distribution of phenotypical antimicrobial resistance in CNS from bovine mastitis

Discussion and Conclusion

The clinical breakpoint of oxacillin may not be appropriate for CNS of bovine mastitis. Confirmation of the presence of *mecA* is necessary. Methicillin-resistant and multiresistant CNS are present in bovine mastitis milk in Switzerland and are resistant to antibiotics which are widely used in the treatment of mastitis. Exact identification of staphylococci and antibiogram is recommended prior therapy.

Genetic properties of multidrug-resistant *Staphylococcus epidermidis* isolates from bovine mastitis

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Poster presented at the International Symposium on Staphylococci and Staphylococcal Infections in Lyon, France, 26-30 August 2012

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Keywords: CNS, milk, methicillin-resistance, antibiotic, animal

Coagulase-negative staphylococci (CNS) have become the most commonly isolated microorganisms from bovine milk in many countries and are regarded as important mastitis pathogens. *S. epidermidis* is one of the CNS that frequently persists after antibiotic treatment. In this study, we determined whether specific multidrug-resistant *S. epidermidis* clones are associated with bovine mastitis.

The *S. epidermidis* isolates were identified using MALDI-TOF MS and genotyped by MLST. Antibiotic resistance profile was tested by broth dilution and microarray. The presence of the biofilm-formation operon *ica* and of the arginine catabolic mobile element ACME, a putative pathogenic island conferring growth and survival advantage of the bacteria in the hosts, were determined by PCR. Out of 363 cows suffering from mastitis, 13 were found to be infected with a multidrug-resistant *S. epidermidis*, including 6 methicillin-resistant *S. epidermidis* (MRSE). The MRSE displayed resistance to antibiotics like the beta-lactams (*mecA*; *blaZ*), macrolides and lincosamides [*erm*(C)], tetracycline [*tet*(K)], aminoglycosides streptomycin [*str* or *ant*(6)-*Ia*], and gentamicin and kanamycin [*aac*(6')-*Ie* – *aph*(2)-*Ia*; *aph*(3)-*III*], and trimethoprim [*dfr*(A)]. They belonged to CC59 (n=2), CC55, CC57, CC119, and CC291. The remaining multidrug-resistant *S. epidermidis* displayed resistance to penicillin (*blaZ*), macrolides (*msr*), macrolides and lincosamides [*erm*(C)], tetracycline [*tet*(K)], and streptomycin (*str*) and belonged to CC111 (n=4), CC184, CC57, and CC5. None of the isolates contained the *ica* operon. The presence of the ACME operon was mainly associated to CC111 in methicillin-susceptible, but multidrug-resistant *S. epidermidis*, and to CC59 and CC119 in MRSE.

The use of antibiotics in dairy farms has likely selected for multidrug-resistant *S. epidermidis*. The isolates are resistant to antibiotics which are widely used in the treatment of mastitis. They belonged to specific clonal lineages, with CC111 being the most common. The presence of the ACME in *S. epidermidis* may favor colonization of bovine udder and contribute to the spread of multidrug-resistant *S. epidermidis* in dairy cows.

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Introduction

Coagulase-negative staphylococci (CNS) have become the most commonly isolated microorganisms from bovine milk in many countries and are regarded as important mastitis pathogens. *S. epidermidis* is one of the CNS that frequently persists after antimicrobial treatment. In this study, we determined whether specific multidrug-resistant *S. epidermidis* clones are associated with bovine mastitis.

Material and Methods

The *S. epidermidis* isolates were identified using MALDI-TOF MS and genotyped by PFGE and MLST. Antibiotic resistance profile was tested by broth dilution method and microarray. The presence of the biofilm-formation operon *ica* and of the arginine catabolic mobile element (ACME), a putative pathogenic island conferring growth and survival advantage of the bacteria in the hosts, were determined by PCR.

Results

Out of 363 cows suffering from mastitis, 13 were found to be infected with a multidrug-resistant *S. epidermidis* strain, including 6 methicillin-resistant *S. epidermidis* (MRSE) strains.

Antimicrobial resistance genes: The MRSE (n = 6) displayed resistance to antibiotics like the beta-lactams (*mecA*; *blaZ*), the macrolides and the lincosamides [*erm*(C)], tetracycline [*tet*(K)], the aminoglycosides like streptomycin [*str* or *ant*(6)-*la*], gentamicin and kanamycin [*aac*(6')-*le* – *aph*(2)-*la*; *aph*(3)-*III*], and trimethoprim [*dfr*(A)]. The remaining multidrug-resistant *S. epidermidis* isolates displayed resistance to penicillin (*blaZ*), macrolides (*msr*), macrolides and lincosamides [*erm*(C)], tetracycline [*tet*(K)], and streptomycin (*str*) (Table 1).

Genotyping of methicillin-resistant isolates: Two different *mecA* genes whose sequences differed slightly from each other were detected in *S. epidermidis* isolates (Fig.1). MRSE contained SCCmec type IV (n=4), type V (n=1) and one non-typeable SCCmec related to type IV and VI (Fig.1).

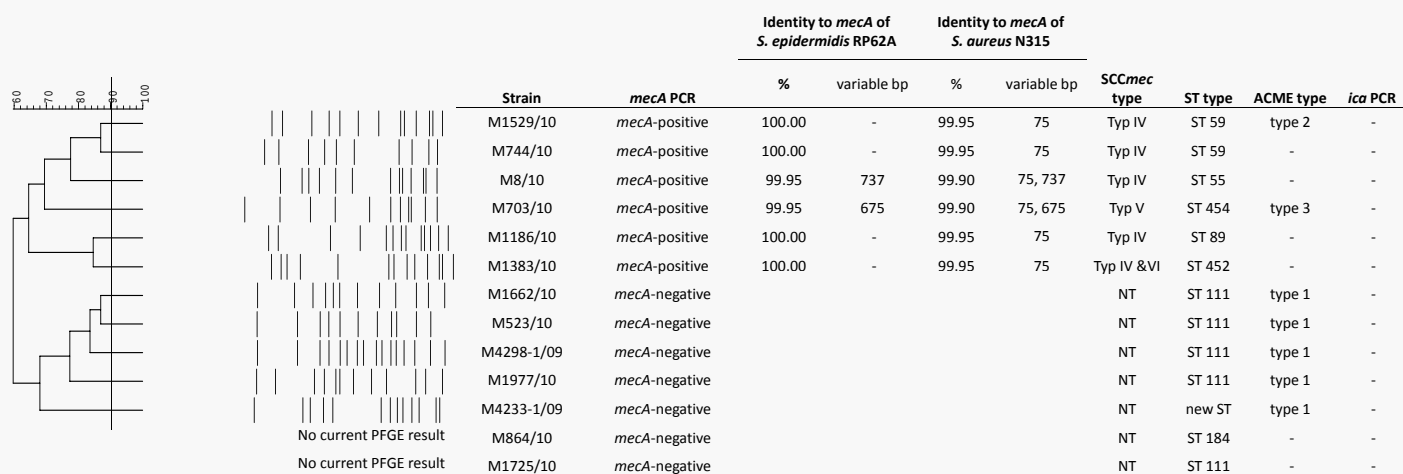
Clonality: MRSE belonged to ST59 (n = 2), ST55, ST89, and to two new ST types (ST452 and ST454), whereas MSSE belonged to ST111 (n=4), ST184, ST293, and a new ST. PFGE analysis of the *S. epidermidis* (n = 11) isolates showed that the strains were not clonally related. However, MRSE and MSSE clustered in two different PFGE branches (Fig. 1).

ACME and *ica* operons: The presence of the ACME operons was mainly associated to ST111 in methicillin-susceptible, but multidrug-resistant *S. epidermidis* isolates, and to ST59 and ST454 in MRSE. None of the *S. epidermidis* isolates contained the biofilm-formation operon *ica* (Fig.1).

Table 1: Antimicrobial resistance in *S. epidermidis* strains from clinical and subclinical mastitis

Strains	Mastitis	Resistances
M1529/10	subclinical	<i>erm</i> (C): CLI, ERY; <i>tet</i> (K): TET; <i>str</i> : STR; <i>blaZ</i> : PEN; <i>aac</i> (6')- <i>le</i> – <i>aph</i> (2')- <i>la</i> : GEN-KAN; <i>dfr</i> (A): TMP; <i>mecA</i> : OXA
M1186/10	clinical	<i>erm</i> (C): CLI, ERY; <i>tet</i> (K): TET; <i>ant</i> (6)- <i>la</i> : STR; <i>blaZ</i> : PEN; <i>aac</i> (6')- <i>le</i> – <i>aph</i> (2')- <i>la</i> : GEN-KAN; <i>mecA</i> : OXA
M744/10	subclinical	<i>str</i> : STR; <i>cat</i> _{PC221} : CHL; <i>aac</i> (6')- <i>le</i> – <i>aph</i> (2')- <i>la</i> : GEN-KAN; <i>erm</i> (C): CLI, ERY; <i>mecA</i> : OXA
M1383/10	subclinical	<i>erm</i> (C): CLI, ERY; <i>blaZ</i> : PEN; <i>aac</i> (6')- <i>le</i> – <i>aph</i> (2')- <i>la</i> : GEN-KAN; <i>mecA</i> : OXA
M8/10	clinical	<i>ant</i> (6)- <i>la</i> : STR; <i>blaZ</i> : PEN; <i>cat</i> _{PC221} : CHL; <i>aph</i> (3')- <i>III</i> : KAN; <i>erm</i> (C): CLI, ERY; <i>mecA</i> : OXA
M703/10	subclinical	<i>blaZ</i> : PEN; <i>msr</i> : ERY; <i>mecA</i> : OXA
M4298-1/09	subclinical	<i>tet</i> (K): TET; <i>str</i> : STR; <i>blaZ</i> : PEN; <i>cat</i> _{PC221} : CHL
M4233-1/09	subclinical	<i>erm</i> (C): CLI-ERY; <i>tet</i> (K): TET; <i>str</i> : STR
M523/10	clinical	<i>tet</i> (K): TET; <i>str</i> : STR; <i>blaZ</i> : PEN
M864/10	subclinical	<i>blaZ</i> : PEN; <i>tet</i> (K): TET
M1725/10	clinical	<i>str</i> : STR; <i>msr</i> : ERY
M1662/10	clinical	<i>blaZ</i> : PEN
M1977/10	subclinical	<i>str</i> : STR

Figure 1. PFGE, SCCmec types, ST types, and ACME types of 13 *S. epidermidis* isolates



Discussion & Conclusion

The use of antibiotics in dairy farms has likely selected for multidrug-resistant *S. epidermidis*. The isolates are resistant to antibiotics which are widely used in the treatment of mastitis. They belonged to specific sequence types, with ST111 being the most common. The presence of the ACME in *S. epidermidis* may favor colonization of bovine udder and contribute to the spread of multidrug-resistant *S. epidermidis* in dairy cows.

Antibiotic resistance in coagulase-negative staphylococci from bovine mastitis

Yvonne Frey and Vincent Perreten

Institute of Veterinary Bacteriology, Vetsuisse Faculty, University of Bern, Bern, Switzerland

Oral presentation held at the workshop “Bovine mastitis research in Switzerland” of the VPH institute of the Vetsuisse-Faculty Bern, in Bern; 16 March 2012

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Vetsuisse Faculty, University of Bern



Workshop bovine mastitis research in Switzerland; March 16, 2012

Introduction and origin of the milk samples

- > CNS = coagulase-negative staphylococci
Cocci, Gram – positive; Coagulase – negative
- > CNS were isolated from milk which was sent to the IVB for diagnostic in 2009 & 2010
 - dairy milk cows
 - cantons: BE, JU, FR, VD and LU
- > CNS were isolated from:
 - clinical mastitis (n=121)
 - subclinical mastitis (n = 276)



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Workshop bovine mastitis research in Switzerland; March 16, 2012

Identification of CNS

- > Identification of the isolates using MALDI TOF
 - Mass spectrometer (Matrix-Assisted-Laser-Desorption/Ionization–Time-Of-Flight–Mass-Spectrometry)
 - Mass analysis of ribosomal proteins
(< 1min for species identification)
- > MALDI TOF results were validated with 16S rDNA sequences.



3

Workshop bovine mastitis research in Switzerland; March 16, 2012

Incidence of CNS...

... in clinical mastitis (n= 121) ... in subclinical mastitis (n=276)

— <i>S. xylosus</i>	42.2 %	— <i>S. xylosus</i>	39.9 %
— <i>S. chromogenes</i>	15.7 %	— <i>S. chromogenes</i>	15.9 %
— <i>S. sciuri</i>	9.9 %	— <i>S. haemolyticus</i>	9.4 %
— <i>S. haemolyticus</i>	6.6 %	— <i>S. sciuri</i>	9.1 %
— <i>S. devriesi</i>	5.8 %	— <i>S. simulans</i>	5.1 %
— <i>S. vitulinus</i>	4.1 %	— <i>S. devriesi</i>	4.7 %
— <i>S. warneri</i>	3.3 %	— <i>S. epidermidis</i>	3.6 %
— <i>S. epidermidis</i>	3.3 %	— <i>S. succinus</i>	2.9 %
— <i>S. fleuretti</i>	3.3 %	— <i>S. warneri</i>	2.2 %
— others	each < 1.0 %	— <i>S. fleuretti</i>	1.8 %
		— others	each < 1.5 %



4

Workshop bovine mastitis research in Switzerland; March 16, 2012

Antimicrobial resistance testing

- > Determination of the phenotypical antibiotic resistance
 - Micro dilution in Mueller Hinton Bouillon
 - Sensititre susceptibility plates with 19 different antibiotics
 - Resistance breakpoints → EUCAST guidelines (www.eucast.org)
- > Determination of the antibiotic resistance genes
 - Microarray



5

Workshop bovine mastitis research in Switzerland; March 16, 2012

Phenotypical antimicrobial resistance in CNS

> Resistance in %

	ERY	CLI	TET	STR	GEN	KAN	FUS	SMX	TMP	OXA	PEN
CNS from clinical mastitis (n=121)	5.8	3.3	16.5	9.1	5.8	5.0	39.7	11.6	5.0	58.7	21.5
CNS from subclinical mastitis (n=276)	6.8	4.3	13.3	10.8	1.8	2.2	62.1	2.9	4.3	43.0	24.4

indicator for methicillin-resistance
→ Confirmation by detection of *mecA*



6



Methicillin-resistant CNS

- > Confirmation by micro array and PCR:
→ 32.2% (n=64) of the oxacillin-resistant CNS carried a *mecA*.
- > In total, 15% of CNS contained the *mecA* gene.

mecA gene → resistance to all β -lactam-antibiotics and combinations of β -lactam/ β -lactamase inhibitors

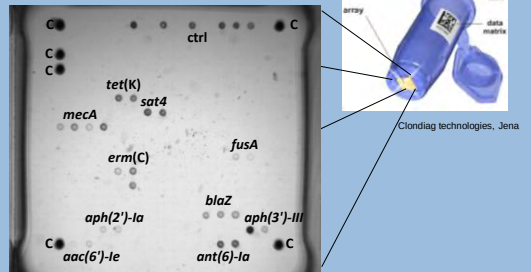
Methicillin-resistant staphylococci often are resistant to several other antibiotics.

7



Detection of AB – resistance genes

- > Antibiotic-resistance genes of *S. epidermidis* (M1186/10) from clinical mastitis



8



Antibiotic Resistances of the *S. epidermidis* isolate M1186/10

Beta-Lactams:

- Amoxicillin/clavulanic acid
- Cephalosporins
- Oxacillin
- Penicillin

Macrolides (e.g. Spiramycin)

Lincosamides (e.g. Clindamycin)

Tetracyclines

Aminoglycosides:

- Gentamicin/Kanamycin
- Kanamycin/Neomycin
- Streptomycin

Resistance genes:

- mecA*
- blaZ*
- erm(C)*
- tetK*
- aac(6')-Ie – aph(2')-Ia*
- aph(3')-III*
- ant(6)-Ia*

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Multiresistance in methicillin-resistant CNS

- > Methicillin-resistant CNS from bovine mastitis showed additional resistances to:

— Tetracycline	19.1%	6% of all tested CNS were multiresistant!
— Streptomycin	14.7%	
— Erythromycin	8.8%	
— Clindamycin	8.8%	
— Gentamicin/Kanamycin	8.8%	
— Penicillin	7.4%	
— Kanamycin/Neomycin	5.9%	
— Trimethoprim	4.4%	

The most frequently occurring strains were *S. epidermidis* and *S. sciuri*.

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Summary

- > The most common CNS – species in mastitis milk were: *S. xylosus*, *S. chromogenes*, *S. sciuri* and *S. haemolyticus* as identified by using rapid identification tool (MALDI TOF).
- > 15% of the isolates carried a *mecA* gene, some of them were resistant to other classes of drugs.
- > No significant differences were observed between resistances in CNS from subclinical and clinical mastitis.
- > Multiresistant CNS are present in bovine mastitis; some CNS isolates have resistance genes making them resistant to frequently used antibiotic in bovine mastitis treatment.
- > Antibigram is recommended for targeted antimicrobial treatment.

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- > Vincent Perreten
- > Joan Pena Rodriguez
- > Andreas Thomann
- > Sybille Schwendener
- > Juliette Wipf
- > Alexandra Collaud

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APPENDIX

Strains used in this thesis

name, storage number, species, anamnesis, antibiotic resistance profiles and detected antibiotic resistance genes
115 isolates from clinical mastitis

Name	Storage number	Species	Anamnesis	Break-points µg/ml	MIC = minimal inhibitory concentration in µg/ml											Resistance genes								
					CLI	TET	RIF	STR	FUS	PEN	CHL	KAN	TIA	SYN	VAN	GEN	TMP	ERY	CIP	OXA	LZD	MUP	SMX	
M8/10	21335	<i>S. epidermidis</i>	clinical	<=0.12	1	<=0.016	>32	<=0.5	>2	64	>64	<=0.5	<=0.5	<=1	<=1	<=2	>8	<=0.25	4	<=1	<=0.5	>256	ant(6)-Ia; cat _{p221} ; aph(3)-III; erm(C); mecA	
M36/10	21342	<i>S. xyloso</i>	clinical	<=0.12	<=0.5	0.03	<=4	1	<=0.12	8	<=4	<=0.5	1	2	<=1	<=2	<=0.25	0.5	0.5	2	<=0.5	<=64	n.r.	
M39-1/10	21343	<i>S. devriesei</i>	clinical	<=0.12	<=0.5	<=0.016	<=4	<=0.5	0.25	<=4	<=4	<=0.5	<=0.5	<=1	<=1	8	<=0.25	<=0.25	<=0.25	2	1	<=64	blaZ	
M39-2/10	21344	<i>S. xyloso</i>	clinical	<=0.12	<=0.5	0.03	<=4	<=0.5	<=0.12	<=4	<=4	<=0.5	<=0.5	2	<=1	<=2	<=0.25	<=0.25	0.5	<=1	<=0.5	<=64	n.r.	
M50/10	21347	<i>S. vitulinus</i>	clinical	<=0.12	>16	<=0.016	<=4	1	<=0.12	<=4	<=4	<=0.5	<=0.5	<=1	<=1	4	<=0.25	<=0.25	0.5	0.5	2	<=64	mecA; ter(K)	
M65-2/10	21348	<i>S. vitulinus</i>	clinical	0.25	<=0.5	<=0.016	<=4	2	<=0.12	8	<=4	<=0.5	<=0.5	<=1	<=1	8	<=0.25	<=0.25	0.5	0.5	2	<=64	n.r.	
M143/10	21369	<i>S. fleurettii</i>	clinical	<=0.12	<=0.5	<=0.016	<=4	1	0.25	<=4	<=4	<=0.5	<=0.5	<=1	<=1	<=2	<=0.25	<=0.25	0.5	4	2	<=64	mecA	
M150/10	21371	<i>S. chromogenes</i>	clinical	<=0.12	<=0.5	<=0.016	<=4	<=0.5	1	<=4	<=4	<=0.5	<=0.5	<=1	<=1	<=2	<=0.25	<=0.25	<=0.25	<=1	<=0.5	<=64	blaZ	
M269/10	21425	<i>S. chromogenes</i>	clinical	<=0.12	<=0.5	<=0.016	<=4	<=0.5	<=0.12	8	<=4	<=0.5	<=0.5	2	<=1	<=2	0.5	<=0.25	<=0.25	<=0.25	2	<=0.5	<=64	n.r.
M277/10	21427	<i>S. fleurettii</i>	clinical	<=0.12	<=0.5	<=0.016	<=4	2	0.5	8	<=4	<=0.5	1	<=1	<=1	<=2	<=0.25	<=0.25	>8	2	<=0.5	<=64	mecA	
M324/10	21435	<i>S. xyloso</i>	clinical	<=0.12	<=0.5	0.03	<=4	2	<=0.12	8	<=4	<=0.5	1	2	<=1	<=2	<=0.25	<=0.25	0.5	2	<=0.5	<=64	n.r.	
M341/10	21436	<i>S. chromogenes</i>	clinical	<=0.12	<=0.5	<=0.016	<=4	<=0.5	<=0.12	8	<=4	<=0.5	<=0.5	2	<=1	<=2	<=0.25	<=0.25	<=0.25	2	<=0.5	<=64	n.r.	
M344/10	21437	<i>S. xyloso</i>	clinical	0.25	<=0.5	<=0.016	<=4	4	<=0.12	8	<=4	<=0.5	1	<=1	<=1	<=2	0.5	<=0.25	0.5	2	<=0.5	<=64	n.r.	
M346/10	21438	<i>S. devriesei</i>	clinical	<=0.12	<=0.5	<=0.016	<=4	<=0.5	<=0.12	<=4	<=4	<=0.5	<=0.5	<=1	<=1	4	<=0.25	<=0.25	<=0.25	<=1	<=0.5	<=64	n.r.	
M347/10	21439	<i>S. xyloso</i>	clinical	0.25	<=0.5	0.03	<=4	1	<=0.12	8	<=4	<=0.5	1	2	<=1	<=2	0.5	<=0.25	<=0.25	0.5	2	<=0.5	<=64	n.r.
M348/10	21440	<i>S. xyloso</i>	clinical	0.25	<=0.5	<=0.016	<=4	2	<=0.12	<=4	<=4	<=0.5	2	2	<=1	<=2	<=0.25	<=0.25	0.5	0.5	<=1	<=0.5	<=64	n.r.
M361/10	21444	<i>S. xyloso</i>	clinical	<=0.12	<=0.5	<=0.016	<=4	<=0.5	<=0.12	<=4	<=4	<=0.5	<=0.5	<=1	<=1	<=2	<=0.25	<=0.25	<=0.25	<=1	<=0.5	<=64	n.r.	
M404-1/10	21456	<i>S. fleurettii</i>	clinical	<=0.12	<=0.5	<=0.016	<=4	4	<=0.12	<=4	<=4	<=0.5	2	<=1	<=1	<=2	<=0.25	<=0.25	<=0.25	<=1	<=0.5	<=64	mecA	
M441/10	21467	<i>S. xyloso</i>	clinical	<=0.12	<=0.5	0.06	8	1	<=0.12	<=4	<=4	<=0.5	1	<=1	<=1	<=2	0.5	<=0.25	0.5	<=1	<=0.5	<=64	n.r.	
M545/10	3128	<i>S. fleurettii</i>	clinical	0.25	>16	<=0.016	<=4	4	0.25	<=4	<=4	<=0.5	1	<=1	<=1	<=2	<=0.25	<=0.25	0.5	2	<=1	<=0.5	<=64	mecA; ter(K)
M549-1/10	3129	<i>S. vitulinus</i>	clinical	<=0.12	<=0.5	<=0.016	<=4	<=0.5	<=0.12	<=4	<=4	<=0.5	<=0.5	<=1	<=1	<=2	<=0.25	<=0.25	<=0.25	<=1	<=0.5	<=64	sr; ter(K)	
M549-2/10	31210	<i>S. chromogenes</i>	clinical	<=0.12	<=0.5	<=0.016	<=4	<=0.5	1	<=4	<=4	<=0.5	<=0.5	<=1	<=1	<=2	<=0.25	<=0.25	<=0.25	<=1	<=0.5	256	blaZ	
M573/10	31220	<i>S. devriesei</i>	clinical	<=0.12	<=0.5	<=0.016	<=4	<=0.5	<=0.12	<=4	<=4	<=0.5	<=0.5	<=1	<=1	4	<=0.25	<=0.25	<=0.25	<=1	<=0.5	<=64	n.r.	
M645/10	31245	<i>S. xyloso</i>	clinical	<=0.12	<=0.5	<=0.016	<=4	2	<=0.12	<=4	<=4	<=0.5	1	2	<=1	<=2	<=0.25	<=0.25	0.5	0.5	<=1	<=0.5	<=64	n.r.
M683/10	31257	<i>S. equorum</i>	clinical	0.25	<=0.5	<=0.016	8	<=0.5	<=0.12	8	<=4	<=0.5	<=0.5	2	<=1	<=2	<=0.25	<=0.25	<=0.25	<=1	<=0.5	<=64	n.r.	
M711-1/10	31261	<i>S. hyicus</i>	clinical	<=0.12	<=0.5	<=0.016	<=4	<=0.5	<=0.12	<=4	<=4	<=0.5	<=0.5	<=1	<=1	<=2	<=0.25	<=0.25	<=0.25	<=1	<=0.5	<=64	n.r.	
M711-2/10	31268	<i>S. chromogenes</i>	clinical	<=0.12	<=0.5	<=0.016	<=4	<=0.5	2	<=4	<=4	<=0.5	<=0.5	<=1	<=1	<=2	<=0.25	<=0.25	<=0.25	<=1	<=0.5	<=64	blaZ	
M799/10	3136	<i>S. xyloso</i>	clinical	0.25	<=0.5	<=0.016	<=4	2	<=0.12	<=4	<=4	<=0.5	2	2	<=1	<=2	<=0.25	<=0.25	<=0.25	<=1	<=0.5	<=64	n.r.	
M800/10	3137	<i>S. xyloso</i>	clinical	<=0.12	<=0.5	<=0.016	<=4	1	<=0.12	<=4	<=4	<=0.5	1	2	<=1	4	<=0.25	<=0.25	<=0.25	<=1	<=0.5	<=64	n.r.	
M811/10	31313	<i>S. xyloso</i>	clinical	<=0.12	<=0.5	<=0.016	<=4	2	<=0.12	<=4	<=4	<=0.5	1	2	<=1	<=2	0.5	0.5	0.5	<=1	<=0.5	<=64	n.r.	
M874/10	31333	<i>S. haemolyticus</i>	clinical	<=0.12	<=0.5	<=0.016	<=4	<=0.5	<=0.12	<=4	<=4	<=0.5	<=0.5	<=1	<=1	<=2	<=0.25	<=0.25	<=0.25	<=1	<=0.5	<=64	n.r.	
M886/10	31339	<i>S. xyloso</i>	clinical	<=0.12	<=0.5	0.03	16	2	<=0.12	<=4	<=4	<=0.5	<=0.5	2	<=1	<=2	<=0.25	<=0.25	<=0.25	<=1	<=0.5	<=64	n.r.	
M1067/10	31378	<i>S. varneri</i>	clinical	<=0.12	<=0.5	<=0.016	<=4	<=0.5	<=0.12	<=4	<=4	<=0.5	<=0.5	<=1	<=1	<=2	<=0.25	<=0.25	<=0.25	<=1	<=0.5	<=64	n.r.	
M1082/10	31411	<i>S. xyloso</i>	clinical	<=0.12	<=0.5	0.03	<=4	2	<=0.12	<=4	<=4	<=0.5	1	2	<=1	4	<=0.25	<=0.25	0.5	0.5	<=1	<=0.5	<=64	n.r.
M1125-1/10	31424	<i>S. haemolyticus</i>	clinical	<=0.12	<=0.5	<=0.016	<=4	<=0.5	<=0.12	<=4	<=4	<=0.5	<=0.5	<=1	<=1	<=2	<=0.25	<=0.25	<=0.25	<=1	<=0.5	<=64	n.r.	
M1125-2/10	31425	<i>S. fleurettii</i>	clinical	<=0.12	<=0.5	<=0.016	8	4	0.25	<=4	<=4	<=0.5	<=0.5	<=1	<=1	<=2	<=0.25	<=0.25	4	2	<=0.5	<=64	mecA	
M1186/10	31444	<i>S. epidermidis</i>	clinical	>4	>16	<=0.016	>32	<=0.5	>2	<=4	>64	<=0.5	<=0.5	<=1	16	<=2	>8	<=0.25	>8	<=1	<=0.5	>512	ant(6)-Ia; aac(6)-Ie; aph(2)-Ia; aph(3)-III; blaZ; erm(C); mecA; ter(K)	
M1215/10	31457	<i>S. sciuri</i>	clinical	0.5	<=0.5	<=0.016	<=4	4	<=0.12	<=4	<=4	<=0.5	2	<=1	<=1	4	<=0.25	0.5	1	2	2	<=64	mecA; ter(K)	
M1262/10	31467	<i>S. xyloso</i>	clinical	<=0.12	>16	<=0.016	<=4	<=0.5	<=0.12	<=4	<=4	<=0.5	1	<=1	<=1	<=2	<=0.25	<=0.25	<=0.25	<=1	<=0.5	<=64	n.r.	
M1296/10	31476	<i>S. xyloso</i>	clinical	<=0.12	<=0.5	0.03	<=4	1	<=0.12	<=4	<=4	<=0.5	<=0.5	2	<=1	<=2	<=0.25	<=0.25	1	<=1	<=0.5	<=64	n.r.	
M1349/10	4127	<i>S. varneri</i>	clinical	<=0.12	>16	<=0.016	<=4	<=0.5	<=0.12	<=4	<=4	<=0.5	<=0.5	<=1	<=1	<=2	<=0.25	<=0.25	<=0.25	<=1	<=0.50			

Name	Storage number	Species	Anamnesis	Break-points µg/ml	MIC = minimal inhibitory concentration in µg/ml										Resistance genes									
					CLI	TET	RIF	STR	FUS	PEN	CHL	KAN	TIA	SYN	VAN	GEN	TMP	ERY	CIP	OXA	LZD	MUP	SMX	
M1571/10	4347	<i>S. xylosum</i>	clinical	>0.5	<=0.5	>0.5	>0.5	<=4	2	<=0.12	8	<=4	1	2	<=1	<=2	0.5	<=0.25	0.5	2	<=0.5	<=64	n.r.	
M1595-1/10	4358	<i>S. scuri</i>	clinical	<=0.12	<=0.5	<=0.016	<=4	<=4	4	<=0.12	<=4	<=4	>4	2	<=1	<=2	<=0.25	1	1	2	<=0.5	<=64	mecH1	
M1595-2/10	4359	<i>S. xylosum</i>	clinical	<=0.12	<=0.5	<=0.016	<=4	<=4	4	<=0.12	<=4	<=4	>4	1	<=1	<=2	<=0.25	<=0.25	<=1	<=1	<=0.5	<=64	n.r.	
M1618/10	4369	<i>S. xylosum</i>	clinical	0.5	1	<=0.016	<=4	<=4	2	0.25	8	<=4	>4	2	<=1	<=2	0.5	0.5	2	1	<=1	<=64	blaZ	
M1658-1/10	4372	<i>S. chromogenes</i>	clinical	<=0.12	<=0.5	<=0.016	<=4	<=4	<=0.5	<=1	<=4	<=4	>4	<=0.5	<=1	<=2	<=0.25	<=0.25	<=1	<=1	<=0.5	<=64	blaZ	
M1658-2/10	4373	<i>S. xylosum</i>	clinical	<=0.12	>16	<=0.016	<=4	<=4	1	<=0.12	<=4	<=4	1	2	<=1	8	<=0.25	<=0.25	<=1	<=1	<=0.5	<=64	ter(K)	
M1662/10	4411	<i>S. epidermidis</i>	clinical	<=0.12	<=0.5	<=0.016	<=4	<=4	<=0.5	0.25	<=4	<=4	<=0.5	2	<=1	<=2	<=0.25	<=0.25	<=1	<=1	<=0.5	<=64	blaZ; dfr(A)	
M1691-1/10	4431	<i>S. haemolyticus</i>	clinical	<=0.12	<=0.5	<=0.016	16	<=4	16	0.5	<=4	<=4	<=0.5	<=1	<=1	<=2	<=0.25	<=0.25	<=1	<=1	<=0.5	<=64	blaZ	
M1725-1/10	4439	<i>S. epidermidis</i>	clinical	<=0.12	2	<=0.016	<=4	<=4	>32	>4	<=0.12	<=4	<=0.5	2	<=1	<=2	>8	<=0.25	<=0.25	<=1	<=0.5	512	msr; str	
M1736/10	4442	<i>S. auricularis</i>	clinical	<=0.12	<=0.5	<=0.016	<=4	<=4	<=0.5	<=0.12	<=4	<=4	<=0.5	<=1	<=1	4	<=0.25	<=0.25	<=1	<=1	<=0.5	128	n.r.	
M1863/10	5219	<i>S. xylosum</i>	clinical	0.25	<=0.5	0.06	<=4	<=4	1	<=0.12	8	<=4	>4	2	<=1	<=2	0.5	0.5	2	<=1	<=0.5	<=64	n.r.	
M1905/10	5224	<i>S. devriesii</i>	clinical	<=0.12	<=0.5	<=0.016	<=4	<=4	<=0.5	<=0.12	<=4	<=4	<=0.5	<=1	<=1	<=2	<=0.25	<=0.25	<=1	<=1	<=0.5	<=64	n.r.	
M1915/10	5227	<i>S. chromogenes</i>	clinical	<=0.12	<=0.5	<=0.016	<=4	<=4	<=0.5	1	<=4	<=4	<=0.5	<=1	<=1	<=2	<=0.25	<=0.25	<=1	<=1	<=0.5	<=64	blaZ	
M1956/10	5241	<i>S. devriesii</i>	clinical	<=0.12	<=0.5	<=0.016	<=4	<=4	<=0.5	<=0.12	<=4	<=4	<=0.5	<=1	<=1	4	<=0.25	<=0.25	<=1	<=1	<=0.5	<=64	n.r.	
M1965/10	5249	<i>S. scuri</i>	clinical	<=0.12	>16	<=0.016	<=4	32	4	<=0.12	<=4	>64	>4	2	<=1	16	>32	<=0.25	0.5	<=1	<=1	<=0.5	aac(6)-Ie; aph(2)-Ia; dfr(D); mecH1; str; ter(K)	
M1972/10	5251	<i>S. chromogenes</i>	clinical	<=0.12	<=0.5	<=0.016	<=4	<=4	<=0.5	>2	8	<=4	<=0.5	<=1	<=1	<=2	0.5	<=0.25	0.5	2	<=0.5	<=64	blaZ	
M1990/10	5257	<i>S. chromogenes</i>	clinical	<=0.12	<=0.5	<=0.016	<=4	<=4	<=0.5	<=0.12	<=4	<=4	<=0.5	<=1	<=1	<=2	<=0.25	<=0.25	<=1	<=1	<=0.5	<=64	n.r.	
M1997-2/10	5259	<i>S. xylosum</i>	clinical	<=0.12	<=0.5	<=0.016	<=4	<=4	1	<=0.12	64	<=4	<=0.5	2	<=1	4	<=0.25	<=0.25	<=1	<=1	<=0.5	<=64	cat_p233	
M2122/10	5326	<i>S. varneri</i>	clinical	<=0.12	<=0.5	<=0.016	<=4	<=4	<=0.5	<=0.12	<=4	<=4	<=0.5	<=1	<=1	4	<=0.25	<=0.25	<=1	<=1	<=0.5	<=64	n.r.	
M2165/10	5345	<i>S. chromogenes</i>	clinical	<=0.12	<=0.5	<=0.016	<=4	<=4	<=0.5	<=0.12	8	<=4	<=0.5	2	<=1	<=2	<=0.25	<=0.25	2	<=1	<=0.5	256	n.r.	
M2169/10	5349	<i>S. chromogenes</i>	clinical	<=0.12	<=0.5	<=0.016	<=4	<=4	<=0.5	<=0.12	<=4	<=4	<=0.5	<=1	<=1	<=2	<=0.25	<=0.25	<=1	<=1	<=0.5	<=64	n.r.	
M2174/10	5351	<i>S. xylosum</i>	clinical	<=0.12	<=0.5	<=0.016	<=4	<=4	<=0.5	<=0.12	<=4	<=4	<=0.5	<=1	<=1	<=2	>8	<=0.25	<=0.25	<=1	<=0.5	<=64	msr; mph(C)	
M2200/10	5364	<i>S. xylosum</i>	clinical	<=0.12	<=0.5	0.03	<=4	<=4	2	<=0.12	<=4	<=4	<=0.5	2	<=1	<=2	<=0.25	0.5	0.5	<=1	<=0.5	<=64	n.r.	
M2200/10	5364	<i>S. xylosum</i>	clinical	<=0.12	<=0.5	0.03	<=4	<=4	2	<=0.12	<=4	<=4	<=0.5	1	2	<=1	<=0.25	0.5	0.5	<=1	<=0.5	<=64	n.r.	
M2220/10	5370	<i>S. chromogenes</i>	clinical	<=0.12	<=0.5	<=0.016	<=4	<=4	<=0.5	<=0.12	8	<=4	1	<=0.5	<=1	<=2	<=0.25	<=0.25	4	2	<=0.5	<=64	n.r.	
M2307/10	5420	<i>S. chromogenes</i>	clinical	<=0.12	<=0.5	<=0.016	<=4	<=4	<=0.5	<=0.12	8	<=4	<=0.5	2	<=1	<=2	<=0.25	<=0.25	<=1	<=1	<=0.5	256	n.r.	
M2395/10	5461	<i>S. xylosum</i>	clinical	0.25	>16	0.12	<=4	<=4	2	<=0.12	8	<=4	>4	1	2	<=1	<=2	0.5	<=0.25	0.5	2	<=0.5	<=64	ter(K)
M3574/09	12111	<i>S. chromogenes</i>	clinical	<=0.12	<=0.5	<=0.016	<=4	<=4	<=0.5	<=0.12	<=4	<=4	<=0.5	<=1	<=1	<=2	<=0.25	<=0.25	<=1	<=1	<=0.5	512	n.r.	
M3592/09	12118	<i>S. xylosum</i>	clinical	<=0.12	<=0.5	<=0.016	<=4	<=4	4	<=0.12	8	<=4	1	<=0.5	<=1	<=2	1	<=0.25	<=0.25	2	<=0.5	512	n.r.	
M3636/09	12440	<i>S. xylosum</i>	clinical	<=0.12	<=0.5	<=0.016	<=4	<=4	1	<=0.12	<=4	<=4	<=0.5	<=1	<=1	<=2	<=0.25	<=0.25	<=1	<=1	<=0.5	<=64	n.r.	
M3665/09	1251	<i>S. haemolyticus</i>	clinical	<=0.12	<=0.5	<=0.016	<=4	<=4	<=0.5	<=0.12	<=4	<=4	<=0.5	2	<=1	<=2	<=0.25	<=0.25	<=1	<=1	<=0.5	>512	n.r.	
M3668-1/09	1252	<i>S. haemolyticus</i>	clinical	<=0.12	<=0.5	<=0.016	<=4	<=4	<=0.5	<=0.12	<=4	<=4	<=0.5	<=1	<=1	<=2	<=0.25	<=0.25	<=1	<=1	<=0.5	<=64	str	
M3668-2/09	1253	<i>Staph. sp.</i>	clinical	<=0.12	<=0.5	<=0.016	16	<=4	2	<=0.12	<=4	<=4	1	<=0.5	<=1	<=2	0.5	<=0.25	0.5	<=1	<=0.5	<=64	n.r.	
M3709-2/09	12175	<i>S. haemolyticus</i>	clinical	<=0.12	<=0.5	<=0.016	<=4	<=4	<=0.5	<=0.12	<=4	<=4	<=0.5	<=1	<=1	<=2	<=0.25	<=0.25	<=1	<=1	<=0.5	>512	n.r.	
M3719/09	12178	<i>S. chromogenes</i>	clinical	<=0.12	<=0.5	<=0.016	<=4	<=4	<=0.5	1	<=0.12	<=4	<=0.5	<=1	<=1	<=2	<=0.25	<=0.25	<=1	<=1	<=0.5	<=64	blaZ	
M3727/09	1314	<i>S. saprophyticus</i>	clinical	<=0.12	<=0.5	<=0.016	<=4	<=4	4	0.25	<=4	<=4	<=0.5	1	<=1	<=2	<=0.25	<=0.25	<=1	<=1	<=0.5	<=64	blaZ	
M3738/09	1317	<i>S. simulans</i>	clinical	<=0.12	<=0.5	<=0.016	<=4	<=4	<=0.5	<=0.12	8	<=4	<=0.5	<=1	<=1	<=2	<=0.25	<=0.25	2	<=1	<=0.5	256	n.r.	
M3741-1/09	1312	<i>Staph. sp.</i>	clinical	<=0.12	<=0.5	<=0.016	<=4	<=4	2	<=0.12	<=4	<=4	<=0.5	<=1	<=1	<=2	8	1	<=0.25	2	<=0.5	<=64	mph(C)	
M3741-2/09	1313	<i>S. scuri</i>	clinical	0.5	<=0.5	<=0.016	<=4	<=4	4	<=0.12	<=4	<=4	>4	2	<=1	4	<=0.25	1	1	2	<=1	<=64	mecH1	
M3742-1/09	1314	<i>S. xylosum</i>	clinical	0.25	<=0.5	0.06	<=4	<=4	2	<=0.12	<=4	<=4	>4	2	<=1	<=2	<=0.25	0.5	0.5	2	<=0.5	<=64	n.r.	
M3777-1/09	1324	<i>S. xylosum</i>	clinical	<=0.12	>16	<=0.016	<=4	<=4	2	<=0.12	<=4	<=4	<=0.5	<=1	<=1	<=2	0.5	<=0.25	0.5	2	<=0.5	<=64	ter(K)	
M3777-2/09	1325	<i>S. chromogenes</i>	clinical	<=																				

255 isolates from subclinical mastitis

Name	Storage number	Species	Anamnesis	Break-points µg/ml	MIC = minimal inhibitory concentration in µg/ml										Resistance genes									
					CLI	TET	RIF	STR	FUS	PEN	CHL	KAN	TIA	SYN	VAN	GEN	TMP	ERY	CIP	OXA	LZD	MUP	SMX	
M13-1/10	21339	<i>S. xylosum</i>	subclinical	<=0.12	<=0.5	0.06	<=4	1	<=0.12	<=4	<=4	<=4	2	1	2	<=1	<=2	<=0.25	0.5	0.5	<=1	<=0.5	<=64	n.r.
M13-2/10	21336	<i>S. scuri</i>	subclinical	0.25	<=0.5	<=0.016	<=4	4	<=0.12	<=4	<=4	<=4	>4	2	<=1	<=1	<=2	<=0.25	0.5	0.5	<=1	<=0.5	<=64	mecA1
M17/10	21337	<i>S. chromogenes</i>	subclinical	<=0.12	<=0.5	<=0.016	<=4	<=0.5	<=0.12	<=4	<=4	<=4	<=4	<=0.5	<=1	<=1	<=2	<=0.25	<=0.25	<=0.25	<=1	<=0.5	<=64	n.r.
M18/10	21338	<i>S. vitulinus</i>	subclinical	<=0.12	<=0.5	<=0.016	<=4	1	<=0.12	<=4	<=4	<=4	1	<=0.5	<=1	<=1	4	<=0.25	<=0.25	0.5	2	<=0.5	<=64	n.r.
M31/10	21340	<i>S. chromogenes</i>	subclinical	<=0.12	<=0.5	<=0.016	32	<=0.5	1	<=0.12	<=4	<=4	<=4	<=0.5	<=0.5	<=1	<=2	<=0.25	<=0.25	0.5	<=1	<=0.5	<=64	blaZ, str
M35/10	21341	<i>S. haemolyticus</i>	subclinical	<=0.12	<=0.5	<=0.016	32	<=0.5	0.5	<=0.12	<=4	<=4	<=4	<=0.5	<=0.5	<=1	<=2	<=0.25	<=0.25	<=0.25	<=1	<=0.5	<=64	blaZ, str
M46/10	21345	<i>S. chromogenes</i>	subclinical	<=0.12	>16	<=0.016	32	<=0.5	1	<=0.12	<=4	<=4	<=4	<=0.5	<=0.5	<=1	<=2	<=0.25	<=0.25	<=0.25	<=1	<=0.5	<=64	blaZ, str; ter(K)
M47/10	21346	<i>S. chromogenes</i>	subclinical	<=0.12	>16	<=0.016	>32	<=0.5	>2	<=0.12	<=4	>64	<=4	<=0.5	<=0.5	<=1	16	<=0.25	<=0.25	1	2	<=0.5	<=64	aac(6)-Ic; aph(2)-Ia; blaZ, str; ter(L)
M67/10	21349	<i>S. xylosum</i>	subclinical	<=0.12	<=0.5	<=0.016	<=4	2	<=0.12	<=4	<=4	<=4	<=0.5	<=0.5	<=1	<=1	<=2	0.5	<=0.25	<=0.25	<=1	<=0.5	<=64	n.r.
M68-1/10	21350	<i>S. xylosum</i>	subclinical	0.25	<=0.5	0.03	<=4	1	<=0.12	<=4	<=4	<=4	>4	1	<=1	<=1	<=2	<=0.25	<=0.25	0.5	<=1	<=0.5	<=64	n.r.
M124/10	21362	<i>S. xylosum</i>	subclinical	0.5	<=0.5	0.06	<=4	1	<=0.12	<=4	<=4	<=4	>4	2	<=1	<=1	<=2	<=0.25	<=0.25	0.5	<=1	<=0.5	<=64	n.r.
M130/10	21364	<i>S. haemolyticus</i>	subclinical	>4	<=0.5	<=0.016	<=4	<=0.5	<=0.12	<=4	<=4	<=4	<=4	<=0.5	2	<=1	<=2	<=0.25	<=0.25	<=0.25	<=1	<=0.5	<=64	erm(C)
M133-2/10	21366	<i>S. chromogenes</i>	subclinical	<=0.12	<=0.5	<=0.016	>32	<=0.5	<=0.5	<=0.12	<=4	<=4	<=4	<=0.5	<=0.5	<=1	<=2	<=0.25	<=0.25	0.5	<=1	<=0.5	<=64	blaZ, str
M161/10	21372	<i>S. hyicus</i>	subclinical	<=0.12	<=0.5	<=0.016	<=4	<=0.5	<=0.12	<=4	<=4	<=4	<=4	<=0.5	<=0.5	<=1	<=2	<=0.25	<=0.25	<=0.25	<=1	<=0.5	<=64	n.r.
M163/10	21373	<i>S. haemolyticus</i>	subclinical	<=0.12	<=0.5	<=0.016	<=4	<=0.5	<=0.12	<=4	<=4	<=4	<=4	<=0.5	<=1	<=1	<=2	<=0.25	<=0.25	<=0.25	<=1	<=0.5	<=64	blaZ
M173-2/10	21377	<i>S. xylosum</i>	subclinical	0.25	>16	<=0.016	<=4	2	<=0.12	<=4	<=4	<=4	>4	1	<=1	<=1	<=2	<=0.25	<=0.25	<=0.25	<=1	<=0.5	<=64	blaZ ter(L)
M192/10	2143	<i>S. chromogenes</i>	subclinical	<=0.12	<=0.5	<=0.016	8	<=0.5	<=0.12	8	<=4	<=4	<=4	<=0.5	<=0.5	<=1	<=2	<=0.25	<=0.25	<=0.25	<=1	<=0.5	256	n.r.
M232/10	21411	<i>S. hyicus</i>	subclinical	<=0.12	<=0.5	<=0.016	<=4	<=0.5	<=0.12	<=4	<=4	<=4	<=4	<=0.5	<=0.5	<=1	<=2	<=0.25	<=0.25	<=0.25	<=1	<=0.5	<=64	n.r.
M243-2/10	21414	<i>S. chromogenes</i>	subclinical	<=0.12	<=0.5	<=0.016	16	<=0.5	<=0.12	<=4	<=4	<=4	<=4	<=0.5	<=0.5	<=1	<=2	<=0.25	<=0.25	<=0.25	<=1	<=0.5	<=64	blaZ
M250/10	21426	<i>S. xylosum</i>	subclinical	0.25	>16	0.06	<=4	1	<=0.12	<=4	<=4	<=4	>4	1	<=1	<=1	<=2	<=0.25	<=0.25	<=0.25	<=1	<=0.5	<=64	blaZ ter(K)
M251/10	21416	<i>S. xylosum</i>	subclinical	<=0.12	<=0.5	0.06	<=4	2	<=0.12	<=4	<=4	<=4	>4	1	<=1	<=1	<=2	<=0.25	<=0.25	0.5	<=1	<=0.5	<=64	n.r.
M292/10	21431	<i>S. succinus</i>	subclinical	<=0.12	<=0.5	0.12	<=4	<=0.5	<=0.12	8	<=4	<=4	2	1	2	<=1	<=2	<=0.25	<=0.25	<=0.25	2	<=0.5	<=64	n.r.
M349/10	21441	<i>S. xylosum</i>	subclinical	0.25	<=0.5	<=0.016	<=4	1	<=0.12	<=4	<=4	<=4	>4	1	2	<=1	<=2	<=0.25	<=0.25	<=0.25	<=1	<=0.5	<=64	n.r.
M353-1/10	21442	<i>S. xylosum</i>	subclinical	0.25	<=0.5	<=0.016	<=4	1	<=0.12	<=4	<=4	<=4	>4	1	2	<=1	<=2	<=0.25	<=0.25	<=0.25	<=1	<=0.5	<=64	n.r.
M353-2/10	21443	<i>S. scuri</i>	subclinical	0.25	<=0.5	<=0.016	<=4	2	<=0.12	<=4	<=4	<=4	>4	2	<=1	<=1	<=2	<=0.25	<=0.25	0.5	<=1	<=0.5	<=64	mecA1
M372/10	21447	<i>S. xylosum</i>	subclinical	<=0.12	<=0.5	<=0.016	<=4	2	<=0.12	<=4	<=4	<=4	>4	1	<=1	<=1	<=2	<=0.25	<=0.25	0.5	<=1	<=0.5	<=64	n.r.
M374/10	21448	<i>S. vitulinus</i>	subclinical	<=0.12	<=0.5	<=0.016	<=4	1	<=0.12	<=4	<=4	<=4	>4	<=0.5	<=1	<=1	<=2	<=0.25	<=0.25	0.5	<=1	<=0.5	<=64	n.r.
M376-1/10	21449	<i>S. xylosum</i>	subclinical	<=0.12	<=0.5	<=0.016	<=4	1	<=0.12	<=4	<=4	<=4	>4	<=0.5	<=0.5	<=1	<=2	<=0.25	<=0.25	0.5	<=1	<=0.5	<=64	n.r.
M376-2/10	21450	<i>S. scuri</i>	subclinical	0.25	<=0.5	<=0.016	<=4	4	<=0.12	<=4	<=4	<=4	>4	2	<=1	<=1	<=2	<=0.25	<=0.25	0.5	<=1	<=0.5	<=64	n.r.
M383/10	21451	<i>S. similans</i>	subclinical	<=0.12	>16	<=0.016	32	<=0.5	<=0.12	<=4	<=4	<=4	>4	<=0.5	<=1	<=1	<=2	<=0.25	<=0.25	<=0.25	<=1	<=0.5	<=64	mecA1
M400/10	21454	<i>S. auricularis</i>	subclinical	<=0.12	<=0.5	<=0.016	<=4	<=0.5	<=0.12	<=4	<=4	<=4	>4	<=0.5	<=1	<=1	<=2	<=0.25	<=0.25	<=0.25	<=1	<=0.5	128	blaZ
M409/10	21458	<i>S. chromogenes</i>	subclinical	<=0.12	<=0.5	<=0.016	<=4	<=0.5	<=0.12	8	<=4	<=4	>4	<=0.5	<=0.5	<=1	<=2	0.5	<=0.25	<=0.25	2	<=0.5	<=64	n.r.
M414/10	21459	<i>S. chromogenes</i>	subclinical	<=0.12	<=0.5	<=0.016	32	<=0.5	1	<=0.12	<=4	<=4	<=4	<=0.5	<=0.5	<=1	<=2	<=0.25	<=0.25	<=0.25	<=1	<=0.5	<=64	blaZ, str
M416-1/10	21460	<i>S. devriesei</i>	subclinical	<=0.12	<=0.5	<=0.016	32	<=0.5	0.25	<=0.12	<=4	<=4	<=4	<=0.5	<=0.5	<=1	<=2	<=0.25	<=0.25	<=0.25	<=1	<=0.5	<=64	blaZ, str
M416-2/10	21461	<i>S. equorum</i>	subclinical	<=0.12	<=0.5	<=0.016	<=4	<=0.5	<=0.12	<=4	<=4	<=4	2	1	<=1	<=1	<=2	8	<=0.25	<=0.25	<=1	<=0.5	<=64	n.r.
M425/10	21462	<i>S. chromogenes</i>	subclinical	>4	>16	<=0.016	16	<=0.5	0.25	64	<=4	<=4	<=0.5	<=0.5	<=1	<=1	<=32	>8	<=0.25	<=0.25	<=1	<=0.5	<=64	erm(B); ter(L); blaZ; cat p _{CD1} ; dfr(K)
M439/10	21466	<i>S. xylosum</i>	subclinical	0.25	<=0.5	<=0.016	<=4	1	<=0.12	<=4	<=4	<=4	>4	1	2	<=1	<=2	<=0.25	<=0.25	0.5	<=1	<=0.5	<=64	n.r.
M532-1/10	3126	<i>S. scuri</i>	subclinical	0.5	<=0.5	<=0.016	<=4	4	<=0.12	<=4	<=4	<=4	>4	2	<=1	<=1	<=2	<=0.25	<=0.25	1	<=1	<=0.5	<=64	mecA1
M532-2/10	3127	<i>S. xylosum</i>	subclinical	<=0.12	<=0.5	0.06	<=4	1	<=0.12	<=4	<=4	<=4	>4	1	2	<=1	<=2	<=0.25	<=0.25	<				

Name	Storage number	Species	Anamnesis	Break-points µg/ml	MIC = minimal inhibitory concentration in µg/ml										Resistance genes								
					CLI	TET	RIF	STR	FUS	PEN	CHL	KAN	TIA	SYN	VAN	GEN	TMP	ERY	CIP	OXA	LZD	MUP	SMX
M860/10	31336	<i>S. chromogenes</i>	subtechnical	>0.5	<0.12	<0.5	<0.016	<4	>1.0	>0.125	>8.0	>8.0	>2.0	>2.0	>2.0	>1.0	>4.0	>2.0	>1.0	>0.25	>4.0	>256	>128
M864/10	31329	<i>S. epidermidis</i>	subtechnical	<0.12	>16	<0.5	<0.016	<4	<0.5	1	<4	<4	<0.5	<0.5	<1	<1	<2	<0.25	<0.25	0.5	<1	<0.5	<64
M917/10	31349	<i>S. chromogenes</i>	subtechnical	<0.12	<0.5	<0.5	<0.016	<4	<0.5	0.25	<4	<4	<0.5	<0.5	2	<1	<2	<0.25	<0.25	<0.25	<1	<0.5	128
M974/10	31362	<i>S. chromogenes</i>	subtechnical	<0.12	<0.5	<0.5	<0.016	16	<0.5	<0.12	<4	<4	<0.5	<0.5	<1	<1	<2	0.5	<0.25	<0.25	<1	<0.5	<64
M1032/10	31367	<i>S. xylus</i>	subtechnical	0.25	<0.5	<0.5	<0.016	<4	<0.5	<0.12	8	<4	<0.5	<0.5	2	<1	<2	<0.25	<0.25	<0.25	<1	<0.5	<64
M1048/10	31373	<i>S. lentus</i>	subtechnical	1	<0.5	<0.5	<0.016	<4	2	<0.12	8	<4	>4	2	2	<1	4	0.5	1	1	2	1	<64
M1049/10	31374	<i>Staph. sp.</i>	subtechnical	0.5	<0.5	<0.5	<0.016	<4	1	<0.12	<4	<4	>4	2	2	<1	<2	8	0.5	0.5	<1	<0.5	<64
M1051/10	31375	<i>S. sciuri</i>	subtechnical	0.5	<0.5	<0.5	<0.016	<4	4	<0.12	<4	<4	>4	2	<1	<1	4	<0.25	0.5	0.5	<1	<0.5	<64
M1058/10	31377	<i>S. sciuri</i>	subtechnical	0.5	<0.5	<0.5	<0.016	<4	2	<0.12	<4	<4	>4	2	<1	<1	<2	<0.25	0.5	0.5	<1	1	<64
M1085-5/10	31412	<i>S. chromogenes</i>	subtechnical	<0.12	<0.5	0.12	<0.016	<4	4	<0.12	<4	<4	2	1	<1	<1	<2	<0.25	<0.25	0.5	<1	<0.5	<64
M1086/10	31381	<i>S. xylus</i>	subtechnical	0.25	<0.5	<0.5	<0.016	<4	<0.5	<0.12	<4	<4	>4	<0.5	2	<1	<2	<0.25	<0.25	0.5	<1	<0.5	<64
M1093-2/10	3148	<i>S. sciuri</i>	subtechnical	0.25	<0.5	<0.5	<0.016	<4	4	<0.12	<4	<4	>4	2	<1	<1	<2	<0.25	0.5	1	<1	2	<64
M1094-1/10	31414	<i>S. varneri</i>	subtechnical	<0.12	>16	<0.5	<0.016	>32	<0.5	0.25	<4	<4	<0.5	<0.5	<1	<1	<2	<0.25	<0.25	<0.25	<1	<0.5	<64
M1101-1/10	31416	<i>S. haemolyticus</i>	subtechnical	<0.12	<0.5	<0.5	<0.016	<4	<0.5	0.5	<4	<4	<0.5	<0.5	<1	<1	<2	<0.25	<0.25	<0.25	<1	<0.5	<64
M1130-1/10	31427	<i>S. sciuri</i>	subtechnical	0.5	<0.5	<0.5	<0.016	<4	4	<0.12	<4	<4	>4	2	<1	<1	4	<0.25	0.5	0.5	2	1	<64
M1130-2/10	31428	<i>S. haemolyticus</i>	subtechnical	<0.12	<0.5	<0.5	<0.016	<4	<0.5	<0.12	<4	<4	<0.5	<0.5	<1	<1	<2	<0.25	<0.25	<0.25	<1	<0.5	<64
M1143/10	31430	<i>S. chromogenes</i>	subtechnical	<0.12	<0.5	<0.5	<0.016	<4	<0.5	<0.12	<4	<4	<0.5	<0.5	<1	<1	<2	<0.25	<0.25	<0.25	<1	<0.5	<64
M1154-1/10	31431	<i>S. xylus</i>	subtechnical	<0.12	<0.5	<0.5	<0.016	<4	2	<0.12	<4	<4	<0.5	<0.5	<1	<1	<2	<0.25	<0.25	<0.25	<1	<0.5	<64
M1176/10	31436	<i>S. xylus</i>	subtechnical	0.25	>16	0.06	<0.016	<4	1	<0.12	8	<4	>4	1	2	<1	<2	0.5	<0.25	0.5	2	<0.5	<64
M1191-2/10	31441	<i>S. fleuretti</i>	subtechnical	<0.12	<0.5	<0.5	<0.016	<4	4	<0.12	<4	<4	>4	1	<1	<1	<2	<0.25	0.5	0.5	2	<0.5	<64
M1201/10	31447	<i>S. sciuri</i>	subtechnical	0.5	>16	<0.5	<0.016	32	4	<0.12	64	<4	>4	2	<1	<1	<2	<0.25	1	1	<1	2	<64
M1217/10	31448	<i>S. xylus</i>	subtechnical	<0.12	<0.5	0.06	<0.016	<4	2	<0.12	<4	<4	>4	2	<1	<1	<2	<0.25	<0.25	0.5	<1	<0.5	<64
M1219-2/10	31452	<i>S. xylus</i>	subtechnical	<0.12	<0.5	<0.5	<0.016	<4	2	<0.12	<4	<4	<0.5	<0.5	<1	<1	<2	<0.25	<0.25	<0.25	<1	<0.5	<64
M1228/10	31451	<i>S. varneri</i>	subtechnical	<0.12	<0.5	<0.5	<0.016	<4	<0.5	<0.12	<4	<4	<0.5	<0.5	<1	<1	<2	<0.25	<0.25	<0.25	<1	<0.5	<64
M1236-1/10	31458	<i>S. devriesi</i>	subtechnical	<0.12	<0.5	<0.5	<0.016	<4	<0.5	<0.12	<4	<4	<0.5	<0.5	<1	<1	<2	<0.25	<0.25	<0.25	<1	<0.5	<64
M1231-1/10	31454	<i>S. xylus</i>	subtechnical	<0.12	<0.5	<0.5	<0.016	<4	2	<0.12	8	<4	1	1	2	<1	<2	<0.25	<0.25	<0.25	<1	<0.5	<64
M1231-2/10	31455	<i>S. succinus</i>	subtechnical	<0.12	<0.5	0.12	<0.016	<4	<0.5	<0.12	8	<4	1	2	2	<1	<2	<0.25	0.5	0.5	2	<0.5	<64
M1245-1/10	31460	<i>S. sciuri</i>	subtechnical	0.25	<0.5	<0.5	<0.016	<4	4	<0.12	<4	<4	>4	2	<1	<1	<2	0.5	<0.25	0.5	2	<0.5	<64
M1245-2/10	31461	<i>S. xylus</i>	subtechnical	<0.12	>16	<0.5	<0.016	<4	1	<0.12	<4	<4	<0.5	<0.5	2	<1	4	<0.25	<0.25	1	2	2	<64
M1254/10	31463	<i>S. xylus</i>	subtechnical	0.25	>16	<0.5	<0.016	<4	<0.5	<0.12	8	<4	>4	1	2	<1	<2	<0.25	<0.25	<0.25	<1	<0.5	<64
M1256/10	31465	<i>S. varneri</i>	subtechnical	<0.12	<0.5	<0.5	<0.016	32	<0.5	1	<4	<4	<0.5	<0.5	<1	<1	<2	0.5	<0.25	<0.25	2	<0.5	<64
M1263/10	31469	<i>S. haemolyticus</i>	subtechnical	<0.12	<0.5	<0.5	<0.016	<4	<0.5	<0.12	<4	<4	<0.5	<0.5	<1	<1	<2	<0.25	<0.25	<0.25	<1	<0.5	<64
M1292-1/10	31477	<i>S. simulans</i>	subtechnical	0.25	<0.5	<0.5	<0.016	8	<0.5	<0.12	8	<4	2	<0.5	<1	<1	4	<0.25	<0.25	<0.25	2	<0.5	<64
M1300/10	31480	<i>S. devriesi</i>	subtechnical	<0.12	<0.5	<0.5	<0.016	<4	<0.5	0.25	<4	<4	<0.5	<0.5	<1	<1	<2	<0.25	0.5	<0.25	<1	<0.5	<64
M1312/10	4121	<i>S. devriesi</i>	subtechnical	<0.12	<0.5	<0.5	<0.016	32	<0.5	0.5	<4	<4	<0.5	<0.5	<1	<1	<2	<0.25	<0.25	<0.25	<1	<0.5	<64
M1351/10	41230	<i>S. xylus</i>	subtechnical	0.25	<0.5	<0.5	<0.016	<4	1	<0.12	<4	<4	>4	2	4	<1	<2	<0.25	<0.25	<0.25	2	<0.5	<64
M1370/10	41240	<i>S. saprophyticus</i>	subtechnical	<0.12	<0.5	<0.5	<0.016	<4	4	0.5	<4	<4	2	1	<1	<1	4	<0.25	<0.25	1	<1	<0.5	<64
M1375-1/10	41236	<i>S. devriesi</i>	subtechnical	<0.12	<0.5	<0.5	<0.016	<4	<0.5	<0.12	<4	<4	<0.5	<0.5	<1	<1	<2	<0.25	<0.25	<0.25	<1	<0.5	<64
M1375-2/10	41237	<i>S. xylus</i>	subtechnical	<0.12	>16	<0.5	<0.016	<4	2	<0.12	<4	<4	<0.5	<0.5	2	<1	<2	<0.25	<0.25	<0.25	<1	<0.5	<64
M1376/10	41238	<i>S. xylus</i>	subtechnical	<0.12	<0.5	<0.5	<0.016	<4	1	<0.12	<4	<4	>4	1	2	<1	<2	<0.25	<0.25	<0.25	<1	<0.5	<64
M1383/10	41242	<i>S. epidermidis</i>	subtechnical	>4	1	<0.5	<0.016	<4	<0.5	1	<4	64	<0.5	<0.5	2	4	<2	>8	<0.25	1	<1	<0.5	256
M1400/10	41248	<i>S. xylus</i>	subtechnical	<0.12	<0.5	<0.5	<0.016	<4	2	<0.12	<4	<4	1	<0.5	2	<1	<2	<0.25	<0.25	0.5	<1	<0.5	<64
M1405/10	41253	<i>S. devriesi</i>	subtechnical	<0.12	<0.5	<0.5	<0.016	<4	<0.5	0.25	<4	<4	<0.5	<0.5	<1	<1	<2	<0.25	<0.25	<0.25	<1	<0.5	<64
M1410-1/10	41254	<i>S. haemolyticus</i>	subtechnical	>4	<0.5	0.06	<0.016	16	4	<0.12	<4	8	<0.5	<0.5	<1	2	<2	>8	0.5	<0.25	<0.25	<1	<0.5
M1413-2/10	41259	<i>S. chromogenes</i>	subtechnical	<0.12	<0.5	<0.5	<0.016	<4	<0.5	<0.12	<4	<4	<0.5	<0.5	2	<1	<2	<0.25	<0.25	<0.25	<1	<0.5	<64
M1446-1/10	41315	<i>S. xylus</i>	subtechnical	<0.12	<0.5	<0.5	<0.016	<4	2	0.25	<4	<4	4	1	2	<1	<2	0.5	<0.25	0.5	<1	<0.5	<64
M1450-1/10	41317	<i>S. haemolyticus</i>	subtechnical	<0.12	<0.5	<0.5	<0.016	>32	<0.5	<0.12	<4	<4	2	2	4	<1	<2	<0.25	<0.25	<0.25	<1	<0.5	<64
M1451-2/10	41279	<i>S. xylus</i>	subtechnical	0.25	<0.5	<0.5	<0.016	<4	2	<0.12	<4	<4	>4	2	2	<1	<2	<0.25	<0.25	0.5	<1	<0.5	<64
M1455/10	4134	<i>S. xylus</i>	subtechnical	<0.12	>16	0.03	<0.016	<4	1	<0.12	<4	<4	<0.5	<0.5	2	<1	<2	<0.25	<0.25	<0.25	<1	<0.5	<64
M1457/10	41274	<i>S. chromogenes</i>	subtechnical	<0.12	<0.5	<0.5	<0.016	<4	<0.5	<0.12	<4	<4	<0.5	<0.5	2	<1	<2	0.5	<0.25	<0.25	<1	<0.5	<64
M1469/10	4132	<i>S. devriesi</i>	subtechnical	<0.12	<0.5	<0.5	<0.016	<4	<0.5	0.25	<4	<4	<0.5	<0.5	<1	<1	<2	<0.25	<0.25	<0.25	<1	<0.5	<64
M1470-1/10	41310	<i>S. devriesi</i>	subtechnical	<0.12	<0.5	<0.5	<0.016	<4	<0.5	0.25	<4	<4	<0.5	<0.5	<1	<1	<2	<0.25	<0.25	<0.25			

Name	Storage number	Species	Anamnesis	Break-points µg/ml	MIC = minimal inhibitory concentration in µg/ml										Resistance genes								
					CLI	TET	RIF	STR	FUS	PEN	CHL	KAN	TIA	SYN	VAN	GEN	TMP	ERY	CIP	OXA	LZD	MUP	SMX
M1599/10	4361	<i>Staph. sp.</i>	subclinical	>0.5	<0.5	>2.0	<0.5	<4	<0.5	>0.125	>8.0	<4	>8.0	>2.0	2	<1	<2	0.5	>1.0	<0.25	>4.0	<0.5	>128
M1605-1/10	4363	<i>S. haemolyticus</i>	subclinical	<0.12	<0.5	<0.5	<0.016	<4	<0.5	0.5	<4	<4	<4	<0.5	<1	<1	<2	<0.25	<0.25	0.5	<1	<0.5	<64
M1605-2/10	4364	<i>S. chromogenes</i>	subclinical	<0.12	<0.5	<0.5	<0.016	>32	<0.5	0.5	<4	<4	<4	<0.5	<1	<1	<2	<0.25	<0.25	<0.25	<1	<0.5	<64
M1607/10	4367	<i>S. varneri</i>	subclinical	<0.12	<0.5	<0.5	<0.016	<4	<0.5	1	<4	<4	<4	<0.5	<1	<1	<2	<0.25	<0.25	0.5	<1	<0.5	<64
M1608/10	4368	<i>S. simulans</i>	subclinical	<0.12	<0.5	<0.5	<0.016	<4	<0.5	<0.12	8	<4	<4	<0.5	<1	<1	8	<0.25	<0.25	<0.25	<1	<0.5	128
M1608/10	4368	<i>S. succinus</i>	subclinical	<0.12	<0.5	<0.5	<0.016	<4	<0.5	<0.12	8	<4	<4	1	2	<1	<2	0.5	<0.25	<0.25	2	<0.5	<64
M1622/10	4370	<i>S. capitis</i>	subclinical	<0.12	<0.5	<0.5	<0.016	<4	<0.5	0.25	<4	<4	<4	<0.5	<1	<1	<2	<0.25	<0.25	<0.25	<1	<0.5	<64
M1623/10	4381	<i>S. devriesii</i>	subclinical	<0.12	<0.5	<0.5	<0.016	<4	<0.5	0.25	<4	<4	<4	<0.5	<1	<1	<2	<0.25	<0.25	<0.25	<1	<0.5	<64
M1638-1/10	4371	<i>S. haemolyticus</i>	subclinical	<0.12	<0.5	<0.5	<0.016	<4	<0.5	<0.12	<4	<4	<4	<0.5	2	<1	<2	<0.25	<0.25	<0.25	<1	<0.5	<64
M1638-2/10	4374	<i>S. chromogenes</i>	subclinical	<0.12	<0.5	<0.5	<0.016	<4	<0.5	0.5	<4	<4	<4	<0.5	2	<1	<2	<0.25	<0.25	<0.25	<1	<0.5	<64
M1641-2/10	4378	<i>S. xylois</i>	subclinical	0.25	<0.5	<0.5	<0.016	<4	1	<0.12	<4	<4	<4	<0.5	2	<1	<2	<0.25	<0.25	<0.25	<1	<0.5	<64
M1642-1/10	4379	<i>S. xylois</i>	subclinical	1	<0.5	<0.5	0.12	<4	4	<0.12	<4	<4	<4	<0.5	<1	<1	<2	<0.25	0.5	<0.25	<1	<0.5	<64
M1643/10	4376	<i>S. xylois</i>	subclinical	<0.12	<0.5	<0.5	0.06	<4	2	<0.12	<4	<4	<4	<0.5	2	<1	<2	<0.25	<0.25	0.5	<1	<0.5	<64
M1643/10	444	<i>S. chromogenes</i>	subclinical	<0.12	<0.5	<0.5	<0.016	<4	<0.5	1	<4	<4	<4	<0.5	<1	<1	<2	<0.25	<0.25	<0.25	<1	<0.5	<64
M1677-2/10	448	<i>S. xylois</i>	subclinical	<0.12	>16	<0.5	<0.016	<4	1	<0.12	<4	<4	<4	<0.5	1	<1	4	<0.25	<0.25	0.5	<1	<0.5	<64
M1679-1/10	4411	<i>S. scituri</i>	subclinical	0.25	<0.5	<0.5	<0.016	<4	2	<0.12	<4	<4	<4	<0.5	2	<1	<2	<0.25	<0.25	0.5	<1	<0.5	<64
M1679-2/10	4412	<i>S. xylois</i>	subclinical	0.25	<0.5	<0.5	<0.016	<4	1	<0.12	<4	<4	<4	<0.5	1	<1	<2	<0.25	<0.25	0.5	<1	<0.5	<64
M1680-1/10	4413	<i>S. scituri</i>	subclinical	0.5	<0.5	<0.5	<0.016	<4	4	<0.12	<4	<4	<4	<0.5	2	<1	4	<0.25	1	1	2	2	<64
M1680-2/10	4414	<i>S. xylois</i>	subclinical	0.5	<0.5	<0.5	<0.016	<4	4	<0.12	<4	<4	<4	<0.5	2	<1	<2	<0.25	<0.25	0.5	<1	<0.5	<64
M1684-1/10	4420	<i>S. xylois</i>	subclinical	0.25	<0.5	<0.5	<0.016	<4	<0.5	<0.12	<4	<4	<4	<0.5	1	<1	<2	<0.25	<0.25	<0.25	<1	<0.5	<64
M1685/10	4422	<i>S. scituri</i>	subclinical	0.5	<0.5	<0.5	<0.016	<4	4	<0.12	<4	<4	<4	<0.5	2	<1	<2	<0.25	0.5	1	<1	2	<64
M1687-1/10	4423	<i>S. scituri</i>	subclinical	<0.12	<0.5	<0.5	<0.016	<4	2	<0.12	<4	<4	<4	<0.5	2	<1	<2	<0.25	<0.25	<0.25	<1	<0.5	<64
M1687-2/10	4424	<i>S. xylois</i>	subclinical	0.25	<0.5	<0.5	<0.016	<4	2	<0.12	<4	<4	<4	<0.5	1	<1	<2	0.5	<0.25	<0.25	2	<0.5	<64
M1688-1/10	4425	<i>S. scituri</i>	subclinical	0.25	<0.5	<0.5	<0.016	<4	2	<0.12	<4	<4	<4	<0.5	2	<1	<2	<0.25	0.5	0.5	<1	<0.5	<64
M1688-2/10	4426	<i>S. xylois</i>	subclinical	<0.12	<0.5	<0.5	<0.016	<4	1	<0.12	<4	<4	<4	<0.5	2	<1	<2	<0.25	<0.25	<0.25	<1	<0.5	<64
M1689-1/10	4427	<i>S. scituri</i>	subclinical	0.25	<0.5	<0.5	<0.016	<4	4	<0.12	<4	<4	<4	<0.5	2	<1	<2	<0.25	<0.25	0.5	<1	<0.5	<64
M1695/10	4429	<i>S. chromogenes</i>	subclinical	<0.12	<0.5	<0.5	<0.016	>32	<0.5	0.5	<4	<4	<4	<0.5	<1	<1	<2	<0.25	<0.25	0.5	<1	<0.5	<64
M1713/10	4437	<i>S. scituri</i>	subclinical	0.5	<0.5	<0.5	<0.016	16	4	<0.12	<4	<4	<4	<0.5	2	<1	<2	<0.25	1	1	2	2	<64
M1720/10	4432	<i>S. xylois</i>	subclinical	0.25	<0.5	<0.5	<0.016	<4	1	<0.12	<4	<4	<4	<0.5	1	<1	8	0.5	<0.25	0.5	<1	<0.5	<64
M1796/10	4455	<i>S. xylois</i>	subclinical	0.25	<0.5	<0.5	<0.016	<4	1	<0.12	8	<4	<4	<0.5	2	<1	<2	0.5	<0.25	<0.25	2	<0.5	<64
M1811/10	4448	<i>S. xylois</i>	subclinical	<0.12	<0.5	<0.5	0.03	<4	1	<0.12	<4	<4	<4	<0.5	2	<1	<2	<0.25	<0.25	<0.25	<1	<0.5	<64
M1813/10	4471	<i>S. haemolyticus</i>	subclinical	<0.12	<0.5	<0.5	<0.016	<4	<0.5	<0.12	<4	<4	<4	<0.5	<1	<1	<2	<0.25	<0.25	<0.25	<1	<0.5	<64
M1814-1/10	4472	<i>S. epidermidis</i>	subclinical	<0.12	<0.5	<0.5	<0.016	<4	<0.5	<0.12	<4	<4	<4	<0.5	2	<1	<2	<0.25	0.5	<0.25	<1	<0.5	<64
M1814-2/10	4466	<i>S. hominis</i>	subclinical	<0.12	<0.5	<0.5	<0.016	<4	<0.5	0.25	<4	<4	<4	<0.5	2	<1	<2	<0.25	<0.25	<0.25	<1	<0.5	<64
M1816/10	4473	<i>S. xylois</i>	subclinical	<0.12	<0.5	<0.5	0.06	<4	1	<0.12	<4	<4	<4	<0.5	<1	<1	<2	<0.25	<0.25	0.5	<1	<0.5	<64
M1835/10	5121	<i>S. xylois</i>	subclinical	<0.12	<0.5	>16	<0.016	<4	2	<0.12	<4	<4	<4	<0.5	2	<1	<2	<0.25	<0.25	<0.25	<1	<0.5	<64
M1840/10	5122	<i>S. xylois</i>	subclinical	<0.12	<0.5	<0.5	<0.016	16	<0.5	<0.12	<4	<4	<4	<0.5	<1	<1	<2	<0.25	<0.25	<0.25	<1	<0.5	<64
M1853/10	5128	<i>S. xylois</i>	subclinical	<0.12	<0.5	<0.5	<0.016	<4	2	<0.12	<4	<4	<4	<0.5	<1	<1	<2	0.5	<0.25	<0.25	<1	<0.5	<64
M1869/10	5218	<i>S. haemolyticus</i>	subclinical	<0.12	<0.5	<0.5	<0.016	<4	<0.5	0.25	<4	<4	<4	<0.5	<1	<1	<2	<0.25	<0.25	<0.25	<1	<0.5	<64
M1877/10	5216	<i>S. chromogenes</i>	subclinical	<0.12	<0.5	<0.5	<0.016	<4	<0.5	0.25	<4	<4	<4	<0.5	<1	<1	<2	<0.25	<0.25	<0.25	<1	<0.5	<64
M1895/10	5217	<i>S. chromogenes</i>	subclinical	<0.12	<0.5	<0.5	<0.016	<4	<0.5	<0.12	<4	<4	<4	<0.5	<1	<1	<2	<0.25	<0.25	<0.25	<1	<0.5	<64
M1936/10	5231	<i>S. chromogenes</i>	subclinical	<0.12	<0.5	<0.5	<0.016	<4	<0.5	2	8	<4	<4	<0.5	<1	<1	<2	0.5	<0.25	0.5	2	<0.5	128
M1961/10	5245	<i>S. fleuretti</i>	subclinical	0.25	<0.5	<0.5	<0.016	<4	4	0.5	<4	<4	<4	<0.5	1	<1	4	<0.25	0.5	>8	2	<0.5	<64
M1976/10	5253	<i>S. scituri</i>	subclinical	0.5	<0.5	<0.5	<0.016	<4	2	<0.12	<4	<4	<4	<0.5	2	<1	<2	<0.25	<0.25	0.5	2	<0.5	<64
M1977/10	5255	<i>S. epidermidis</i>	subclinical	<0.12	<0.5	<0.5	<0.016	>32	<0.5	<0.12	<4	<4	<4	<0.5	2	<1	<2	<0.25	<0.25	<0.25	<1	<0.5	<64
M1981/10	5254	<i>S. chromogenes</i>	subclinical	<0.12	<0.5	<0.5	<0.016	<4	1	0.5	<4	<4	<4	<0.5	<1	<1	<2	<0.25	<0.25	<0.25	<1	<0.5	<64
M2014-1/10	5272	<i>S. chromogenes</i>	subclinical	<0.12	<0.5	<0.5	<0.016	<4	<0.5	<0.12	<4	<4	<4	<0.5	<1	<1	<2	<0.25	<0.25	<0.25	<1	<0.5	<64
M2014-2/10	5273	<i>S. xylois</i>	subclinical	<0.12	>16	<0.5	<0.016	<4	<0.5	<0.12	<4	<4	<4	<0.5	<1	<1	<2	<0.25	<0.25	0.5	<1	<0.5	<64
M2019/10	5274	<i>S. xylois</i>	subclinical	<0.12	<0.5	<0.5	<0.016	<4	2	<0.12	<4	<4	<4	<0.5	<1	<1	<2	<0.25	<0.25	<0.25	<1	<0.5	<64
M2045/10	5311	<i>S. xylois</i>	subclinical	<0.12	<0.5	<0.5	<0.016	<4	2	<0.12	<4	<4	<4	<0.5	<1	<1	<2	<0.25	<0.25	<0.25	<1	<0.5	<64
M2046/10	5312	<i>S. haemolyticus</i>	subclinical	<0.12	<0.5	<0.5	<0.016	<4	<0.5	<0.12	<4	<4	<4	<0.5	<1	<1	<2	<0.25	<0.25	<0.25	<1	<0.5	<64
M2048/10	5325	<i>S. simulans</i>	subclinical	<0.12	<0.5	<0.5	<0.016	<4	<0.5	<0.12	<4	<4	<4	<0.5	<1	<1	4						

Name	Storage number	Species	Anamnesis	Break-points µg/ml	MIC = minimal inhibitory concentration in µg/ml										Resistance genes									
					CLI	TET	RIF	STR	FUS	PEN	CHL	KAN	TIA	SYN	VAN	GEN	TMP	ERY	CIP	OXA	LZD	MUP	SMX	
M2255/10	5148	<i>Staph. sp.</i>	subclinical	<0.12	<0.5	<0.5	<0.016	<4	<0.5	<0.12	<4	<4	<4	<0.5	<1	<1	<2	<0.25	<0.25	<0.25	<1	<0.5	128	n.r.
M2258/10	5149	<i>S. haemolyticus</i>	subclinical	<0.12	<0.5	<0.016	<0.016	<4	<0.5	<0.12	<4	<4	<4	<0.5	<1	<1	<2	<0.25	<0.25	<0.25	<1	<0.5	<64	n.r.
M2311-1/10	51436	<i>S. xylosum</i>	subclinical	0.5	>16	<0.016	<0.016	<4	2	<0.12	<4	<4	<4	<0.5	2	<1	<2	<8	<0.25	<0.25	0.5	2	<0.5	<64
M2313-1/10	51437	<i>S. haemolyticus</i>	subclinical	<0.12	<0.5	<0.016	<0.016	<4	<0.5	<0.12	<4	<4	<4	<0.5	<1	<1	<2	<0.25	<0.25	<0.25	<1	<0.5	<64	<i>msr</i> , <i>mph</i> (C); <i>ter</i> (K)
M3074-1/09	14130	<i>S. xylosum</i>	subclinical	0.25	>16	<0.016	<0.016	<4	<0.5	<0.12	<4	<4	<4	1	2	<1	<2	0.5	<0.25	<0.25	0.5	<1	<0.5	<64
M3551/09	1211	<i>S. xylosum</i>	subclinical	<0.12	<0.5	<0.016	<0.016	<4	2	<0.12	<4	<4	<4	<0.5	2	<1	<2	<0.25	<0.25	<0.25	<1	<0.5	<64	<i>ter</i> (K)
M3556/09	1216	<i>S. simulans</i>	subclinical	<0.12	<0.5	<0.016	<0.016	<4	<0.5	<0.12	<4	<4	<4	<0.5	<1	<1	4	<0.25	<0.25	<0.25	<1	<0.5	128	n.r.
M3567-1/09	1217	<i>S. xylosum</i>	subclinical	<0.12	<0.5	<0.016	<0.016	<4	<0.5	<0.12	<4	<4	<4	<0.5	2	<1	<2	<0.25	<0.25	<0.25	<1	<0.5	<64	n.r.
M3569-1/09	1212	<i>S. xylosum</i>	subclinical	<0.12	>16	0.06	0.06	8	2	<0.12	<4	<4	<4	<0.5	2	<1	<2	<0.25	<0.25	<0.25	0.5	<1	<0.5	<64
M3579/09	12115	<i>S. xylosum</i>	subclinical	0.25	<0.5	<0.016	<0.016	8	2	<0.12	<4	<4	<4	<0.5	2	<1	<2	<0.25	<0.25	<0.25	0.5	<1	<0.5	<64
M3607/09	1223	<i>S. succinus</i>	subclinical	<0.12	<0.5	0.06	0.06	<4	<0.5	<0.12	<4	<4	<4	<0.5	2	<1	<2	<0.25	<0.25	<0.25	0.5	<1	<0.5	<64
M3608-1/09	12124	<i>S. scituri</i>	subclinical	0.25	<0.5	<0.016	<0.016	<4	2	<0.12	<4	<4	<4	<0.5	2	<1	4	<0.25	0.5	0.5	0.5	<1	2	<64
M3608-2/09	1225	<i>S. xylosum</i>	subclinical	<0.12	<0.5	<0.016	<0.016	<4	2	<0.12	<4	<4	<4	<0.5	2	<1	<2	<0.25	<0.25	<0.25	0.5	<1	<0.5	<64
M3617-1/09	12127	<i>S. xylosum</i>	subclinical	<0.12	>16	<0.016	<0.016	<4	2	<0.12	<4	<4	<4	<0.5	<1	<1	<2	<0.25	<0.25	<0.25	<1	<0.5	<64	<i>ter</i> (K)
M3619/09	12129	<i>S. xylosum</i>	subclinical	0.25	<0.5	<0.016	<0.016	<4	1	<0.12	8	<4	<4	1	<0.5	<1	<2	0.5	<0.25	0.5	2	<0.5	<64	n.r.
M3628/09	12134	<i>S. xylosum</i>	subclinical	0.25	<0.5	0.06	0.06	<4	1	<0.12	<4	<4	<4	<0.5	2	<1	<2	<0.25	<0.25	<0.25	0.5	<1	<0.5	<64
M3629/09	12135	<i>S. chromogenes</i>	subclinical	<0.12	<0.5	<0.016	<0.016	>32	<0.5	>2	8	<4	<4	<0.5	<1	<1	<2	<0.25	<0.25	<0.25	0.5	<1	<0.5	<64
M3631/09	12137	<i>S. chromogenes</i>	subclinical	<0.12	<0.5	<0.016	<0.016	<4	<0.5	>2	8	<4	<4	<0.5	<1	<1	<2	0.5	<0.25	<0.25	<1	<0.5	128	<i>bla</i> Z; <i>str</i>
M3642/09	12138	<i>S. chromogenes</i>	subclinical	<0.12	>16	<0.016	<0.016	<4	<0.5	>2	8	<4	<4	<0.5	<1	<1	<2	<0.25	<0.25	<0.25	0.5	<1	<0.5	<64
M3658-2/09	12148	<i>S. xylosum</i>	subclinical	0.25	<0.5	<0.016	<0.016	<4	2	<0.12	<4	<4	<4	<0.5	2	<1	<2	<0.25	0.5	0.5	0.5	<1	<0.5	<64
M3660/09	12149	<i>S. xylosum</i>	subclinical	0.25	<0.5	<0.016	<0.016	<4	2	<0.12	<4	<4	<4	<0.5	2	<1	<2	<0.25	<0.25	<0.25	0.5	<1	<0.5	<64
M3662/09	12150	<i>S. xylosum</i>	subclinical	<0.12	<0.5	<0.016	<0.016	<4	2	<0.12	<4	<4	<4	<0.5	<1	<1	<2	<0.25	<0.25	<0.25	0.5	<1	<0.5	<64
M3678-2/09	12158	<i>S. scituri</i>	subclinical	0.5	<0.5	<0.016	<0.016	32	4	<0.12	<4	<4	<4	<0.5	<1	<1	4	<0.25	<0.25	<0.25	0.5	<1	<0.5	<64
M3679-2/09	12156	<i>S. xylosum</i>	subclinical	0.25	<0.5	0.03	0.03	<4	1	<0.12	<4	<4	<4	2	<0.5	<1	<2	<0.25	<0.25	<0.25	0.5	<1	2	<64
M3689/09	12162	<i>S. haemolyticus</i>	subclinical	>4	<0.5	0.03	0.03	8	<0.5	<0.12	<4	<4	<4	<0.5	<1	<1	<2	<8	<0.25	<0.25	<0.25	<1	<0.5	<64
M3698-1/09	12169	<i>S. xylosum</i>	subclinical	0.25	<0.5	<0.016	<0.016	<4	2	0.25	8	<4	<4	<0.5	<1	<1	<2	<0.25	<0.25	<0.25	<1	<0.5	<64	<i>erm</i> (C)
M3729-2/09	13131	<i>S. xylosum</i>	subclinical	0.25	<0.5	<0.016	<0.016	<4	2	<0.12	<4	<4	<4	<0.5	2	<1	<2	<0.25	<0.25	<0.25	0.5	<1	<0.5	<64
M3729-3/09	13132	<i>S. chromogenes</i>	subclinical	<0.12	<0.5	<0.016	<0.016	<4	<0.5	>2	<4	<4	<4	<0.5	<1	<1	<2	<0.25	<0.25	<0.25	<1	<0.5	<64	<i>bla</i> Z
M3752/09	13120	<i>S. xylosum</i>	subclinical	0.25	<0.5	0.03	0.03	<4	1	<0.12	<4	<4	<4	<0.5	2	<1	<2	<0.25	<0.25	<0.25	0.5	<1	<0.5	<64
M3761/09	13121	<i>S. xylosum</i>	subclinical	<0.12	<0.5	<0.016	<0.016	<4	1	<0.12	<4	<4	<4	<0.5	<1	<1	8	<0.25	<0.25	<0.25	0.5	<1	<0.5	<64
M3783/09	13129	<i>S. xylosum</i>	subclinical	<0.12	<0.5	0.03	0.03	<4	4	0.25	8	<4	<4	2	<0.5	<1	<2	2	0.5	8	<0.25	<1	<0.5	512
M3786/09	13135	<i>S. haemolyticus</i>	subclinical	<0.12	<0.5	<0.016	<0.016	32	<0.5	0.5	<4	<4	<4	<0.5	<1	<1	<2	<0.25	<0.25	<0.25	<1	<0.5	<64	<i>bla</i> Z; <i>str</i>
M3817/09	13144	<i>S. warneri</i>	subclinical	<0.12	<0.5	<0.016	<0.016	<4	<0.5	<0.12	<4	<4	<4	<0.5	<1	<1	<2	<0.25	<0.25	<0.25	<1	<0.5	<64	n.r.
M3818-1/09	13145	<i>S. chromogenes</i>	subclinical	<0.12	<0.5	<0.016	<0.016	<4	<0.5	0.25	<4	<4	<4	<0.5	<1	<1	<2	<0.25	<0.25	<0.25	<1	<0.5	128	<i>bla</i> Z
M3818-2/09	13146	<i>S. hyicus</i>	subclinical	<0.12	<0.5	<0.016	<0.016	<4	1	<0.12	8	<4	<4	<0.5	<1	<1	<2	<0.25	<0.25	<0.25	2	<0.5	<64	n.r.
M3876/09	13168	<i>S. xylosum</i>	subclinical	0.25	<0.5	<0.016	<0.016	<4	2	<0.12	<4	<4	<4	<0.5	<1	<1	<2	<0.25	<0.25	<0.25	<1	<0.5	<64	n.r.
M3877/09	13169	<i>S. scituri</i>	subclinical	0.25	<0.5	<0.016	<0.016	<4	2	<0.12	<4	<4	<4	<0.5	2	<1	<2	<0.25	<0.25	<0.25	<1	<0.5	<64	n.r.
M3884-1/09	13171	<i>S. xylosum</i>	subclinical	0.25	>16	<0.016	<0.016	<4	2	<0.12	<4	<4	<4	<0.5	2	<1	<2	<0.25	<0.25	<0.25	0.5	<1	<0.5	<64
M3895-2/09	13173	<i>S. succinus</i>	subclinical	<0.12	<0.5	0.12	0.12	<4	<0.5	<0.12	8	<4	<4	<0.5	2	<1	<2	<0.25	<0.25	<0.25	<1	<0.5	<64	<i>mecA1</i>
M3907-2/09	14118	<i>S. simulans</i>	subclinical	<0.12	>16	<0.016	<0.016	<4	<0.5	<0.12	64	<4	<4	<0.5	<1	<1	<2	0.5	<0.25	<0.25	2	<0.5	<64	n.r.
M3961/09	14119	<i>Staph. sp.</i>	subclinical	<0.12	>16	<0.016	<0.016	<4	1	<0.12	<4	<4	<4	<0.5	<1	<1	<2	<0.25	<0.25	<0.25	<1	<0.5	128	<i>ter</i> (K)
M4023/09	14132	<i>S. xylosum</i>	subclinical	<0.12	<0.5	0.06	0.06	<4	1	<0.12	<4	<4	<4	<0.5	<1	<1	<2	<0.25	<0.25	<0.25	0.5	<1	<0.5	<64
M4029/09	14136	<i>S. xylosum</i>	subclinical	<0.12	<0.5	<0.016	<0.016	<4	2	<0.12	<4	<4	<4	<0.5	<1	<1	<2	<0.25	<0.25	<0.25	<1	<0.5	<64	n.r.
M4031/09	14137	<i>S. haemolyticus</i>	subclinical	<0.12	<0.5	<0.016	<0.016	<4	<0.5	<0.12	<4	<4	<4	<0.5	<1	<1	<2	<0.25	<0.25	<0.25	<1	<0.5	<64	n.r.
M4074-2/09	14147	<i>S. scituri</i>	subclinical	0.25	<0.5	<0.016	<0.016	<4	2	<0.12	<4	<4	<4	<0.5	<1	<1	4	<0.25	<0.25	<0.25	<1	<0.5	<64	n.r.
M4187-3/09	14162	<i>S. xylosum</i>	subclinical	<0.12	<0.5	<0.016	<0.016	<4	<0.5	<0.12	8	<4	<4	<0.5	<1	<1	<2	0.5	<0.25	<0.25	<1	<0.5	<64	<i>mecA1</i>
M4195/09	14163	<i>S. xylosum</i>	subclinical	<0.12	<0.5	<0.016	<0.016	<4	1	<0.12	<4	<4	<4	<0.5	<1	<1	<2	<0.25	<0.25	<0.25	2	<0.5	<64	n.r.
M4204/09	14175	<i>S. xylosum</i>	subclinical	<0.12	<0.5	<0.016	<0.016	<4	1	<0.12	<4	<4	<4	<0.5	<1	<1	<2	<0.25	<0.25	<0.25	<1	<0.5	<64	n.r.
M4207-1/09	14169	<i>S. xylosum</i>	subclinical	<0.12	<0.5	0.12	0.12	<4	4	<0.12	<4	<4	<4	1	2	<1	<2	<0.25	<0.25	<0.25	0.5	<1	<0.5	<64
M4207-2/09	14170	<i>S. succinus</i>	subclinical	<0.1																				

Name	Storage number	Species	Anamnesis	Break-points µg/ml	MIC = minimal inhibitory concentration in µg/ml												Resistance genes							
					CLI	TET	RIF	STR	FUS	PEN	CHL	KAN	TIA	SYN	VAN	GEN	TMP	ERY	CIP	OXA	LZD	MUP	SMX	
M4331/09	2/266	<i>Staph. sp.</i>	subclinical	<=0.12	<=0.5	<=0.016	<=4	<=0.5	<=0.12	<=4	8	<=0.5	<=0.5	<=1	<=1	8	<=0.25	<=0.25	<=0.25	<=0.25	<=1	<=0.5	<=128	n.r.
M4341-2/09	2/256	<i>S. xyloos</i>	subclinical	0.5	<=0.5	<=0.016	<=4	<=0.5	<=0.12	<=4	<=4	<=4	2	<=1	<=1	<=2	8	<=0.25	<=0.25	0.5	<=1	<=0.5	<=64	<i>mph(C)</i>
M4347-1/09	2/258	<i>S. denitrisei</i>	subclinical	<=0.12	<=0.5	<=0.016	<=4	<=0.5	0.25	<=4	<=4	<=0.5	<=0.5	<=1	<=1	<=2	<=0.25	<=0.25	0.5	<=0.25	<=1	<=0.5	<=64	<i>blaZ'</i>
M4347-2/09	2/259	<i>S. xyloos</i>	subclinical	<=0.12	<=0.5	0.03	<=4	1	<=0.12	<=4	<=4	2	<=0.5	2	<=1	<=2	<=0.25	<=0.25	0.5	<=1	<=0.5	<=0.5	<=64	n.r.
M4347-3/09	2/260	<i>S. succinus</i>	subclinical	<=0.12	<=0.5	0.12	<=4	<=0.5	<=0.12	8	<=4	<=4	4	1	2	<=2	0.5	<=0.25	0.5	2	<=0.5	<=0.5	<=64	n.r.
M4349-2/09	2/262	<i>S. xyloos</i>	subclinical	0.25	<=0.5	<=0.016	<=4	2	<=0.12	<=4	<=4	<=4	<=4	1	<=1	<=2	0.5	<=0.25	<=0.25	<=0.25	<=1	<=0.5	<=64	n.r.
M4383-1/09	2/276	<i>S. wurneri</i>	subclinical	<=0.12	<=0.5	<=0.016	<=4	<=0.5	<=0.12	<=4	<=4	<=0.5	<=0.5	<=1	<=1	<=2	<=0.25	<=0.25	<=0.25	<=0.25	<=1	<=0.5	<=64	n.r.
M4385-1/09	2/277	<i>S. chromogenes</i>	subclinical	<=0.12	<=0.5	<=0.016	<=4	<=0.5	>2	<=4	<=4	<=4	<=0.5	<=0.5	<=1	<=2	<=0.25	<=0.25	<=0.25	<=0.25	<=1	<=0.5	<=64	<i>blaZ'</i>
M4397/09	2/32	<i>S. chromogenes</i>	subclinical	<=0.12	<=0.5	<=0.016	>32	<=0.5	1	<=4	<=4	<=4	<=0.5	<=0.5	<=1	<=1	<=2	<=0.25	<=0.25	<=0.25	<=0.25	<=1	<=0.5	<i>blaZ', str</i>
M4425/09	2/311	<i>S. simulans</i>	subclinical	0.25	<=0.5	<=0.016	<=4	<=0.5	<=0.12	<=4	<=4	<=4	2	<=0.5	<=1	4	<=0.25	<=0.25	<=0.25	<=0.25	2	<=0.5	<=64	n.r.
M4445/09	2/324	<i>S. equorum</i>	subclinical	0.25	<=0.5	<=0.016	<=4	<=0.5	<=0.12	8	<=4	<=4	2	1	2	<=2	8	<=0.25	<=0.25	<=0.25	2	<=0.5	<=64	<i>mph(C)</i>
M4478/09	2/326	<i>S. scituri</i>	subclinical	0.25	<=0.5	<=0.016	<=4	4	<=0.12	<=4	<=4	<=4	>4	2	<=1	4	<=0.25	<=0.25	0.5	0.5	<=1	2	<=0.5	<i>mecA1</i>
M4506/09	2/333	<i>S. xyloos</i>	subclinical	<=0.12	<=0.5	<=0.016	<=4	4	<=0.12	<=4	<=4	<=4	<=0.5	<=0.5	2	<=1	<=2	<=0.25	<=0.25	0.5	<=1	<=0.5	<=64	n.r.

47 isolates from control milk

Name	Storage number	Species	Anamnesis	Break-points µg/ml	MIC = minimal inhibitory concentration in µg/ml										Resistance genes										
					CLI	TET	RIF	STR	FUS	PEN	CHL	KAN	TIA	SYN	VAN	GEN	TMP	ERY	CIP	OXA	LZD	MUP	SMX		
M107-1/10	21354	<i>S. schleri</i>	control	0.5	<0.5	0.03	<4	4	<0.12	<4	<4	<4	>4	2	<1	<1	4	<0.25	0.5	1	<1	1	<64	<i>mecA-I</i>	
M107-2/10	21355	<i>S. xylois</i>	control	0.25	<0.5	<0.016	<4	2	<0.12	<4	<4	<4	>4	2	2	<1	<2	<0.25	<0.25	0.5	<1	<0.5	<64	n.r.	
M108-1/10	21356	<i>S. simulans</i>	control	<0.12	4	<0.016	<4	<0.5	<0.12	8	<4	<4	1	<0.5	<1	<1	8	<0.25	<0.25	<0.25	2	<0.5	128	<i>ter</i> (K)	
M108-2/10	21357	<i>S. chromogenes</i>	control	<0.12	<0.5	<0.016	<4	<0.5	<0.12	<4	<4	<4	<0.5	<0.5	<1	<1	<2	<0.25	<0.25	<0.25	<1	<0.5	<64	n.r.	
M115-1/10	21359	<i>S. simulans</i>	control	<0.12	>16	<0.016	<4	2	<0.12	<4	<4	<4	<0.5	<0.5	2	<1	<2	<0.25	<0.25	0.5	<1	<0.5	<64	<i>ter</i> (K)	
M119-1/10	21361	<i>S. xylois</i>	control	<0.12	<0.5	<0.016	<4	<0.5	<0.12	<4	<4	<4	<0.5	<0.5	<1	<1	<2	<0.25	<0.25	<0.25	<1	<0.5	<64	n.r.	
M132-1/10	21365	<i>S. scuri</i>	control	<0.12	<0.5	<0.016	<4	4	<0.12	<4	<4	<4	>4	2	<1	<1	<2	<0.25	0.5	0.5	<1	2	<64	<i>mecA-I</i>	
M205-1/10	2146	<i>S. fleuretti</i>	control	1	>16	<0.016	<4	4	0.25	<4	<4	<4	>4	1	<1	<1	<2	8	0.5	8	<1	<0.5	<64	<i>mecA-I</i> ; <i>ter</i> (K)	
M286-1/10	21429	<i>S. warneri</i>	control	<0.12	<0.5	<0.016	<4	<0.5	<0.12	<4	<4	<4	<0.5	<0.5	<1	<1	<2	<0.25	<0.25	<0.25	<1	<0.5	<64	n.r.	
M352-1/10	21463	<i>S. xylois</i>	control	<0.12	<0.5	<0.016	<4	2	<0.12	<4	<4	<4	<0.5	<0.5	2	<1	<2	<0.25	<0.25	<0.25	<1	<0.5	<64	n.r.	
M432-1/10	21480	<i>S. epididymidis</i>	control	<0.12	>16	<0.016	<4	>32	<0.5	0.25	<4	<4	<0.5	<0.5	2	<1	<2	<0.25	<0.25	<0.25	<1	<0.5	<64	<i>blaZ</i> ; <i>str</i> ; <i>ter</i> (K)	
M588-1/10	31223	<i>S. equorum</i>	control	<0.12	<0.5	0.03	<4	<0.5	<0.12	<4	<4	<4	<0.5	<0.5	<1	<1	<2	8	<0.25	<0.25	<0.25	<1	<0.5	<64	<i>mph</i> (C)
M589-1/10	31224	<i>S. cobiti</i>	control	<0.12	<0.5	<0.016	<4	>4	<0.5	>2	8	<4	4	2	<1	<1	<2	<0.25	<0.25	2	2	<0.5	<64	<i>blaZ</i>	
M590-1/10	31226	<i>S. chromogenes</i>	control	<0.12	<0.5	<0.016	<4	<0.5	<0.12	<4	<4	<4	<0.5	<0.5	<1	<1	<2	<0.25	<0.25	0.5	<1	<0.5	<64	n.r.	
M1326-1/10	4129	<i>S. warneri</i>	control	<0.12	>16	<0.016	<4	<0.5	<0.12	<4	<4	<4	<0.5	<0.5	<1	<1	<2	<0.25	<0.25	<0.25	<1	<0.5	<64	<i>ter</i> (K)	
M1327-1/10	41210	<i>S. haemolyticus</i>	control	<0.12	<0.5	<0.016	<4	<0.5	<0.12	<4	<4	<4	<0.5	<0.5	<1	<1	<2	<0.25	<0.25	<0.25	<1	<0.5	<64	n.r.	
M1332-1/10	41214	<i>S. warneri</i>	control	<0.12	>16	<0.016	<4	<0.5	<0.12	<4	<4	<4	<0.5	<0.5	<1	<1	<2	<0.25	<0.25	<0.25	<1	<0.5	<64	<i>ter</i> (K)	
M1334-1/10	41218	<i>S. haemolyticus</i>	control	<0.12	<0.5	<0.016	<4	<0.5	<0.12	<4	<4	<4	<0.5	<0.5	<1	<1	<2	<0.25	<0.25	<0.25	<1	<0.5	<64	n.r.	
M1335-1/10	41215	<i>S. warneri</i>	control	<0.12	>16	<0.016	<4	<0.5	<0.12	<4	<4	<4	<0.5	<0.5	<1	<1	<2	<0.25	<0.25	<0.25	<1	<0.5	<64	<i>ter</i> (K)	
M1336-1/10	41216	<i>S. warneri</i>	control	<0.12	<0.5	<0.016	<4	<0.5	<0.12	<4	<4	<4	<0.5	<0.5	<1	<1	<2	<0.25	<0.25	<0.25	<1	<0.5	<64	<i>ter</i> (K)	
M1343-1/10	41221	<i>S. warneri</i>	control	<0.12	>16	<0.016	<4	<0.5	<0.12	<4	<4	<4	<0.5	<0.5	<1	<1	<2	<0.25	<0.25	<0.25	<1	<0.5	<64	<i>ter</i> (K)	
M1914-1/10	51226	<i>S. haemolyticus</i>	control	<0.12	<0.5	<0.016	<4	<0.5	<0.5	0.25	<4	<4	<0.5	<0.5	<1	<1	4	<0.25	<0.25	<0.25	<1	<0.5	<64	<i>blaZ</i>	
M1975-1/10	51252	<i>S. xylois</i>	control	<0.12	<0.5	0.06	<4	2	<0.12	<4	<4	<4	4	1	2	<1	<2	<0.25	<0.25	0.5	<1	<0.5	<64	n.r.	
M2006-1/10	51266	<i>S. xylois</i>	control	<0.12	<0.5	<0.016	<4	2	<0.12	<4	<4	<4	>4	2	4	<1	<2	<0.25	<0.25	0.5	<1	<0.5	<64	n.r.	
M2026-1/10	51277	<i>S. haemolyticus</i>	control	<0.12	<0.5	<0.016	<4	<0.5	<0.12	<4	<4	<4	<0.5	<0.5	2	<1	4	<0.25	0.5	<0.25	<1	<0.5	<64	n.r.	
M2181-2/10	51356	<i>S. xylois</i>	control	<0.12	<0.5	<0.016	<4	2	<0.12	<4	<4	<4	2	1	2	<1	<2	<0.25	<0.25	0.5	<1	<0.5	<64	n.r.	
M2206-1/10	51371	<i>S. xylois</i>	control	0.25	<0.5	0.06	<4	1	<0.12	<4	<4	<4	>4	2	2	<1	<2	<0.25	<0.25	0.5	<1	<0.5	<64	n.r.	
M3693-2/09	11267	<i>S. warneri</i>	control	<0.12	<0.5	<0.016	<4	<0.5	<0.12	<4	<4	<4	<0.5	<0.5	<1	<1	<2	<0.25	<0.25	<0.25	<1	<0.5	<64	n.r.	
M3724-09	11279	<i>S. xylois</i>	control	<0.12	<0.5	<0.016	<4	1	<0.12	<4	<4	<4	2	1	<1	<1	<2	<0.25	<0.25	<0.25	<1	<0.5	<64	n.r.	
M3747-1/09	11316	<i>S. fleuretti</i>	control	<0.12	<0.5	<0.016	<4	4	<0.12	<4	<4	<4	>4	1	<1	<1	<2	<0.25	<0.25	0.5	<1	<0.5	<64	n.r.	
M3747-2/09	11317	<i>S. equorum</i>	control	0.25	<0.5	<0.016	<4	<0.5	<0.12	8	<4	<4	4	1	<1	<1	<2	<0.25	<0.25	<0.25	2	<0.5	<64	n.r.	
M3794-09	11336	<i>S. chromogenes</i>	control	<0.12	<0.5	<0.016	<4	<0.5	1	<0.12	<4	<4	<0.5	<0.5	<1	<1	<2	<0.25	<0.25	<0.25	<1	<0.5	<64	<i>blaZ</i>	
M3827-09	11347	<i>S. succinus</i>	control	<0.12	<0.5	0.03	<4	<0.5	<0.12	8	<4	<4	1	1	<1	<1	<2	<0.25	<0.25	<0.25	<1	<0.5	<64	<i>ter</i> (K)	
M3873-1/09	11366	<i>S. xylois</i>	control	0.25	>16	0.06	<4	2	<0.12	<4	<4	<4	4	1	2	<1	<2	<0.25	<0.25	<0.25	<1	<0.5	<64	<i>blaZ</i> ; <i>cat</i> ; <i>prc233</i>	
M4091-09	11456	<i>S. cobiti</i>	control	<0.12	1	0.25	<4	4	0.25	64	<4	<4	2	2	<1	<1	<2	>8	<0.25	1	2	<0.5	<64	<i>blaZ</i>	
M4093-09	11457	<i>S. equorum</i>	control	0.25	<0.5	<0.016	8	<0.5	<0.12	8	<4	<4	4	1	2	<1	<2	8	<0.25	0.5	2	<0.5	<64	<i>mph</i> (C)	
M4203-2/09	11474	<i>S. xylois</i>	control	<0.12	>16	0.03	<4	2	<0.12	<4	<4	<4	<0.5	<0.5	<1	<1	<2	<0.25	<0.25	0.5	<1	<0.5	<64	<i>ter</i> (K)	
M4283-09	21220	<i>S. xylois</i>	control	<0.12	<0.5	<0.016	<4	2	<0.12	<4	<4	<4	<0.5	<0.5	<1	<1	<2	0.5	<0.25	0.5	2	<0.5	<64	n.r.	
M4406-09	2133	<i>S. chromogenes</i>	control	<0.12	<0.5	<0.016	<4	<0.5	<0.12	<4	<4	<4	<0.5	<0.5	2	<1	<2	<0.25	<0.25	<0.25	<1	<0.5	<64	n.r.	
M4408-2/09	2138	<i>S. chromogenes</i>	control	<0.12	<0.5	<0.016	<4	<0.5	<0.5	0.25	8	<4	1	<0.5	2	<1	<2	<0.25	<0.25	<0.25	<1	<0.5	<64	<i>blaZ</i>	
M4411-09	2134	<i>S. chromogenes</i>	control	<0.12	<0.5	<0.016	<4	<0.5	<0.12	<4	<4	<4	<0.5	<0.5	<1	<1	<2	<0.25	<0.25	<0.25	<1	<0.5	<64	n.r.	
M4412-09	2139	<i>S. chromogenes</i>	control	<0.12	<0.5	<0.016	<4	<0.5	<0.12	<4	<4	<4	<0.5	<0.5	2	<1	<2	<0.25	<0.25	<0.25	<1	<0.5	<64	n.r.	
M4414-09	2135	<i>S. chromogenes</i>	control	<0.12	<0.5	<0.016	<4	<0.5	<0.12	<4	<4	<4	<0.5	<0.5	<1	<1	<2	<0.25	<0.25	<0.25	<1	<0.5	<64	n.r.	
M4416-09	2136	<i>S. chromogenes</i>	control	<0.12	<0.5	<0.016	<4	<0.5	<0.12	<4	<4	<4	<0.5	<0.5	<1	<1	<2	<0.25	<0.25	<0.25	<1	<0.5	<64	n.r.	
M4422-09	21310	<i>S. chromogenes</i>	control	<0.12	<0.5	<0.016	<4	<0.5	<0.5	0.5	<4	<4	<0.5	<0.5	<1	<1	<2	<0.25	<0.25	<0.25	<1	<0.5	<64	<i>blaZ</i> ; <i>str</i>	
M4438-1/09	21317	<i>S. xylois</i>	control	0.25	<0.5	<0.016	<4	2	<0.12	<4	<4	<4	>4	2	2	<1	<2	<0.25	<0.25	<0.25	<1	<0.5	<64	n.r.	
M4438-2/09	21320	<i>S. succinus</i>	control	0.25	<0.5	<0.016	<4	<0.5	<0.12	8	<4	<4	>4	2	2	<1	<2	0.5	0.5	0.5	2	1	<64	n.r.	

Labelling of genomic DNA (DeRisi method, University of California, San Francisco)

Round A

- 2,0 µl of genomic DNA from lysate
- add 0,5µl **5X Sequenase buffer** + 0,5µl **De RisiA primer** (40pmol/ µl) x No. of samples (→ 1µl mix per sample)
- denature for 2min at 94°C and cool down to 10°C for 5min in thermocycler with “DeRisiA” program
- pipet 1,25 µl of the following mix into the denatured DNA:

Mix for labelling:

for 4 reactions:	for 8 react.:	for 16 react.	for 32 react.:
1 µl 5X Sequenase buffer	2 µl	4 µl	8 µl
1,5 µl 3mM dNTP	3 µl	6 µl	12 µl
0,75 µl 0,1M DTT	1,5 µl	3 µl	6 µl
1,5 µl 500 µg/ml BSA	3 µl	6 µl	12 µl
0,4 µl Sequenase	0,8 µl	1,6 µl	3,2 µl

Caution: add sequenase just before pipeting the mix; directly from the fridge!

- **Program A:**
 - o 8 min at 37°C (ramping from 10°C-37°C within 8 min)
 - o 8 min at 37°C (constant)
 - o 2 min at 94°C
 - o 5 min at 10°C while adding 0,3 µl of the following mix to each sample:
 - for 4 reactions: 0,3 µl **Sequenase** + 0,9 µl **Sequenase Dil buffer**
 - for 8 reactions: 0,6 µl **Sequenase** + 1,8 µl **Sequenase Dil buffer**
 - for 16 reactions: 1,2 µl **Sequenase** + 3,6 µl **Sequenase Dil buffer**
 - o 8 min at 37°C (ramping from 10°C-37°C within 8 min)
 - o 8 min at 37°C (constant)
 - o remove the samples before temperature starts to rise up to 94°C
→ 1 ¾ cycles; duration: ca. 15min
 - o **add 11,0 µl water to the sample**

Round B

Preparation of premix for 8 reactions:	for 12 react.:	for 16 react.	for 32 react.
20 µl 10X PCR buffer von Roche	30 µl	40 µl	80 µl
2 µl 100mM dNTP	3,0 µl	4 µl	8 µl
1,6 µl DeRisiB primer (100pmol/ µl)	2,4 µl	3,2 µl	6,4 µl
146 µl water	219 µl	292 µl	584 µl
2 µl Taq polymerase	3,0 µl	4,0 µl	8,0 µl

- Caution: add polymerase just before pipeting the mix; directly from the fridge!
- fill 21,25 µl from this mix in each PCR tube
- add **3,75µl template** from Round A per reaction
- put in thermocycler and run the “DeRisiB” program (choose „standard 54” and store the following program)

- **Program B:**
 - o 30 sec. at 94°C
 - o 30 sec. at 40°C → 35 cycles; duration: ca. 2h
 - o 2 min at 72 °C

Round C

- | - Preparation of premix for 8 reactions: | for 12 react. | for 16 react. | for 32 react.: |
|--|---------------|---------------|----------------|
| - 20 µl 10X PCR buffer von Roche | 30 µl | 40 µl | 80 µl |
| 21 µl dNTP (1mM dACGTP & 0,65mM dTTP) | 31,5 µl | 42 µl | 84 µl |
| 3,2 µl DeRisi Bbiot | 4,8 µl | 6,4 µl | 12,8 µl |
| 6,2 µl Biotin | 9,3 µl | 12,4 µl | 24,8 µl |
| 2 µl Taq Polymerase | 3,0 µl | 4 µl | 8 µl |
| 119,6 µl water | 179,4 µl | 239,2 µl | 478,4 µl |
- Caution: add polymerase just before pipeting the mix; directly from the fridge!
 - fill **21 µl** of this mix in each PCR tube
 - add **4 µl template** from Round B per reaction
 - put in thermocycler an run the “DeRisiC” program
 - **Program C:**
 - o 30 sec. at 94°C
 - o 30 sec. at 40°C
 - o 30 sec. at 50°C
 - o 2 min. at 72°C
 - 40 cycles; duration: ca. 2h

Micro Array – Hybridization ArrayTubes – peroxidase method

Materials:

- Array tubes (5 pieces in one package)
- Biotin labeled probe (from Round C of the DeRisi PCR)
- Plastic pipettes
- **3XDNA buffer** (hybridization solution)
- **2xSSC/0,2% SDS (1A)**
- **2xSSC/0,01 % Triton (1B)**
- **2xSSC (2)**
- **0,2xSSC (3)**
- 6xSSPE/ 0,005% Triton
- **Blocking powder**
- **Streptavidin – Peroxidase**
- **Peroxidase substrate**

Protocol:

- 1.) Probe denaturation:
 - **10 µl probe** (aus round C) + **90 µl 3DNA buffer** 95°C; 5'
 - cool on ice for 2', after that centrifuge shortly
- 2.) Array washing (tube chips):
 - wash the array with **500µl bidest H2O** 25°C; 5', 550rpm
& take off the liquid with a plastic pipette
 - wash the array with **500µl 3DNA buffer** 25°C; 5', 550rpm
& take off the liquid with a plastic pipette

Caution: Do not touch the fond of the tube!
- 3.) Hybridization:
 - add **denatured probe (100 µl)** to **denatured array**
 - hybridize at least 60 min (better 3h) in the “older” thermocycler at 57°C; slightly shaking (550rpm); put the tubes in the thermocycler directly after adding the DNA – samples

After Hybridization:

- 1.) **Washing**
 - 1 wash with **1A (2xSSC/0,2% SDS)** 500µl; 25°C; 5'; 550 rpm
 - 2 wash with **2 (2xSSC)** 500µl; 25°C; 5'; 550 rpm
 - 3 wash with **3 (0,2xSSC)** 500µl; 25°C; 5'; 550 rpm

Take off the liquid with a plastic pipette, caution: Do not touch the fond of the tube!
- 2.) **Blocking** 100 µl; 30°C; 15'; 550 rpm
 - 2% blocking powder in 6xSSPE/0,005% Triton
 - o for 4 array tubes: **0,01g blocking powder** in 500 µl 6xSSPE/0,005% Triton
 - o for 24 array tubes: **0,06 g blocking powder** in 3000 µl 6xSSPE/0,005% Triton

3.) Conjugation

100 µl; 30°C; 15'; 550 rpm

- dilute **Streptavidin – Peroxidase** 1: 2000 in 6xSSPE/0,005% Triton
 - o for 10 array tubes: 999 µl 6xSSPE/0,005% Triton and 1 µl Streptavidin – Peroxidase
 - o Streptavidin is in the fridge of alexandra's office; drawer Nr. 6; box named „DeRisi”; add it just before pipeting the mix!

4.) Washing after conjugation

- 1 wash with **1B (2xSSC/0,01 % Triton)** 500µl; 25°C; 5'; 550 rpm
- 2 wash with **2 (2xSSC)** 500µl; 25°C; 5'; 550 rpm
- 3 wash with **3 (0,2xSSC)** 500µl; 25°C; 5'; 550 rpm

5.) Precipitation

- add **100 µl Peroxidase-Substrat** and wait for about 10 min
- open the AlereTech program, Icono-clust and Icono-scan; then place the array tube into the ready (25°C) READER and click on the “eye-symbol” in the program
- save the pictures if they're good, otherwise you can wash again with the **Peroxidase-Substrat**

Microarray protocols

Raster

List with >100 antibiotic resistance genes spotted on microarray chip

Five examples for multiresistant CNS strains analyzed by microarray

Software: Iconoclust program, Alere Tech GmbH, Jena, Germany

16x16 o. MM, 150 µm Spotabstand (ID 6854)

Referenzsystem atref_auto11_gerd

16	246	232	233	234	235	236	237	238	239	240	241	242	243	244	245	246
15		218	219	220	221	222	223	224	225	226	227	228	229	230	231	
14	246	203	204	205	206	207	208	209	210	211	212	213	214	215	216	217
13	246	188	189	190	191	192	193	194	195	196	197	198	199	200	201	202
12		173	174	175	176	177	178	179	180	181	182	183	184	185	186	187
11	157	158	159	160	161	162	163	164	165	166	167	168	169	170	171	172
10	141	142	143	144	145	146	147	148	149	150	151	152	153	154	155	156
9	125	126	127	128	129	130	131	132	133	134	135	136	137	138	139	140
8	109	110	111	112	113	114	115	116	117	118	119	120	121	122	123	124
7	93	94	95	96	97	98	99	100	101	102	103	104	105	106	107	108
6	77	78	79	80	81	82	83	84	85	86	87	88	89	90	91	92
5	61	62	63	64	65	66	67	68	69	70	71	72	73	74	75	76
4	45	46	47	48	49	50	51	52	53	54	55	56	57	58	59	60
3	29	30	31	32	33	34	35	36	37	38	39	40	41	42	43	44
2		15	16	17	18	19	20	21	22	23	24	25	26	27	28	
1	246	1	2	3	4	5	6	7	8	9	10	11	12	13	14	246

1 2 3 4 5 6 7 8 9 10 11 12 13 14 15 16

 leer
 Biotin

Entwurf Layout 16x16 ohne MM, 150 µm Spotabstand

Spot-ID	Sondenname	Phenotype
1	be_AAC6-Ie_144	aminoglycoside resistance
2	be_AAC6-Ie_475	aminoglycoside resistance
3	be_AAC6-Ii_396	aminoglycoside resistance
4	be_AAC6-Ii_71	aminoglycoside resistance
5	be_AAC6-Im_15	aminoglycoside resistance
6	be_AAC6-Im_286	aminoglycoside resistance
7	VPAB3_ant3-Ia	aminoglycoside resistance
8	VPAB4_ant3-Ia	aminoglycoside resistance
9	be_ANT4-Ia_118	aminoglycoside resistance
10	be_ANT4-Ia_197	aminoglycoside resistance
11	be_ANT6-Ia_433	aminoglycoside resistance
12	be_ANT6-Ia_576	aminoglycoside resistance
13	be_ant(6')-Ib_179	aminoglycoside resistance
14	be_ant(6')-Ib_435	aminoglycoside resistance
15	be_ANT9-Ia_278	aminoglycoside resistance
16	be_ANT9-Ia_560	aminoglycoside resistance
17	be_APH2-Ia_149	aminoglycoside resistance
18	be_APH2-Ia_292	aminoglycoside resistance
19	be_APH2-Ib_317	aminoglycoside resistance
20	be_APH2-Ib_737	aminoglycoside resistance
21	be_APH2-Ic_346	aminoglycoside resistance
22	be_APH2-Ic_58	aminoglycoside resistance
23	be_APH2-Id_249	aminoglycoside resistance
24	be_APH2-Id_354	aminoglycoside resistance
25	VPAB1_aph2-Ie	aminoglycoside resistance
26	VPAB2_aph2-Ie	aminoglycoside resistance
27	be_APH3-III_136	aminoglycoside resistance
28	be_APH3-III_332	aminoglycoside resistance
29	be_APH3-IVa_20	aminoglycoside resistance
30	be_APH3-IVa_474	aminoglycoside resistance
31	be_aadK_175	aminoglycoside resistance
32	be_aadK_61	aminoglycoside resistance
33	VPAB5_aad9	beta-lactam resistance
34	VPAB6_aad9	beta-lactam resistance
35	be_bla1_201	beta-lactam resistance
36	be_bla1_366	beta-lactam resistance
37	be_bla2_192	beta-lactam resistance
38	be_bla2_246	beta-lactam resistance
39	be_blaZ_718	beta-lactam resistance
40	be_blaZ_811	beta-lactam resistance
41	VPAB55_blaZ_663	beta-lactam resistance
42	be_bleO_18	Bleomycin resistance
43	be_bleO_321	Bleomycin resistance
44	be_cat-86_367	chloramphenicol resistance
45	be_cat-86_605	chloramphenicol resistance
46	be_cat-Bant_402	chloramphenicol resistance
47	be_cat-Bant_613	chloramphenicol resistance

48	VP_cat-pC223_135	chloramphenicol resistance
49	VP_cat-pC223_413	chloramphenicol resistance
50	VP_cat-pC223_561	chloramphenicol resistance
51	be_cat-TC_set_170	chloramphenicol resistance
52	be_cat-TC_set_370	chloramphenicol resistance
53	be_catB_233	chloramphenicol resistance
54	be_catB_27	chloramphenicol resistance
55	be_catDP_set_267	chloramphenicol resistance
56	be_catDP_set_281	chloramphenicol resistance
57	be_catQ_186	chloramphenicol resistance
58	be_catQ_66	chloramphenicol resistance
59	be_catS_224	chloramphenicol resistance
60	be_catS_265	chloramphenicol resistance
61	VP_catpC221_411	chloramphenicol resistance
62	be_catpC221_561	chloramphenicol resistance
63	be_cfr_466	chloramphenicol, florfenicol, clindamycin, linezolid resistance
64	be_cfr_908	chloramphenicol, florfenicol, clindamycin, linezolid resistance
65	be_dfrA_172	trimethoprim resistance
66	be_dfrA_20	trimethoprim resistance
67	VP_dfrG_140	trimethoprim resistance
68	VP_dfrG_26	trimethoprim resistance
69	be_dfrD-Lmon_140	trimethoprim resistance
70	be_dfrD-Lmon_27	trimethoprim resistance
71	be_dfrK_19	trimethoprim resistance
72	be_dfrK_241	trimethoprim resistance
73	be_ereA_108	macrolide,lincosamide resistance
74	be_ereA_791	macrolide,lincosamide resistance
75	be_ereB_639	macrolide,lincosamide resistance
76	be_ereB_1209	macrolide,lincosamide resistance
77	be_ermA_193	macrolide,lincosamide resistance
78	be_ermA_590	macrolide,lincosamide resistance
79	be_ermB_112	macrolide,lincosamide resistance
80	be_ermB_520	macrolide,lincosamide resistance
81	be_ermC_149	macrolide,lincosamide resistance
82	be_ermC_372	macrolide,lincosamide resistance
83	be_ermDJK_289	macrolide,lincosamide resistance
84	be_ermDJK_841	macrolide,lincosamide resistance
85	be_ermF_231	macrolide,lincosamide resistance
86	be_ermF_494	macrolide,lincosamide resistance
87	be_ermG_296	macrolide,lincosamide resistance
88	be_ermG_98	macrolide,lincosamide resistance
89	be_ermQ_398	macrolide,lincosamide resistance
90	be_ermQ_521	macrolide,lincosamide resistance
91	be_ermT_104	macrolide,lincosamide resistance
92	be_ermT_149	macrolide,lincosamide resistance
93	be_ermX_231	macrolide,lincosamide resistance
94	be_ermX_282	macrolide,lincosamide resistance
95	be_ermY_122	macrolide,lincosamide resistance
96	be_ermY_258	macrolide,lincosamide resistance
97	be_erm33_368	macrolide,lincosamide resistance

98	be_erm33_419	macrolide, lincosamide resistance
99	be_erm34_166	macrolide, lincosamide resistance
100	be_erm34_582	macrolide, lincosamide resistance
101	be_erm36_245	macrolide, lincosamide resistance
102	be_erm36_644	macrolide, lincosamide resistance
103	be_fexA_541	macrolide, lincosamide resistance
104	be_fexA_1022	macrolide, lincosamide resistance
105	be_fusA_100	fusidic acid resistance
106	be_fusA_1359	fusidic acid resistance
107	be_fusB_337	fusidic acid resistance
108	be_fusB_453	fusidic acid resistance
109	be_fusC_247	fusidic acid resistance
110	be_fusC_406	fusidic acid resistance
111	be_fusD_329	fusidic acid resistance
112	be_fusD_510	fusidic acid resistance
113	be_lnuA_115	lincosamide resistance
114	be_lnuA_218	lincosamide resistance
115	be_lnuB_169	lincosamide resistance
116	be_lnuB_646	lincosamide resistance
117	VPAB11_lnuC	lincosamide resistance
118	VPAB12_lnuC	lincosamide resistance
119	be_lnuD_98	lincosamide resistance
120	be_lnuD_304	lincosamide resistance
121	be_lsaB_375	lincosamide resistance
122	be_lsaB_1003	lincosamide resistance
123	be_mdtA_355	macrolide, tetracycline resistance
124	be_mdtA_571	macrolide, tetracycline resistance
125	be_mecA_1042	beta-lactam resistance
126	be_mecA_871	beta-lactam resistance
127	VPAB53_mecA_1535	beta-lactam resistance
128	VPAB54_mecA_1583	beta-lactam resistance
129	be_mef_set_193	macrolide resistance
130	be_mef_set_39	macrolide resistance
131	VPAB9_mel	macrolide resistance
132	VPAB10_mel	macrolide resistance
133	VPAB31_merB	macrolide resistance
134	VPAB32_merB	macrolide resistance
135	VPAB51_mphC_281	macrolide resistance
136	VPAB52_mphC_527	macrolide resistance
137	be_mreA_420	macrolide resistance
138	be_mreA_78	macrolide resistance
139	be_msrC_1279	macrolide resistance
140	be_msrC_591	macrolide resistance
141	be_msr_set_289	macrolide resistance
142	be_msr_set_655	macrolide resistance
143	VPAB7_mupR	mupirocin resistance
144	VPAB8_mupR	mupirocin resistance
145	be_norA_1027	norfloxacin resistance
146	be_norA_427	norfloxacin resistance
147	be_sat4_161	streptothricin resistance

148	be_sat4_338	streptothricin resistance
149	be_str_334	streptomycin resistance
150	be_str_480	streptomycin resistance
151	be_tet36_116	tetracycline resistance
152	be_tet36_221	tetracycline resistance
153	VPAB29_tet38	tetracycline resistance
154	VPAB30_tet38	tetracycline resistance
155	be_tet44_171	tetracycline resistance
156	be_tet44_1433	tetracycline resistance
157	be_tetAP_244	tetracycline resistance
158	be_tetAP_290	tetracycline resistance
159	be_tetBP_1211	tetracycline resistance
160	be_tetBP_1538	tetracycline resistance
161	be_tetK_259	tetracycline resistance
162	be_tetK_351	tetracycline resistance
163	be_tetL_1_151	tetracycline resistance
164	be_tetL_1_676	tetracycline resistance
165	be_tetL_2_269	tetracycline resistance
166	be_tetL_2_504	tetracycline resistance
167	be_tetM_1378	tetracycline resistance
168	be_tetM_81	tetracycline resistance
169	be_tetO_571	tetracycline resistance
170	be_tetO_915	tetracycline resistance
171	VPAB13_tetO320	tetracycline resistance
172	VPAB14_tetO320	tetracycline resistance
173	VPAB15_tetW032WO	tetracycline resistance
174	VPAB16_tetW032WO	tetracycline resistance
175	be_tetS_18	tetracycline resistance
176	be_tetS_776	tetracycline resistance
177	be_tetT_1326	tetracycline resistance
178	be_tetT_232	tetracycline resistance
179	be_tetU_133	tetracycline resistance
180	be_tetU_191	tetracycline resistance
181	be_tetW_455	tetracycline resistance
182	be_tetW_66	tetracycline resistance
183	be_tetZ_43	tetracycline resistance
184	be_tetZ_93	tetracycline resistance
185	be_vanA_192	vancomycin resistance
186	be_vanA_884	vancomycin resistance
187	be_vanB_set_151	vancomycin resistance
188	be_vanB_set_65	vancomycin resistance
189	be_vanC-1_497	vancomycin resistance
190	be_vanC-1_77	vancomycin resistance
191	be_vanC_set_184	vancomycin resistance
192	be_vanC_set_37	vancomycin resistance
193	be_vanD1-2-3_388	vancomycin resistance
194	be_vanD1-2-3_532	vancomycin resistance
195	be_vanD4-5_183	vancomycin resistance
196	be_vanD4-5_267	vancomycin resistance
197	VPAB17_vanE	vancomycin resistance

198	VPAB18_vanE	vancomycin resistance
199	be_vanG_362	vancomycin resistance
200	be_vanG_549	vancomycin resistance
201	be_vanZ_328	vancomycin resistance
202	be_vanZ_455	vancomycin resistance
203	be_vatA_288	streptogramin A resistance
204	be_vatA_429	streptogramin A resistance
205	be_vatB_109	streptogramin A resistance
206	be_vatB_9	streptogramin A resistance
207	be_vatC_474	streptogramin A resistance
208	be_vatC_552	streptogramin A resistance
209	be_vatD_234	streptogramin A resistance
210	be_vatD_282	streptogramin A resistance
211	VPAB19_vatE	streptogramin A resistance
212	VPAB20_vatE	streptogramin A resistance
213	be_vatG_19	streptogramin A resistance
214	be_vatG_301	streptogramin A resistance
215	be_vgaA_1389	streptogramin B resistance
216	be_vgaA_195	streptogramin B resistance
217	be_vgaAv_1401	streptogramin B resistance
218	be_vgaAv_174	streptogramin B resistance
219	be_vgaB_569	streptogramin B resistance
220	be_vgaB_649	streptogramin B resistance
221	be_vgaC_160	streptogramin B resistance
222	be_vgaC_860	streptogramin B resistance
223	be_vgaD_92	streptogramin B resistance
224	be_vgaD_800	streptogramin B resistance
225	be_vgaE_169	streptogramin B resistance
226	be_vgaE_834	streptogramin B resistance
227	be_vgbA_142	streptogramin B resistance
228	be_vgbA_281	streptogramin B resistance
229	be_vgbB_273	streptogramin B resistance
230	be_vgbB_444	streptogramin B resistance
231	be_lukPV_156	Penton-Valentine virulence gene
232	be_lukPV_1691	Penton-Valentine virulence gene
233	VPAB27_lukS-Si	leukocidin virulence gene
234	VPAB28_lukS-Si	leukocidin virulence gene
235	VPAB41_16S_Sther	positive control
236	VPAB42_16S_Sther	positive control
237	VPAB43_16S_Llactis	positive control
238	VPAB44_16S_Llactis	positive control
239	VPAB45_16S_Ljohn	positive control
240	VPAB46_16S_Ljohn	positive control
241	VPAB47_16S_Blactis	positive control
242	VPAB48_16S_Blactis	positive control
243	VPAB49_16S_ctrl1	positive control
244	VPAB50_16S_ctrl2	positive control
245		
246	Biotin-Marke_2,5µM	

Protocol

Thu Oct 27 10:10:27 2011

Experimentator: Yvonne

M47-10

Labelling 25.10.2011

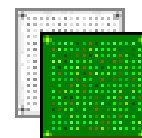
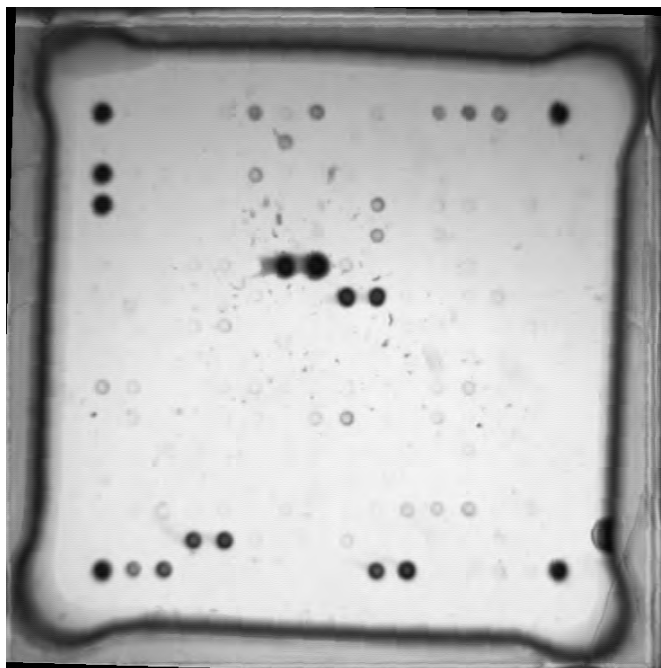
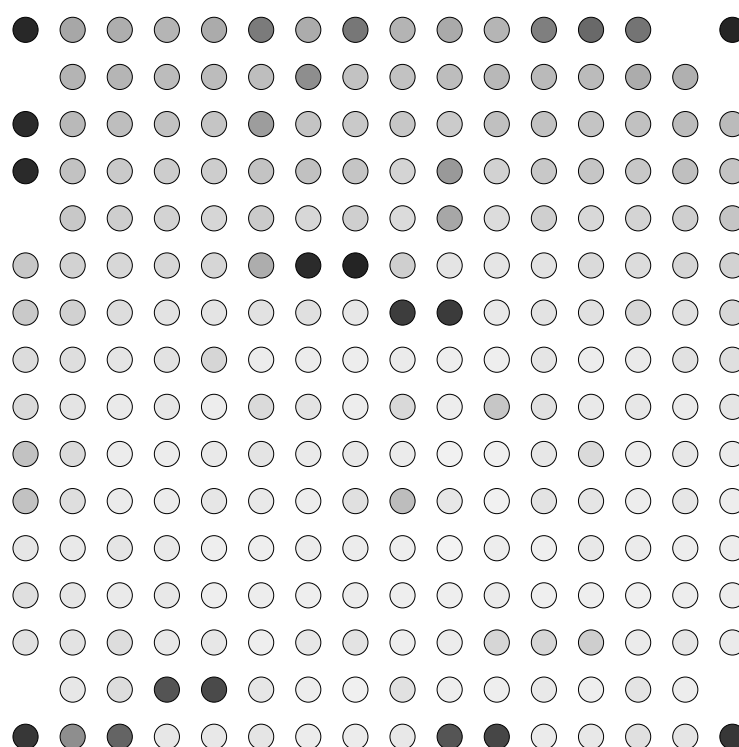


Image M47-10.bmp:

**Results Mean:**

Protocol

Fri Oct 07 10:09:56 2011
Experimentator: Yvonne
M1186-10
Labelling 05.10.2011

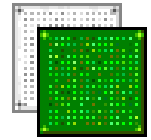
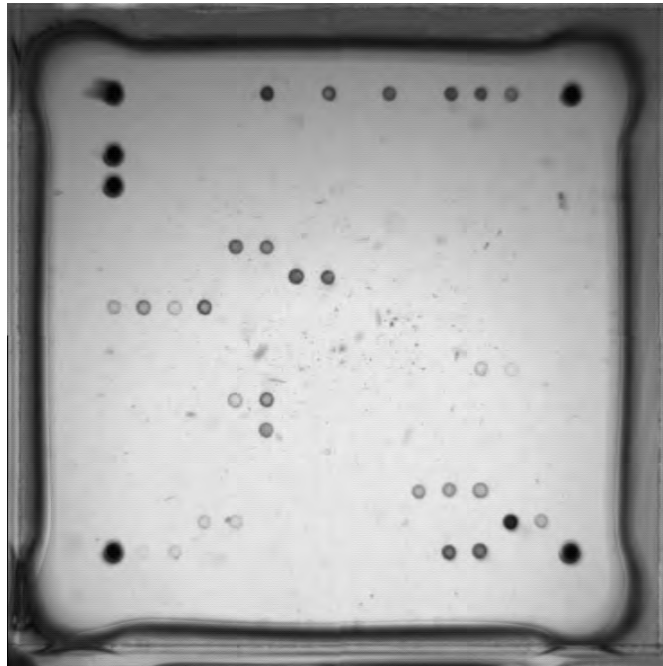
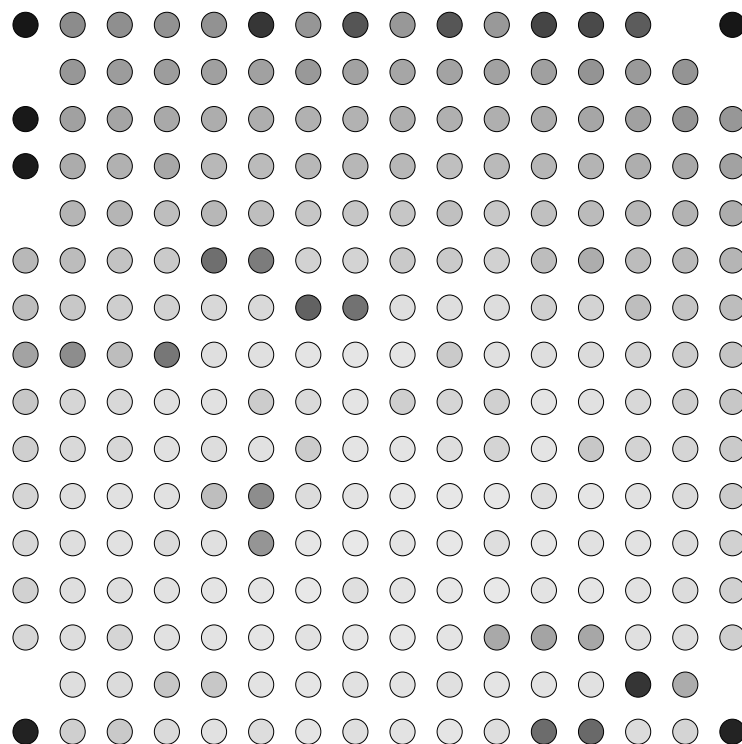


Image M1186-10.bmp:

**Results Mean:**

Protocol

Tue Jan 17 16:56:00 2012
Experimentator: Yvonne Frey
M1383-10n
2ter Versuch
Labelling 10.01.2012

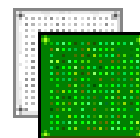
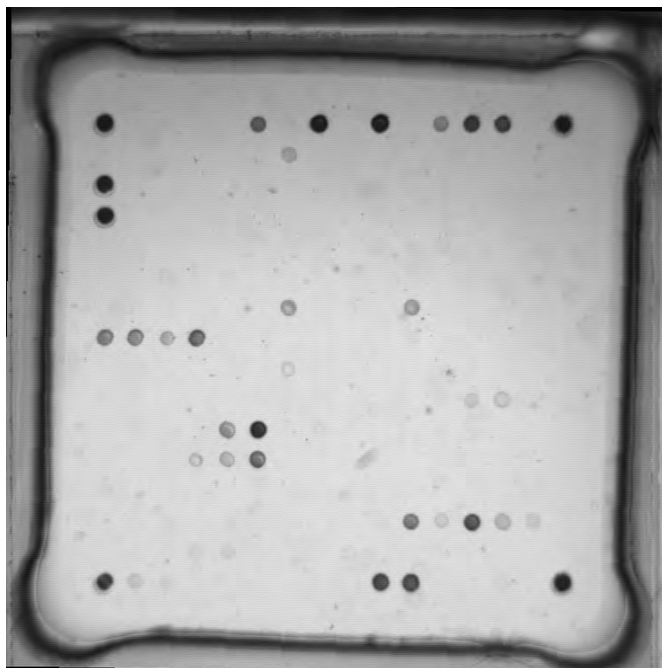
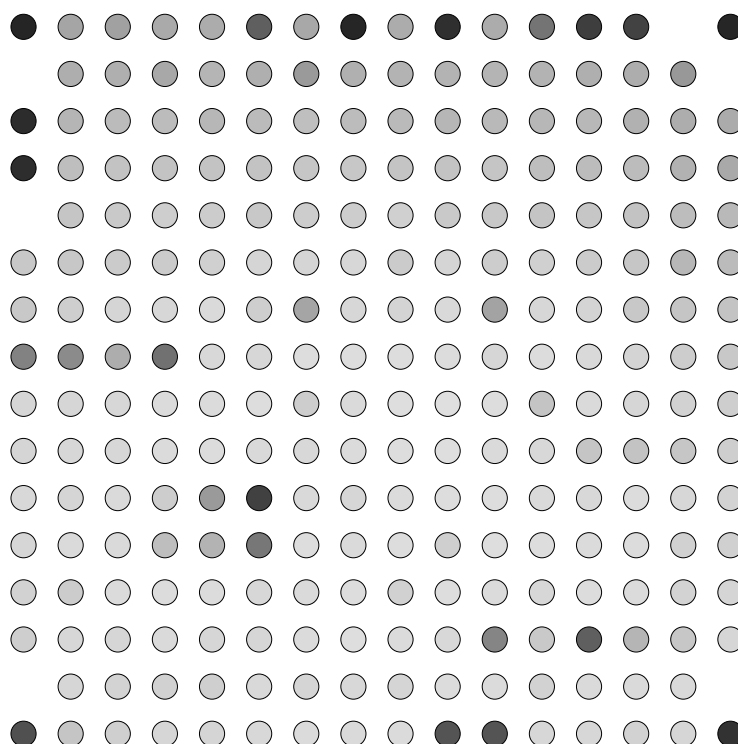


Image M1383-10n.bmp:

**Results Mean:**

Protocol

Fri Oct 21 16:51:55 2011
Experimentator: YVONNE
M1529.10
LABLING 19.10.2011

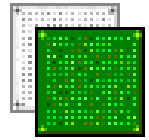
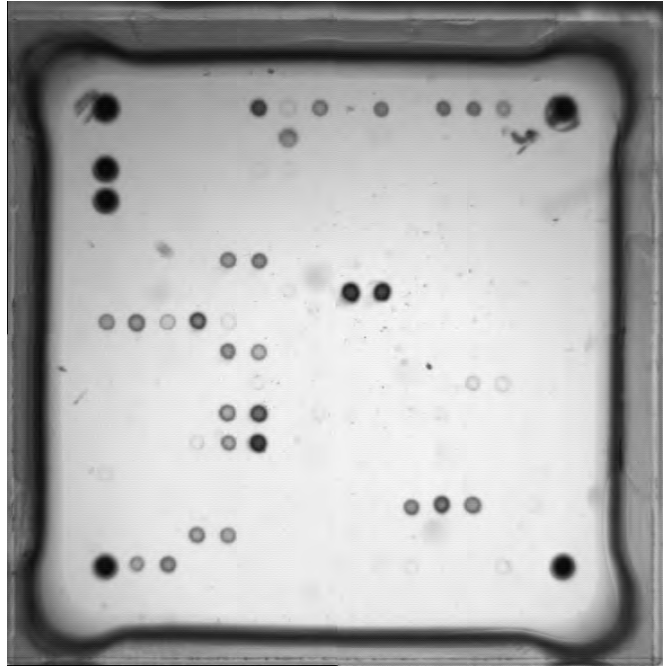
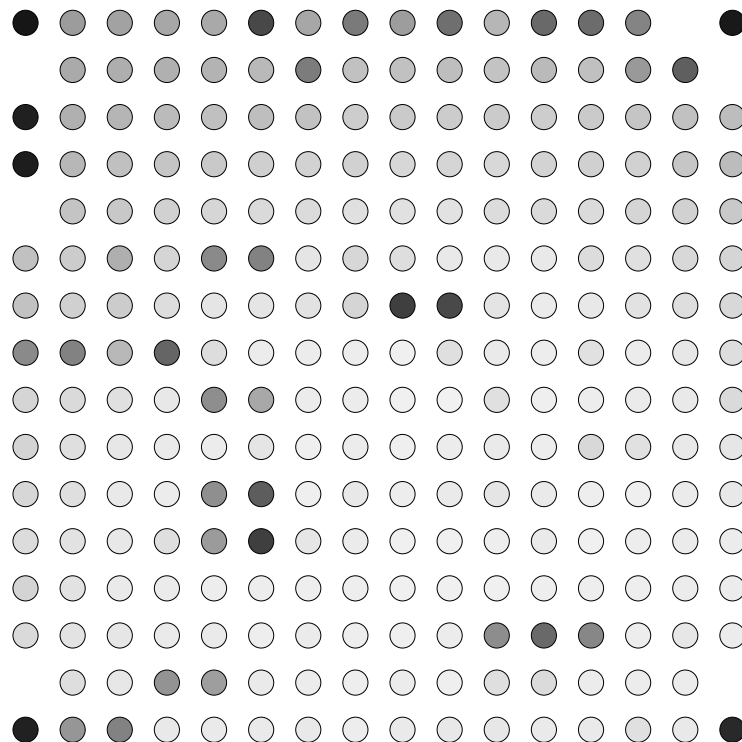


Image M1529.10.bmp:

**Results Mean:**

Protocol

Fri Oct 07 10:26:15 2011
Experimentator: Yvonne
M1570-10
Labelling 05.10.2011

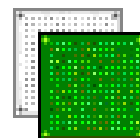
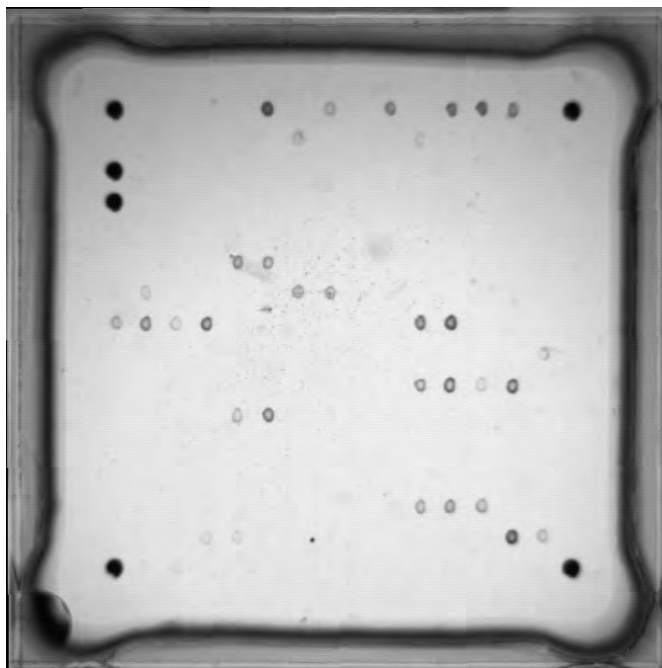
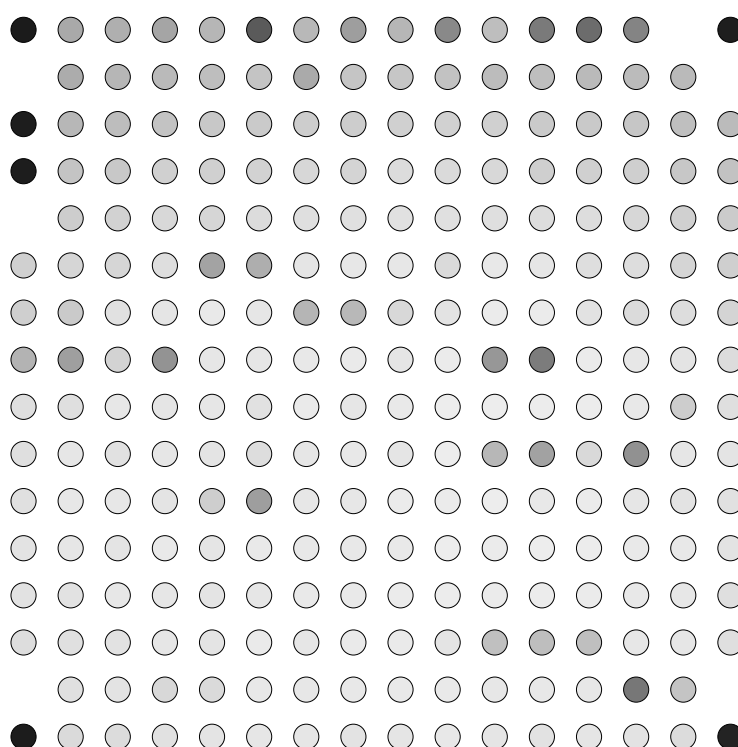
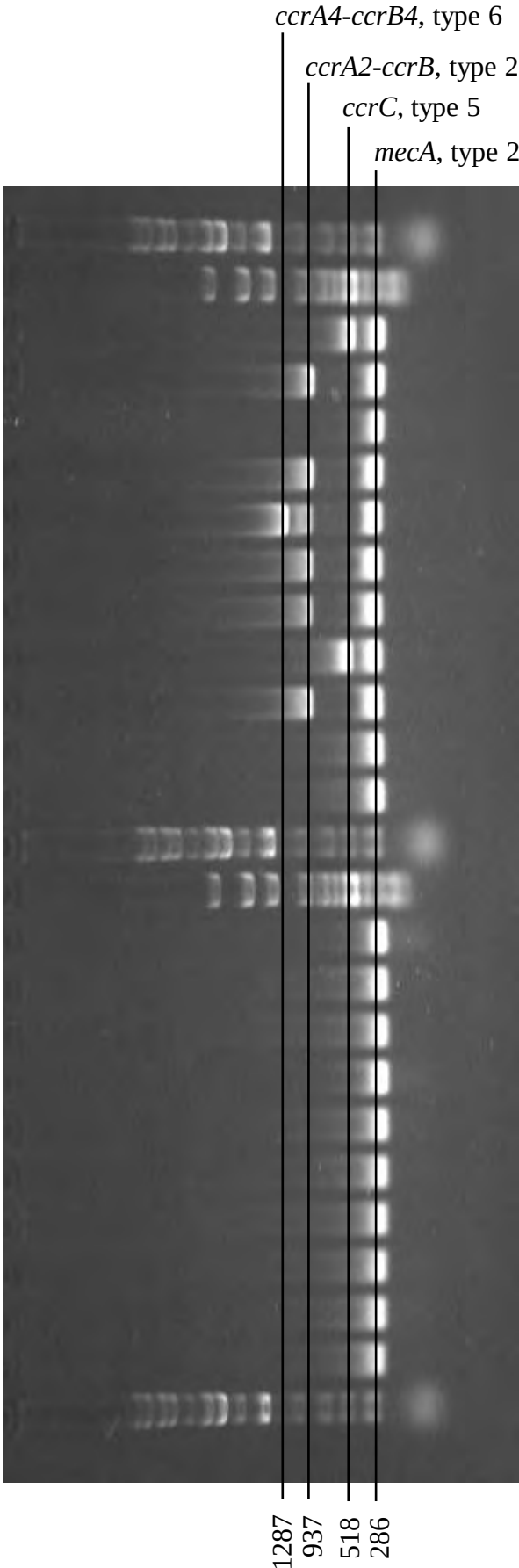


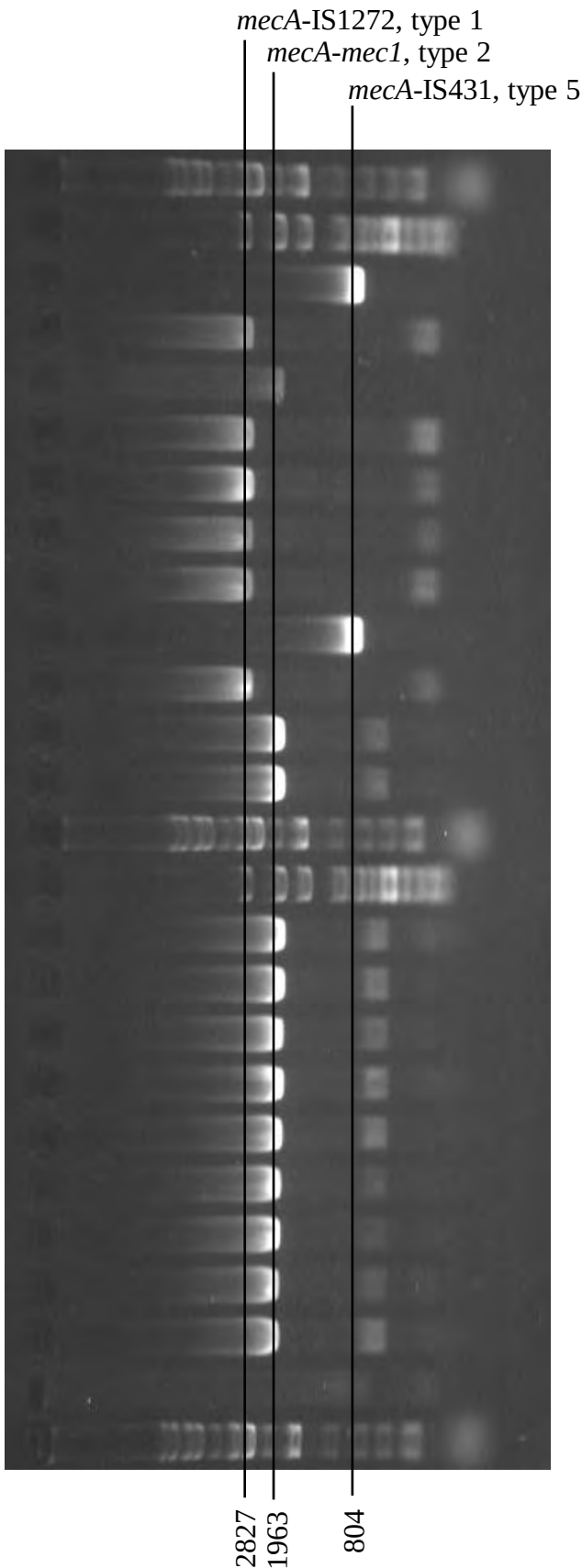
Image M1570-10.bmp:

**Results Mean:**

SCCmec typing of CNS as determined by PCR using method of Kondo et al., 2007. AAC
51(1): 264-74
Kondo – PCR 1, 18.05.2012



Kondo – PCR 2, 18.05.2012



DNA Ladder 1kb

DNA Ladder 10b

control 3: IMD 1532/11

control 2: IMD 1530/11

control 1: IMD 1528/11

M1529/10 *S. epidermidis*

M1383/10 *S. epidermidis*

M1186/10 *S. epidermidis*

M744/10 *S. epidermidis*

M703/10 *S. epidermidis*

M8/10 *S. epidermidis*

M4276-1/09 *S. fleurettii*

M3783/09 *S. fleurettii*

DNA Ladder 1kb

DNA Ladder 10b

M1961/10 *S. fleurettii*

M1191-2/10 *S. fleurettii*

M1125-2/10 *S. fleurettii*

M545/10 *S. fleurettii*

M404-1/10 *S. fleurettii*

M277/10 *S. fleurettii*

M205/10 *S. fleurettii*

M143/10 *S. fleurettii*

M4460/09 *S. fleurettii*

M1570/10 *S. haemolyticus*

DNA Ladder 1kb

Kondo – PCR 1 and 2, 15.06.2012

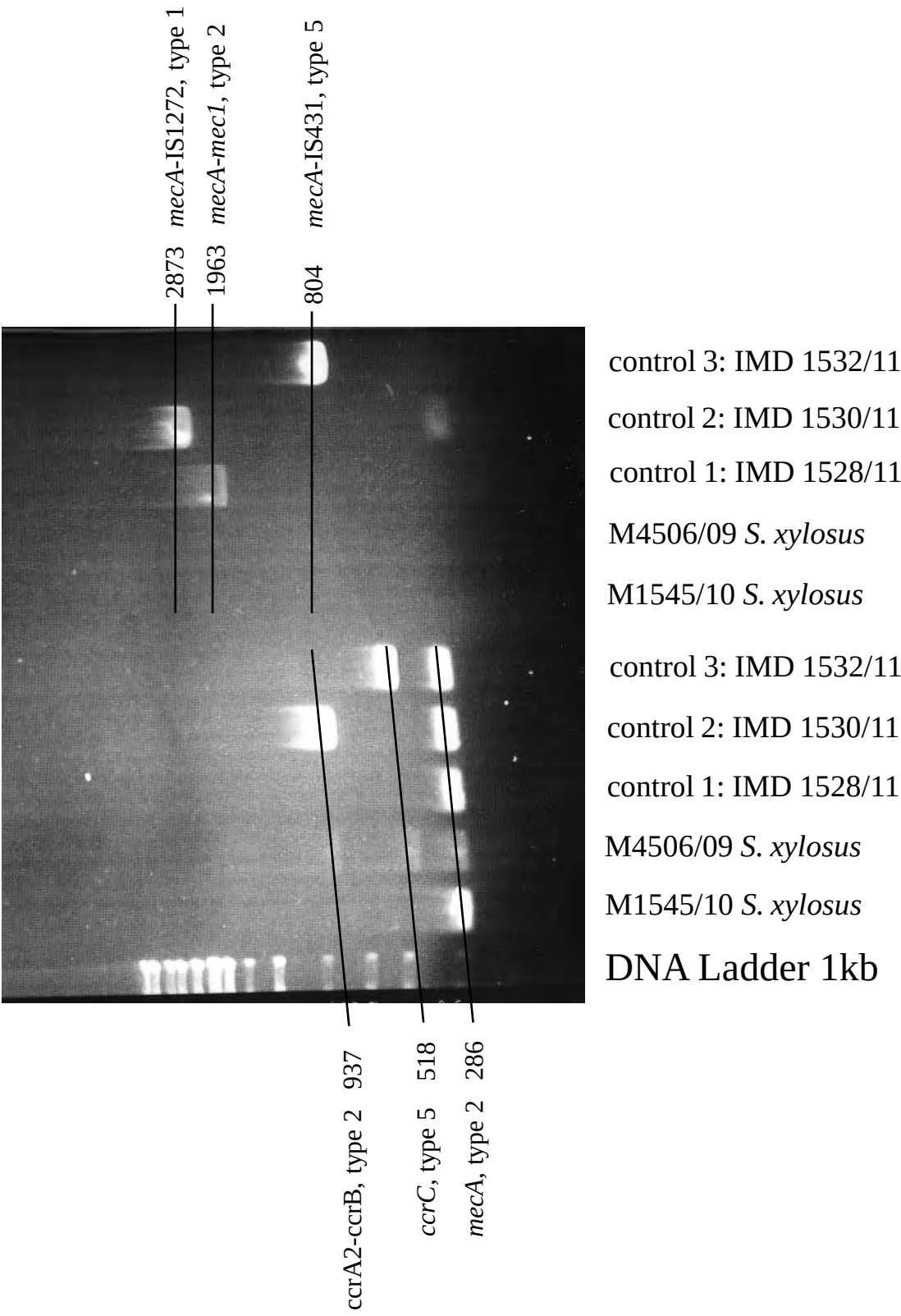


Table. MLST results of 13 (multi-)resistant *S. epidermidis* strains

isolates	species	arcC	aroE	gtr	mutS	pyr	tpiA	yqiL	ST (= sequence type)
M8/10	<i>S. epidermidis</i>	1	2	1	1	2	1	10	ST55
M523/10	<i>S. epidermidis</i>	1	13	2	2	2	1	7	ST111
M703/10	<i>S. epidermidis</i>	1	1	2	1	2	1	8	ST454
M744/10	<i>S. epidermidis</i>	2	1	1	1	2	1	1	ST59
M864/10	<i>S. epidermidis</i>	16	1	2	1	2	1	1	ST184
M1186/10	<i>S. epidermidis</i>	1	1	2	1	2	1	1	ST89
M1383/10	<i>S. epidermidis</i>	2	1	1	2	2	16	1	ST452
M1529/10	<i>S. epidermidis</i>	2	1	1	1	2	1	1	ST59
M1662/10	<i>S. epidermidis</i>	1	13	2	2	2	1	7	ST111
M1725/10	<i>S. epidermidis</i>	1	1	39	36	2	1	1	ST453
M1977/10	<i>S. epidermidis</i>	1	13	2	2	2	1	7	ST111
M4233-1/09	<i>S. epidermidis</i>	1	1	2	2	2*	1	1	New type
M4298-1/09	<i>S. epidermidis</i>	1	13	2	2	2	1	7	ST111

Pulsed Field Gel Electrophoresis (PFGE) for Staphylococci

Part 1: Plugs Preparation

- make an overnight culture of the strain on a blood agar plate
- weight an empty 1.5ml Eppendorff tube
- suspend the culture in $\approx 500\mu\text{l}$ **TE-Buffer** (Tris 10mM, EDTA 100mM, pH 8.0) in this 1.5ml Eppendorff tube
- centrifuge the suspension at 14'000 rpm, 4min, through away the supernatant and dry the cell pellet
- weight the dried cell pellet and resuspend them according to the following formula in x μl **TE-Buffer**:

$$x \mu\text{l} = \frac{\text{weight of the cell pellet [mg]} \times 300 [\mu\text{l}]}{10 [\text{mg}]}$$

- prepare an 1.5% agarose gel (**Seakem Gold agarose**): 0.3 g agarose + 20ml **0,5x TBE** and put it in a waterbath at 50°C
- pipette 200-300 μl gel + 200-300 μl cell suspension, mix well and mould in the plug (BioRad)
- leave in ice to solidify
- create several plugs (3-4) of each sample
- incubate the plugs in 1.2ml **TE Buffer**, pH 8.0 + 5U (= 3 μl) **Lysostaphine** at 37°C for 5-6 hours
- transfer the plugs in 2ml Eppendorff tubes, incubate overnight with **EDTA 0.5M**, 1% **N-Lauroylsarcosine**, 2mg/ml **Proteinase K**, pH 8.0 at 50°C
- transfer the plugs in a little petri dish and wash 5x 30min. with **TE Buffer** at room temperature
- store at 4°C in **EDTA 0.5M**, pH 8.0 for many months or in **TE Buffer** for many days

Part 2: Digestion

- (if the plugs were conserved in **EDTA 0.5M**, equilibrate in **TE Buffer**)
- equilibrate the plugs in **restriction buffer A** for 30 min at 4°C (360 μl H₂O + 40 μl restriction buffer A)
- digest the DNA for 5 hours with 50U **restriction enzyme (SmaI)** in 300 μl at 25°C (265 μl H₂O + 30 μl restriction buffer A + 5 μl enzyme (SmaI))

Part 3: Electrophoresis

- make an 1% gel (1g **Bio Red agarose** in 100ml **0,5x TBE**)
- place plugs in slots and seal the gel (1/4 of a plug) with **low melting agarose**
- equilibrate the buffer ($\approx 2\text{L}$ **TBE 0.5x** + 9mg/l **Thioruea**) in the machine at 12°C
- after migration, stain gel 45min. with 1U/ μl **EtBr** ($\approx 20 \mu\text{l}$ EtBr in 100ml H₂O)
- rinse 2x 20min with **H₂O**
- Electrophoresis conditions for *S. intermedius*, *S. aureus*:
SmaI: 6V/cm, Temperature: 12°C, running time: 21h; Start: 5sec., End: 40sec.

Primers used for this thesis about Antimicrobial resistance in CNS from bovine milk

Primer name	Sequence (5' –3')	Primer design, reference or source
aac(6')-aph(2')-F	CAGAGCCCTTGGGAAGATGAAG	Vakulenko et al., 2003. AAC 47:1423-1426
aac(6')-aph(2')-R	CCTCGTGTAATTCATGTTCTGGC	
AIPS 27	CTAACACTGAACCCCAATG	ACME-arcA ; Diep et al. 2008. JID, 197 : 1523-1530
AIPS 28	GAGCCAGAAGTACGCGAG	
AIPS 45	GCAAACTCTGTAAATGGTCTGTTC	ACME-opp3AB; Diep et al. 2008. JID, 197 : 1523-1530
AIPS 46	GAAGATTGGCAGCACAAAGTG	
ant6-I-F2	CAAGCGCAAGGGAGTATG	
ant6-I-R1	CTCTTCTATATCAGCGGC	
aph3-III-F	CCGCTGCGTAAAGATAC	Perreten et al. 2005. J. Clin. Microbiol. 43: 2291-2302
aph3-III-R	GTCATACCACTTGTCGC	
arcA-F	GAGCCAGAAGTACGCGAG	Barbier et al. 2011. JAC 66 :29-36
arcA-R	CACGTAACTTGCTAGAACGAG	
blaZ-F	CAGTTCACATGCCAAAGAG	Perreten et al. 2005. J. Clin.
blaZ-R1	CAGCAGGTGTGAAGTATC	Riesen et al. 2010. Int. J. Syst. Evol. Microbiol.
catpC221-F1	CTGAATCTGATAACAACTTCACT	Institute of Veterinary Bacteriology, Bern, redesigned by VP & ChS
catpC221-R	TGAAGCATGGTAACCATCAC	Schnellmann et al. 2006. J. Clin Microbiol 44:4444-4454
catpC223-F2	AATAGTGAGGGGAATTG	Institute of Veterinary Bacteriology, Bern, redesigned by VP & ChS
catpC223-R	ACATGGTAACCATCACATAC	Schnellmann et al. 2006. J. Clin Microbiol 44:4444-4454
dfrA-F	CCTTGGCACTTACCAAATG	Perreten et al. 2005. J. Clin. Microbiol. 43: 2291-2302
dfrA-R	CTGAAGATTCGACTTCCC	
dfrA-IS431	CCATGCCACGAAATTAGCAT	for acquired <i>dfr</i> (A) in <i>S. epidermidis</i> , use with dfrA-F
dfrD-F	TTCTTTAATTGTTGCGATGG	
dfrD-R	TTAACGAAATTCTCTCATATATATG	
dfrG-F	ATGAAAGTTTCTTTGATTGCTGC	
dfrG-R	CCCTTTTGGGCAATAACC	
dfrK-F	ATGAAAGTTTCTTTAATTGCTGCG	
dfrK-R	TCAAGAAGCTTTTCGCTCATAA	
erm(33)-F	GAAATYGGCTCAGGAAAGG	
erm(A)-R	TTAGTGAAACAATTGTAACTATTG	Institute of Veterinary Bacteriology, Bern
erm(F)-F	CGGGTCAGCACTTACTATTG	Institute of Veterinary Bacteriology, Bern
erm(F)-R	GGACCTACCTCATAGACAAG	Institute of Veterinary Bacteriology, Bern
erm(G)-F	CATTTCCTAGCCACAATCCAT	Institute of Veterinary Bacteriology, Bern

erm(G)-R	TTAAATAGCGATACAAATTGTTG	Institute of Veterinary Bacteriology, Bern
erm(Q)F	GAGGAAAAGTTGATATTAGTG	
erm(Q)R	GACGCAATCTACACTAGG	
ermX-F	ACCACAAAGATCATCAACTC	
ermX-R	GGAAGAGATCGATCCAGT	
icA-P5	AAGATGTTGGCTGTGATTAC	Gu et al. 2005. J. Hosp. Infection: 61: 342-348
icA-P3	CAACAAGTTGAAGGCATATC	
lnuA-1	GGTGGCTGGGGGTAGATGATTAACTGG	
lnuA-2	GCTTCTTTTGAATAACATGGTATTTTCGATC	
mecA-1	AAAATCGATGGTAAAGTTGGC	Louie et al. 2002. J. Clin. Microbiol. 40: 2786-2790
mecA-2	AGTTCTGCAGTACCGGATTTC	
mecA-F	ATGAAAAAGATAAAAATGTTCAC	Schnellmann et al. 2006. J. Clin. Microbiol 44:4444-4454
mecA-R	TTATTCACTATATCGTATTTTATTAC	
mecA-F1	GATGATACMTTCGTTCAC	
mecA-R1	CGAGTGCTACTCTAGCAA	
mecA-R2	TCCTGTATTGGCCAAATTC	
mecA-F4	TACTGATTAAACCCAGTAC	
mecA-P4	TCCAGATTACAACCTCACCCAGG	Oliveira and Milheirico 2007 mecA
mecA-F5	CATTGATCGCAACGTTCAATTT	Francois et al. 2005. J Clin
mecA-F6	ATGGCAAAGATATTCAACTAAC	This study
mecA-R6	ATTGTAATTGCTATTATCGTCAAC	This study
mecA-F7	GATAACACCTGCTACAC	This study
mecA-R7	AAGGAGAAGTAAACAGC	This study
mecAv-F	AAAAGATAAAATCTTGGGGTG	Anneals in mecA variants of CNS
mecAv-R	CCTTGTTTCATYTTGAGTTC	
mecA LA251-F	CAGCCAGATTCAATTGTACC	
mecA LA251-R	AACATCGTACGATGGGGTAC	
mecA1-sc-F	ATTAATCATCGCCATCGTGA	This study
mecA1-sc-R	TTTGTATCTTGATTCATATTTGAACA	This study
mecAscK1-F	CATATATATATTATACGCTCATC	This study
mecAscK3-F	GATTTAATCAAGACTTGCAATTC	This study
mecAsc-R	TTCAATGGCATCAATTGTTTC	This study
mphC-F	CATTGAATGAATCGGGAC	Perreten et al. 2005. J. Clin. Microbiol. 43: 2291-2302
mphC-R	TTCATACGCCGATTCTCC	
strpS194-F	TATTGCTCTCGAGGGTTC	Schnellmann et al. 2006. J. Clin Microbiol 44:4444-4454

strpS194-R	CTTTCTATATCCATTTCATCTC	Perreten et al. 2005. J. Clin. Microbiol. 43: 2291-2302
tet(K)-1	TTAGGTGAAGGTTAGGTCC	
tet(K)-2	GCAAACTCATTCAGAAAGCA	
Kondo multiplex PCR 1:		
mA1	TGCTATCCACCCTCAAAACAGG	Type II.1 <i>mecA</i> (mA1-mA2) Kondo et al. 2007
mA2	AACGTTGTAACCAACCCCAAGA	Type II.1 Kondo et al. 2007
α1	AACCTATATCATCAATCAGTACGT	Type I.1 <i>ccrA1-ccrB</i> Kondo et al. 2007
α2	TAAAGGCATCAATGCACAAACACT	Type II.1 <i>ccrA2-ccrB</i> Kondo et al. 2007
α3	AGCTCAAAAGCAAGCAATAGAAAT	III.1 <i>ccrA3-ccrB</i> Kondo et al. 2007
βc	ATTGCCCTTGATAAATAGCCITCT	<i>ccrB1, ccrB2,ccrB3</i> Type I.1, II.1, III.1 Kondo et al. 2007
α4.2	GTATCAATGCACCAGAACTT	Type VI <i>ccrA4-ccrB4</i> Kondo et al. 2007
β4.2	TTGCGACTCTCTTGCGGTTT	Type VI Kondo et al. 2007
γR	CCTTTATAGACTGGATTATTCAAAATAT	type V <i>ccrC</i> Kondo et al. 2007
γF	CGTCTATTACAAGATGTTAAGGATAAT	type V Kondo et al. 2007
Kondo multiplex PCR 2:		
mI6	CATAACTTCCCATTCTGCAGATG	Type II.1 <i>mecA-mecI</i> Kondo et al. 2007
IS7	ATGCTTAATGATAGCATCCGAATG	Type I.1 <i>mecA-IS1272</i> upstream of <i>mecA</i> Kondo et al. 2007
IS2 (iS-2)	TGAGGTTATTCAGATATTTTCGATGT	Type V <i>mecA-IS431</i> upstream of <i>mecA</i> Kondo et al. 2007
mA7	ATATACCAAAACCCGACAACTACA	Type I.1, II.1 <i>mecA</i> Kondo et al. 2007
MLST – Primer:		
arcC-F	TGTGATGAGCACGCTACCGTTAG	Thomas et al. 2007 JMC.01934-06: 616-619
arcC-R	TCCAAGTAAACCCATCGGTCTG	
aroE-F	CATTGGATTACCTCTTTTGTTCAGC	Thomas et al. 2007 JMC.01934-06: 616-619
aroE-R	CAAGCGAAATCTGTTGGGG	
gtr-F	CAGCCAAATCTTTTATGACTTTT	Thomas et al. 2007 JMC.01934-06: 616-619
gtr-R	GTGATTAAAGGTATTGATTTGAAT	
gtr-R1	GAAACTGAAGTGATTAAAGG	This study
mutS-F3	GATAAAGAATAAGGGTTGTGAA	Thomas et al. 2007 JMC.01934-06: 616-619
mutS-R3	GTAATCGTCTCAGTTATCATGTT	
pyr-F2	GTTACTAATACTTTTGCTGTGTTT	Thomas et al. 2007 JMC.01934-06: 616-619
pyr-R4	GTAGAAATGTAAAAGAGACTAAAATGAA	
tpi-F2	ATCCAATTAGACGCTTTAGTAAC	Thomas et al. 2007 JMC.01934-06: 616-619
tpi-R2	TTAATGATGCGCCACCTACA	
yqil-F2	CACGCATAGTATTAGCTGAAG	Thomas et al. 2007 JMC.01934-06: 616-619
yqil-R2	CTAATGCCCTTCATCTTGAGAAATAA	

Sequence alignment of methicillin resistance gene *mecA* in *S. epidermidis* isolates

using MultAlign Program <http://multalin.toulouse.inra.fr/multalin/>

Reference strains:

- *S. aureus* N315
- *S. epidermidis* RPA62A

S. aurN315	1	ATGAAAAAGA	TAAAAATTGT	TCCACTTATT	TAAATAGTTG	TAGTTGTCGG	GTTTGGTATA	TATTTTATG	CTTCCAAGA	TAAAGAAATT	AATAATAC	TTGATGCAAT	TGAAGATAAA	AATTTCAAA	130
	S. epirP62A	ATGAAAAAGA	TAAAAATTGT	TCCACTTATT	TAAATAGTTG	TAGTTGTCGG	GTTTGGTATA	TATTTTATG	CTTCCAAGA	TAAAGAAATT	AATAATAC	TTGATGCAAT	TGAAGATAAA	AATTTCAAA	130
	M744/10	ATGAAAAAGA	TAAAAATTGT	TCCACTTATT	TAAATAGTTG	TAGTTGTCGG	GTTTGGTATA	TATTTTATG	CTTCCAAGA	TAAAGAAATT	AATAATAC	TTGATGCAAT	TGAAGATAAA	AATTTCAAA	130
	M1529/10	ATGAAAAAGA	TAAAAATTGT	TCCACTTATT	TAAATAGTTG	TAGTTGTCGG	GTTTGGTATA	TATTTTATG	CTTCCAAGA	TAAAGAAATT	AATAATAC	TTGATGCAAT	TGAAGATAAA	AATTTCAAA	130
	M1383/10	ATGAAAAAGA	TAAAAATTGT	TCCACTTATT	TAAATAGTTG	TAGTTGTCGG	GTTTGGTATA	TATTTTATG	CTTCCAAGA	TAAAGAAATT	AATAATAC	TTGATGCAAT	TGAAGATAAA	AATTTCAAA	130
	M1186/10	ATGAAAAAGA	TAAAAATTGT	TCCACTTATT	TAAATAGTTG	TAGTTGTCGG	GTTTGGTATA	TATTTTATG	CTTCCAAGA	TAAAGAAATT	AATAATAC	TTGATGCAAT	TGAAGATAAA	AATTTCAAA	130
	M703/10	ATGAAAAAGA	TAAAAATTGT	TCCACTTATT	TAAATAGTTG	TAGTTGTCGG	GTTTGGTATA	TATTTTATG	CTTCCAAGA	TAAAGAAATT	AATAATAC	TTGATGCAAT	TGAAGATAAA	AATTTCAAA	130
	M8/10	ATGAAAAAGA	TAAAAATTGT	TCCACTTATT	TAAATAGTTG	TAGTTGTCGG	GTTTGGTATA	TATTTTATG	CTTCCAAGA	TAAAGAAATT	AATAATAC	TTGATGCAAT	TGAAGATAAA	AATTTCAAA	130
	Consensus	ATGAAAAAGA	TAAAAATTGT	TCCACTTATT	TAAATAGTTG	TAGTTGTCGG	GTTTGGTATA	TATTTTATG	CTTCCAAGA	TAAAGAAATT	AATAATAC	TTGATGCAAT	TGAAGATAAA	AATTTCAAA	130
S. aurN315	131	AAGTTTATAA	AGATAGCAGT	TATATTTCTA	AAAGCGATAA	TGGTGAAGTA	GAAATGACTG	AACGTCGGAT	AAAAATATAT	AATAGTTTATG	CGGTTAAAGA	TATAAACACATT	CAGGATCGTA	AAATAAAAA	260
	S. epirP62A	AAGTTTATAA	AGATAGCAGT	TATATTTCTA	AAAGCGATAA	TGGTGAAGTA	GAAATGACTG	AACGTCGGAT	AAAAATATAT	AATAGTTTATG	CGGTTAAAGA	TATAAACACATT	CAGGATCGTA	AAATAAAAA	260
	M744/10	AAGTTTATAA	AGATAGCAGT	TATATTTCTA	AAAGCGATAA	TGGTGAAGTA	GAAATGACTG	AACGTCGGAT	AAAAATATAT	AATAGTTTATG	CGGTTAAAGA	TATAAACACATT	CAGGATCGTA	AAATAAAAA	260
	M1529/10	AAGTTTATAA	AGATAGCAGT	TATATTTCTA	AAAGCGATAA	TGGTGAAGTA	GAAATGACTG	AACGTCGGAT	AAAAATATAT	AATAGTTTATG	CGGTTAAAGA	TATAAACACATT	CAGGATCGTA	AAATAAAAA	260
	M1383/10	AAGTTTATAA	AGATAGCAGT	TATATTTCTA	AAAGCGATAA	TGGTGAAGTA	GAAATGACTG	AACGTCGGAT	AAAAATATAT	AATAGTTTATG	CGGTTAAAGA	TATAAACACATT	CAGGATCGTA	AAATAAAAA	260
	M1186/10	AAGTTTATAA	AGATAGCAGT	TATATTTCTA	AAAGCGATAA	TGGTGAAGTA	GAAATGACTG	AACGTCGGAT	AAAAATATAT	AATAGTTTATG	CGGTTAAAGA	TATAAACACATT	CAGGATCGTA	AAATAAAAA	260
	M703/10	AAGTTTATAA	AGATAGCAGT	TATATTTCTA	AAAGCGATAA	TGGTGAAGTA	GAAATGACTG	AACGTCGGAT	AAAAATATAT	AATAGTTTATG	CGGTTAAAGA	TATAAACACATT	CAGGATCGTA	AAATAAAAA	260
	M8/10	AAGTTTATAA	AGATAGCAGT	TATATTTCTA	AAAGCGATAA	TGGTGAAGTA	GAAATGACTG	AACGTCGGAT	AAAAATATAT	AATAGTTTATG	CGGTTAAAGA	TATAAACACATT	CAGGATCGTA	AAATAAAAA	260
	Consensus	AAGTTTATAA	AGATAGCAGT	TATATTTCTA	AAAGCGATAA	TGGTGAAGTA	GAAATGACTG	AACGTCGGAT	AAAAATATAT	AATAGTTTATG	CGGTTAAAGA	TATAAACACATT	CAGGATCGTA	AAATAAAAA	260
S. aurN315	261	AGTATCTAAA	AATAAAAAAC	GAGTAGATGC	TCAATATATA	ATTAAAAACA	ACTACGGTAA	CATTGATCGC	AACGTTCAAT	TTAATTTTGT	TAAAGAAGAT	GGTATGTGGA	AGTTAGATTG	GGATCATAGC	390
	S. epirP62A	AGTATCTAAA	AATAAAAAAC	GAGTAGATGC	TCAATATATA	ATTAAAAACA	ACTACGGTAA	CATTGATCGC	AACGTTCAAT	TTAATTTTGT	TAAAGAAGAT	GGTATGTGGA	AGTTAGATTG	GGATCATAGC	390
	M744/10	AGTATCTAAA	AATAAAAAAC	GAGTAGATGC	TCAATATATA	ATTAAAAACA	ACTACGGTAA	CATTGATCGC	AACGTTCAAT	TTAATTTTGT	TAAAGAAGAT	GGTATGTGGA	AGTTAGATTG	GGATCATAGC	390
	M1529/10	AGTATCTAAA	AATAAAAAAC	GAGTAGATGC	TCAATATATA	ATTAAAAACA	ACTACGGTAA	CATTGATCGC	AACGTTCAAT	TTAATTTTGT	TAAAGAAGAT	GGTATGTGGA	AGTTAGATTG	GGATCATAGC	390
	M1383/10	AGTATCTAAA	AATAAAAAAC	GAGTAGATGC	TCAATATATA	ATTAAAAACA	ACTACGGTAA	CATTGATCGC	AACGTTCAAT	TTAATTTTGT	TAAAGAAGAT	GGTATGTGGA	AGTTAGATTG	GGATCATAGC	390
	M1186/10	AGTATCTAAA	AATAAAAAAC	GAGTAGATGC	TCAATATATA	ATTAAAAACA	ACTACGGTAA	CATTGATCGC	AACGTTCAAT	TTAATTTTGT	TAAAGAAGAT	GGTATGTGGA	AGTTAGATTG	GGATCATAGC	390
	M703/10	AGTATCTAAA	AATAAAAAAC	GAGTAGATGC	TCAATATATA	ATTAAAAACA	ACTACGGTAA	CATTGATCGC	AACGTTCAAT	TTAATTTTGT	TAAAGAAGAT	GGTATGTGGA	AGTTAGATTG	GGATCATAGC	390
	M8/10	AGTATCTAAA	AATAAAAAAC	GAGTAGATGC	TCAATATATA	ATTAAAAACA	ACTACGGTAA	CATTGATCGC	AACGTTCAAT	TTAATTTTGT	TAAAGAAGAT	GGTATGTGGA	AGTTAGATTG	GGATCATAGC	390
	Consensus	AGTATCTAAA	AATAAAAAAC	GAGTAGATGC	TCAATATATA	ATTAAAAACA	ACTACGGTAA	CATTGATCGC	AACGTTCAAT	TTAATTTTGT	TAAAGAAGAT	GGTATGTGGA	AGTTAGATTG	GGATCATAGC	390
S. aurN315	391	GTCATTATTTC	CAGGAATGCA	GAAAGACCAA	AGCATACATA	TTGAAAAATTT	AAAATCAGAA	CGTGGTAAAA	TTTTAGACCG	AAACAATGTG	GAATTTGGCCA	ATACAGGAAC	AGCATATGAG	ATAGGCATCG	520
	S. epirP62A	GTCATTATTTC	CAGGAATGCA	GAAAGACCAA	AGCATACATA	TTGAAAAATTT	AAAATCAGAA	CGTGGTAAAA	TTTTAGACCG	AAACAATGTG	GAATTTGGCCA	ATACAGGAAC	AGCATATGAG	ATAGGCATCG	520
	M744/10	GTCATTATTTC	CAGGAATGCA	GAAAGACCAA	AGCATACATA	TTGAAAAATTT	AAAATCAGAA	CGTGGTAAAA	TTTTAGACCG	AAACAATGTG	GAATTTGGCCA	ATACAGGAAC	AGCATATGAG	ATAGGCATCG	520
	M1529/10	GTCATTATTTC	CAGGAATGCA	GAAAGACCAA	AGCATACATA	TTGAAAAATTT	AAAATCAGAA	CGTGGTAAAA	TTTTAGACCG	AAACAATGTG	GAATTTGGCCA	ATACAGGAAC	AGCATATGAG	ATAGGCATCG	520
	M1383/10	GTCATTATTTC	CAGGAATGCA	GAAAGACCAA	AGCATACATA	TTGAAAAATTT	AAAATCAGAA	CGTGGTAAAA	TTTTAGACCG	AAACAATGTG	GAATTTGGCCA	ATACAGGAAC	AGCATATGAG	ATAGGCATCG	520
	M1186/10	GTCATTATTTC	CAGGAATGCA	GAAAGACCAA	AGCATACATA	TTGAAAAATTT	AAAATCAGAA	CGTGGTAAAA	TTTTAGACCG	AAACAATGTG	GAATTTGGCCA	ATACAGGAAC	AGCATATGAG	ATAGGCATCG	520
	M703/10	GTCATTATTTC	CAGGAATGCA	GAAAGACCAA	AGCATACATA	TTGAAAAATTT	AAAATCAGAA	CGTGGTAAAA	TTTTAGACCG	AAACAATGTG	GAATTTGGCCA	ATACAGGAAC	AGCATATGAG	ATAGGCATCG	520
	M8/10	GTCATTATTTC	CAGGAATGCA	GAAAGACCAA	AGCATACATA	TTGAAAAATTT	AAAATCAGAA	CGTGGTAAAA	TTTTAGACCG	AAACAATGTG	GAATTTGGCCA	ATACAGGAAC	AGCATATGAG	ATAGGCATCG	520
	Consensus	GTCATTATTTC	CAGGAATGCA	GAAAGACCAA	AGCATACATA	TTGAAAAATTT	AAAATCAGAA	CGTGGTAAAA	TTTTAGACCG	AAACAATGTG	GAATTTGGCCA	ATACAGGAAC	AGCATATGAG	ATAGGCATCG	520

521	S. aurn315	TTCCAAAGAA	TGTATCTAAA	AAAGATTATA	AAGCAATCGC	TAAAGAARCTA	AGTATTCTCG	AAGACTATAT	CAAAACAACA	ATGGATCAAA	ATTGGGTACA	AGATGATACC	TTCGTTCCAC	TTAAACCCGT
	.epiRP62A	TTCCAAAGAA	TGTATCTAAA	AAAGATTATA	AAGCAATCGC	TAAAGAARCTA	AGTATTCTCG	AAGACTATAT	CAAAACAACA	ATGGATCAAA	ATTGGGTACA	AGATGATACC	TTCGTTCCAC	TTAAACCCGT
	M174/10	TTCCAAAGAA	TGTATCTAAA	AAAGATTATA	AAGCAATCGC	TAAAGAARCTA	AGTATTCTCG	AAGACTATAT	CAAAACAACA	ATGGATCAAA	ATTGGGTACA	AGATGATACC	TTCGTTCCAC	TTAAACCCGT
	M1529/10	TTCCAAAGAA	TGTATCTAAA	AAAGATTATA	AAGCAATCGC	TAAAGAARCTA	AGTATTCTCG	AAGACTATAT	CAAAACAACA	ATGGATCAAA	ATTGGGTACA	AGATGATACC	TTCGTTCCAC	TTAAACCCGT
	M1383/10	TTCCAAAGAA	TGTATCTAAA	AAAGATTATA	AAGCAATCGC	TAAAGAARCTA	AGTATTCTCG	AAGACTATAT	CAAAACAACA	ATGGATCAAA	ATTGGGTACA	AGATGATACC	TTCGTTCCAC	TTAAACCCGT
	M1186/10	TTCCAAAGAA	TGTATCTAAA	AAAGATTATA	AAGCAATCGC	TAAAGAARCTA	AGTATTCTCG	AAGACTATAT	CAAAACAACA	ATGGATCAAA	ATTGGGTACA	AGATGATACC	TTCGTTCCAC	TTAAACCCGT
	M703/10	TTCCAAAGAA	TGTATCTAAA	AAAGATTATA	AAGCAATCGC	TAAAGAARCTA	AGTATTCTCG	AAGACTATAT	CAAAACAACA	ATGGATCAAA	ATTGGGTACA	AGATGATACC	TTCGTTCCAC	TTAAACCCGT
	Consensus	TTCCAAAGAA	TGTATCTAAA	AAAGATTATA	AAGCAATCGC	TAAAGAARCTA	AGTATTCTCG	AAGACTATAT	CAAAACAACA	ATGGATCAAA	ATTGGGTACA	AGATGATACC	TTCGTTCCAC	TTAAACCCGT
651	S. aurn315	TAAAAAAATG	GATGAATATT	TAAGTGATT	CGCAAAAAAA	TTTTCATCTTA	CAACTAATGA	AACAGAAAGT	CGTAACTATC	CTCTAGGAAA	AGCGACTTCA	CATCTATTAG	GTTATGTTGG	TCCCATTAAC
	.epiRP62A	TAAAAAAATG	GATGAATATT	TAAGTGATT	CGCAAAAAAA	TTTTCATCTTA	CAACTAATGA	AACAGAAAGT	CGTAACTATC	CTCTAGGAAA	AGCGACTTCA	CATCTATTAG	GTTATGTTGG	TCCCATTAAC
	M174/10	TAAAAAAATG	GATGAATATT	TAAGTGATT	CGCAAAAAAA	TTTTCATCTTA	CAACTAATGA	AACAGAAAGT	CGTAACTATC	CTCTAGGAAA	AGCGACTTCA	CATCTATTAG	GTTATGTTGG	TCCCATTAAC
	M1529/10	TAAAAAAATG	GATGAATATT	TAAGTGATT	CGCAAAAAAA	TTTTCATCTTA	CAACTAATGA	AACAGAAAGT	CGTAACTATC	CTCTAGGAAA	AGCGACTTCA	CATCTATTAG	GTTATGTTGG	TCCCATTAAC
	M1383/10	TAAAAAAATG	GATGAATATT	TAAGTGATT	CGCAAAAAAA	TTTTCATCTTA	CAACTAATGA	AACAGAAAGT	CGTAACTATC	CTCTAGGAAA	AGCGACTTCA	CATCTATTAG	GTTATGTTGG	TCCCATTAAC
	M1186/10	TAAAAAAATG	GATGAATATT	TAAGTGATT	CGCAAAAAAA	TTTTCATCTTA	CAACTAATGA	AACAGAAAGT	CGTAACTATC	CTCTAGGAAA	AGCGACTTCA	CATCTATTAG	GTTATGTTGG	TCCCATTAAC
	M703/10	TAAAAAAATG	GATGAATATT	TAAGTGATT	CGCAAAAAAA	TTTTCATCTTA	CAACTAATGA	AACAGAAAGT	CGTAACTATC	CTCTAGGAAA	AGCGACTTCA	CATCTATTAG	GTTATGTTGG	TCCCATTAAC
	M8/10	TAAAAAAATG	GATGAATATT	TAAGTGATT	CGCAAAAAAA	TTTTCATCTTA	CAACTAATGA	AACAGAAAGT	CGTAACTATC	CTCTAGGAAA	AGCGACTTCA	CATCTATTAG	GTTATGTTGG	TCCCATTAAC
	Consensus	TAAAAAAATG	GATGAATATT	TAAGTGATT	CGCAAAAAAA	TTTTCATCTTA	CAACTAATGA	AACAGAAAGT	CGTAACTATC	CTCTAGGAAA	AGCGACTTCA	CATCTATTAG	GTTATGTTGG	TCCCATTAAC
781	S. aurn315	TCTGAAGAA	TAAACAAAA	AGAATATAAA	GGCTATAAAG	ATGATGCAGT	TATTGGTAAA	AAGGACTCG	AAAAACTTTA	CGATAAAAAG	CTCCAACATG	AGATGGCTA	TCGTGTCACA	ATCGTTGACG
	.epiRP62A	TCTGAAGAA	TAAACAAAA	AGAATATAAA	GGCTATAAAG	ATGATGCAGT	TATTGGTAAA	AAGGACTCG	AAAAACTTTA	CGATAAAAAG	CTCCAACATG	AGATGGCTA	TCGTGTCACA	ATCGTTGACG
	M174/10	TCTGAAGAA	TAAACAAAA	AGAATATAAA	GGCTATAAAG	ATGATGCAGT	TATTGGTAAA	AAGGACTCG	AAAAACTTTA	CGATAAAAAG	CTCCAACATG	AGATGGCTA	TCGTGTCACA	ATCGTTGACG
	M1529/10	TCTGAAGAA	TAAACAAAA	AGAATATAAA	GGCTATAAAG	ATGATGCAGT	TATTGGTAAA	AAGGACTCG	AAAAACTTTA	CGATAAAAAG	CTCCAACATG	AGATGGCTA	TCGTGTCACA	ATCGTTGACG
	M1383/10	TCTGAAGAA	TAAACAAAA	AGAATATAAA	GGCTATAAAG	ATGATGCAGT	TATTGGTAAA	AAGGACTCG	AAAAACTTTA	CGATAAAAAG	CTCCAACATG	AGATGGCTA	TCGTGTCACA	ATCGTTGACG
	M1186/10	TCTGAAGAA	TAAACAAAA	AGAATATAAA	GGCTATAAAG	ATGATGCAGT	TATTGGTAAA	AAGGACTCG	AAAAACTTTA	CGATAAAAAG	CTCCAACATG	AGATGGCTA	TCGTGTCACA	ATCGTTGACG
	M703/10	TCTGAAGAA	TAAACAAAA	AGAATATAAA	GGCTATAAAG	ATGATGCAGT	TATTGGTAAA	AAGGACTCG	AAAAACTTTA	CGATAAAAAG	CTCCAACATG	AGATGGCTA	TCGTGTCACA	ATCGTTGACG
	M8/10	TCTGAAGAA	TAAACAAAA	AGAATATAAA	GGCTATAAAG	ATGATGCAGT	TATTGGTAAA	AAGGACTCG	AAAAACTTTA	CGATAAAAAG	CTCCAACATG	AGATGGCTA	TCGTGTCACA	ATCGTTGACG
	Consensus	TCTGAAGAA	TAAACAAAA	AGAATATAAA	GGCTATAAAG	ATGATGCAGT	TATTGGTAAA	AAGGACTCG	AAAAACTTTA	CGATAAAAAG	CTCCAACATG	AGATGGCTA	TCGTGTCACA	ATCGTTGACG
911	S. aurn315	ATAATAGCAA	TACAATCGCA	CATACATTAA	TAGAGAAAAA	GAAAAAAGAT	GGCAAAGATA	TTCAACTAAC	TATTGATGCT	AAAAGTTCAAA	AGAGTATTTA	TAACAACATG	AAAAATGATT	ATGGCTCAGG
	.epiRP62A	ATAATAGCAA	TACAATCGCA	CATACATTAA	TAGAGAAAAA	GAAAAAAGAT	GGCAAAGATA	TTCAACTAAC	TATTGATGCT	AAAAGTTCAAA	AGAGTATTTA	TAACAACATG	AAAAATGATT	ATGGCTCAGG
	M174/10	ATAATAGCAA	TACAATCGCA	CATACATTAA	TAGAGAAAAA	GAAAAAAGAT	GGCAAAGATA	TTCAACTAAC	TATTGATGCT	AAAAGTTCAAA	AGAGTATTTA	TAACAACATG	AAAAATGATT	ATGGCTCAGG
	M1529/10	ATAATAGCAA	TACAATCGCA	CATACATTAA	TAGAGAAAAA	GAAAAAAGAT	GGCAAAGATA	TTCAACTAAC	TATTGATGCT	AAAAGTTCAAA	AGAGTATTTA	TAACAACATG	AAAAATGATT	ATGGCTCAGG
	M1383/10	ATAATAGCAA	TACAATCGCA	CATACATTAA	TAGAGAAAAA	GAAAAAAGAT	GGCAAAGATA	TTCAACTAAC	TATTGATGCT	AAAAGTTCAAA	AGAGTATTTA	TAACAACATG	AAAAATGATT	ATGGCTCAGG
	M1186/10	ATAATAGCAA	TACAATCGCA	CATACATTAA	TAGAGAAAAA	GAAAAAAGAT	GGCAAAGATA	TTCAACTAAC	TATTGATGCT	AAAAGTTCAAA	AGAGTATTTA	TAACAACATG	AAAAATGATT	ATGGCTCAGG
	M703/10	ATAATAGCAA	TACAATCGCA	CATACATTAA	TAGAGAAAAA	GAAAAAAGAT	GGCAAAGATA	TTCAACTAAC	TATTGATGCT	AAAAGTTCAAA	AGAGTATTTA	TAACAACATG	AAAAATGATT	ATGGCTCAGG
	M8/10	ATAATAGCAA	TACAATCGCA	CATACATTAA	TAGAGAAAAA	GAAAAAAGAT	GGCAAAGATA	TTCAACTAAC	TATTGATGCT	AAAAGTTCAAA	AGAGTATTTA	TAACAACATG	AAAAATGATT	ATGGCTCAGG
	Consensus	ATAATAGCAA	TACAATCGCA	CATACATTAA	TAGAGAAAAA	GAAAAAAGAT	GGCAAAGATA	TTCAACTAAC	TATTGATGCT	AAAAGTTCAAA	AGAGTATTTA	TAACAACATG	AAAAATGATT	ATGGCTCAGG
1041	S. aurn315	TACTGCTATC	CACCCTCAAA	CAGGTGAATT	ATTAGCACTT	GTAAGGCACAC	CTTTCATATGA	CGTCTATCCA	TTTATGTTATG	GCATGAGTAA	CGAAGAATAT	AATAAATTAA	CCGAAGATAA	AAAAGAACCT
	.epiRP62A	TACTGCTATC	CACCCTCAAA	CAGGTGAATT	ATTAGCACTT	GTAAGGCACAC	CTTTCATATGA	CGTCTATCCA	TTTATGTTATG	GCATGAGTAA	CGAAGAATAT	AATAAATTAA	CCGAAGATAA	AAAAGAACCT
	M174/10	TACTGCTATC	CACCCTCAAA	CAGGTGAATT	ATTAGCACTT	GTAAGGCACAC	CTTTCATATGA	CGTCTATCCA	TTTATGTTATG	GCATGAGTAA	CGAAGAATAT	AATAAATTAA	CCGAAGATAA	AAAAGAACCT
	M1529/10	TACTGCTATC	CACCCTCAAA	CAGGTGAATT	ATTAGCACTT	GTAAGGCACAC	CTTTCATATGA	CGTCTATCCA	TTTATGTTATG	GCATGAGTAA	CGAAGAATAT	AATAAATTAA	CCGAAGATAA	AAAAGAACCT
	M1383/10	TACTGCTATC	CACCCTCAAA	CAGGTGAATT	ATTAGCACTT	GTAAGGCACAC	CTTTCATATGA	CGTCTATCCA	TTTATGTTATG	GCATGAGTAA	CGAAGAATAT	AATAAATTAA	CCGAAGATAA	AAAAGAACCT
	M1186/10	TACTGCTATC	CACCCTCAAA	CAGGTGAATT	ATTAGCACTT	GTAAGGCACAC	CTTTCATATGA	CGTCTATCCA	TTTATGTTATG	GCATGAGTAA	CGAAGAATAT	AATAAATTAA	CCGAAGATAA	AAAAGAACCT
	M703/10	TACTGCTATC	CACCCTCAAA	CAGGTGAATT	ATTAGCACTT	GTAAGGCACAC	CTTTCATATGA	CGTCTATCCA	TTTATGTTATG	GCATGAGTAA	CGAAGAATAT	AATAAATTAA	CCGAAGATAA	AAAAGAACCT
	M8/10	TACTGCTATC	CACCCTCAAA	CAGGTGAATT	ATTAGCACTT	GTAAGGCACAC	CTTTCATATGA	CGTCTATCCA	TTTATGTTATG	GCATGAGTAA	CGAAGAATAT	AATAAATTAA	CCGAAGATAA	AAAAGAACCT
	Consensus	TACTGCTATC	CACCCTCAAA	CAGGTGAATT	ATTAGCACTT	GTAAGGCACAC	CTTTCATATGA	CGTCTATCCA	TTTATGTTATG	GCATGAGTAA	CGAAGAATAT	AATAAATTAA	CCGAAGATAA	AAAAGAACCT
1170	S. aurn315	TACTGCTATC	CACCCTCAAA	CAGGTGAATT	ATTAGCACTT	GTAAGGCACAC	CTTTCATATGA	CGTCTATCCA	TTTATGTTATG	GCATGAGTAA	CGAAGAATAT	AATAAATTAA	CCGAAGATAA	AAAAGAACCT
	.epiRP62A	TACTGCTATC	CACCCTCAAA	CAGGTGAATT	ATTAGCACTT	GTAAGGCACAC	CTTTCATATGA	CGTCTATCCA	TTTATGTTATG	GCATGAGTAA	CGAAGAATAT	AATAAATTAA	CCGAAGATAA	AAAAGAACCT
	M174/10	TACTGCTATC	CACCCTCAAA	CAGGTGAATT	ATTAGCACTT	GTAAGGCACAC	CTTTCATATGA	CGTCTATCCA	TTTATGTTATG	GCATGAGTAA	CGAAGAATAT	AATAAATTAA	CCGAAGATAA	AAAAGAACCT
	M1529/10	TACTGCTATC	CACCCTCAAA	CAGGTGAATT	ATTAGCACTT	GTAAGGCACAC	CTTTCATATGA	CGTCTATCCA	TTTATGTTATG	GCATGAGTAA	CGAAGAATAT	AATAAATTAA	CCGAAGATAA	AAAAGAACCT
	M1383/10	TACTGCTATC	CACCCTCAAA	CAGGTGAATT	ATTAGCACTT	GTAAGGCACAC	CTTTCATATGA	CGTCTATCCA	TTTATGTTATG	GCATGAGTAA	CGAAGAATAT	AATAAATTAA	CCGAAGATAA	AAAAGAACCT
	M1186/10	TACTGCTATC	CACCCTCAAA	CAGGTGAATT	ATTAGCACTT	GTAAGGCACAC	CTTTCATATGA	CGTCTATCCA	TTTATGTTATG	GCATGAGTAA	CGAAGAATAT	AATAAATTAA	CCGAAGATAA	AAAAGAACCT
	M703/10	TACTGCTATC	CACCCTCAAA	CAGGTGAATT	ATTAGCACTT	GTAAGGCACAC	CTTTCATATGA	CGTCTATCCA	TTTATGTTATG	GCATGAGTAA	CGAAGAATAT	AATAAATTAA	CCGAAGATAA	AAAAGAACCT
	M8/10	TACTGCTATC	CACCCTCAAA	CAGGTGAATT	ATTAGCACTT	GTAAGGCACAC	CTTTCATATGA	CGTCTATCCA	TTTATGTTATG	GCATGAGTAA	CGAAGAATAT	AATAAATTAA	CCGAAGATAA	AAAAGAACCT
	Consensus	TACTGCTATC	CACCCTCAAA	CAGGTGAATT	ATTAGCACTT	GTAAGGCACAC	CTTTCATATGA	CGTCTATCCA	TTTATGTTATG	GCATGAGTAA	CGAAGAATAT	AATAAATTAA	CCGAAGATAA	AAAAGAACCT

[illegible]

using MultAlign Program <http://multalin.toulouse.inra.fr/multalin/>

[illegible]

[illegible]

781	S. fleur-SMF	TCTGAAGAAT	TAAACACAAA	AGAAATATAA	GGTTATAAAG	ATGATGCAGT	TGTTGCTAAA	AAGGACTCG	AAAAACTTTA	CGATAAAAAG	CTCCAACATG	AAGATGGCTA	TCGTGTCACA	ATCTTTGACG	910
	S. fleurCCU	TCTGAAGAAT	TAAACACAAA	AGAAATATAA	GGCTATAAAG	ATGATGCAGT	TATTGCTAAA	AAGGACTCG	AAAAACTTTA	CGATAAAAAG	CTCCAACATG	AAGATGGCTA	TCGTGTCACA	ATCGTTGACG	
	M143/10	TCTGAAGAAT	TAAACACAAA	AGAAATATAA	GGCTATAAAG	ATGATGCAGT	TATTGCTAAA	AAGGACTCG	AAAAACTTTA	CGATAAAAAG	CTCCAACATG	AAGATGGCTA	TCGTGTCACA	ATCGTTGACG	
	M277/10	TCTGAAGAAT	TAAACACAAA	AGAAATATAA	GGCTATAAAG	ATGATGCAGT	TATTGCTAAA	AAGGACTCG	AAAAACTTTA	CGATAAAAAG	CTCCAACATG	AAGATGGCTA	TCGTGTCACA	ATCGTTGACG	
	M404-1/10	TCTGAAGAAT	TAAACACAAA	AGAAATATAA	GGCTATAAAG	ATGATGCAGT	TATTGCTAAA	AAGGACTCG	AAAAACTTTA	CGATAAAAAG	CTCCAACATG	AAGATGGCTA	TCGTGTCACA	ATCGTTGACG	
	M545/10	TCTGAAGAAT	TAAACACAAA	AGAAATATAA	GGCTATAAAG	ATGATGCAGT	TATTGCTAAA	AAGGACTCG	AAAAACTTTA	CGATAAAAAG	CTCCAACATG	AAGATGGCTA	TCGTGTCACA	ATCGTTGACG	
	M4276-1/09	TCTGAAGAAT	TAAACACAAA	AGAAATATAA	GGCTATAAAG	ATGATGCAGT	TATTGCTAAA	AAGGACTCG	AAAAACTTTA	CGATAAAAAG	CTCCAACATG	AAGATGGCTA	TCGTGTCACA	ATCGTTGACG	
	M1125-2/10	TCTGAAGAAT	TAAACACAAA	AGAAATATAA	GGCTATAAAG	ATGATGCAGT	TATTGCTAAA	AAGGACTCG	AAAAACTTTA	CGATAAAAAG	CTCCAACATG	AAGATGGCTA	TCGTGTCACA	ATCGTTGACG	
	M1961/10	TCTGAAGAAT	TAAACACAAA	AGAAATATAA	GGCTATAAAG	ATGATGCAGT	TATTGCTAAA	AAGGACTCG	AAAAACTTTA	CGATAAAAAG	CTCCAACATG	AAGATGGCTA	TCGTGTCACA	ATCGTTGACG	
	M1191-2/10	TCTGAAGAAT	TAAACACAAA	AGAAATATAA	GGCTATAAAG	ATGATGCAGT	TATTGCTAAA	AAGGACTCG	AAAAACTTTA	CGATAAAAAG	CTCCAACATG	AAGATGGCTA	TCGTGTCACA	ATCGTTGACG	
	M3783/10	TCTGAAGAAT	TAAACACAAA	AGAAATATAA	GGCTATAAAG	ATGATGCAGT	TATTGCTAAA	AAGGACTCG	AAAAACTTTA	CGATAAAAAG	CTCCAACATG	AAGATGGCTA	TCGTGTCACA	ATCGTTGACG	
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911	S. fleur-SMF	ATAAATAGCAA	TACAAATCGCA	CATACATTAA	TAGAGAAAAA	GAAAAAAGAT	GGCAAAGATA	TTCAACTAAC	TATTGATGCT	AAAGTTCAAA	AGAGTATTTA	TAAACAACATG	AAAAATGATT	ATGGCTCAGG	1040
	S. fleurCCU	ATAAATAGCAA	TACAAATCGCA	CATACATTAA	TAGAGAAAAA	GAAAAAAGAT	GGCAAAGATA	TTCAACTAAC	TATTGATGCT	AAAGTTCAAA	AGAGTATTTA	TAAACAACATG	AAAAATGATT	ATGGCTCAGG	
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	S. fleurCCU	TACTGCTATC	CACCCTCAAA	CAGGTGAATT	ATTAGCACTT	GTAAGCACC	CTTCATATGA	CGTCTATCCA	TTTATGTATG	GCATGAGTAA	CGAAGAATAT	AATAAATTAA	CCGAAGATAA	AAAAAACC	
	M143/10	TACTGCTATC	CACCCTCAAA	CAGGTGAATT	ATTAGCACTT	GTAAGCACC	CTTCATATGA	CGTCTATCCA	TTTATGTATG	GCATGAGTAA	CGAAGAATAT	AATAAATTAA	CCGAAGATAA	AAAAAACC	
	M277/10	TACTGCTATC	CACCCTCAAA	CAGGTGAATT	ATTAGCACTT	GTAAGCACC	CTTCATATGA	CGTCTATCCA	TTTATGTATG	GCATGAGTAA	CGAAGAATAT	AATAAATTAA	CCGAAGATAA	AAAAAACC	
	M404-1/10	TACTGCTATC	CACCCTCAAA	CAGGTGAATT	ATTAGCACTT	GTAAGCACC	CTTCATATGA	CGTCTATCCA	TTTATGTATG	GCATGAGTAA	CGAAGAATAT	AATAAATTAA	CCGAAGATAA	AAAAAACC	
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	M1125-2/10	TACTGCTATC	CACCCTCAAA	CAGGTGAATT	ATTAGCACTT	GTAAGCACC	CTTCATATGA	CGTCTATCCA	TTTATGTATG	GCATGAGTAA	CGAAGAATAT	AATAAATTAA	CCGAAGATAA	AAAAAACC	
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Consensus		TACTGCTATC	CACCCTCAAA	CAGGTGAATT	aTTAGCACTT	GTAAGCACC	CTTCATATGA	CGTCTATCCA	TTTATGTATG	GCATGAGTAA	CGAAGAATAT	AATAAATTAA	CCGAAGATAA	AAAAAACC	

1171	S. fleurSMF	TACAACTTCA	CCAGGTTTCA	CTCAAAAAAT	ATTAAACAGCA	ATGATTGGGT	TAAATAACAA	AACATTAGAC	GATAAAACAA	GTTATAAAAT	CGATGCTAAA	GGTTGGCAAA	1300	
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	M1961/10	TACAACTTCA	CCAGGTTTCA	CTCAAAAAAT	ATTAAACAGCA	ATGATTGGGT	TAAATAACAA	AACATTAGAC	GATAAAACAA	GTTATAAAAT	CGATGCTAAA	GGTTGGCAAA		
	M1191-2/10	TACAACTTCA	CCAGGTTTCA	CTCAAAAAAT	ATTAAACAGCA	ATGATTGGGT	TAAATAACAA	AACATTAGAC	GATAAAACAA	GTTATAAAAT	CGATGCTAAA	GGTTGGCAAA		
	M3783/10	TACAACTTCA	CCAGGTTTCA	CTCAAAAAAT	ATTAAACAGCA	ATGATTGGGT	TAAATAACAA	AACATTAGAC	GATAAAACAA	GTTATAAAAT	CGATGCTAAA	GGTTGGCAAA		
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Consensus	GAATTTTCAA	AAAGGCATGA	AAAACTAGG	TGTTGGTGAA	GATATACCAA	GTGATTATCC	ATTTTATAAT	GCTCAAAATTT	CAAAACAAAA	TTTAGATAAT	TAGTCTGATTC	AGGTTACCGA		

	1951	2007
S. f. <i>smf</i>	AAAGTGTATG	ATGAGCTATA
S. f. <i>fleurCU</i>	AAAGTGTATG	ATGAGCTATA
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M277/10	AAAGTGTATG	ATGAGCTATA
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M205/10	AAAGTGTATG	ATGAGCTATA
M4460/09	AAAGTGTATG	ATGAGCTATA
Consensus	AAAGTGTATG	ATGAGCTATA
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	ATAAAAAAAT	ATAAAAAAAT
	ACGATATAGA	ACGATATAGA
	TGAATAAA	TGAATAAA

***mecA* gene sequence of *S. fleurettii* strain M4460/09 deposited into EMBL/Genbank/DDBJ databases**

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ID    HE861945; SV 1; linear; genomic DNA; STD; PRO; 2164 BP.
XX
ST * private 15-JUL-2012
XX
AC    HE861945;
XX
DT    19-JUN-2012 (Rel. 113, Created)
DT    19-JUN-2012 (Rel. 113, Last updated, Version 0)
XX
DE    Staphylococcus fleurettii mecA gene for penicillin binding protein 2',
DE    strain M4460-09
XX
KW    .
XX
OS    Staphylococcus fleurettii
OC    Bacteria; Firmicutes; Bacillales; Staphylococcus.
XX
RN    [1]
RP    1-2164
RA    Perreten V.;
RT    ;
RL    Submitted (15-JUN-2012) to the INSDC.
RL    University of Berne, Institute of Veterinary Bacteriology, Laenggassstrasse
RL    122, CH - 3001 Berne, SWITZERLAND.
XX
RN    [2]
RA    Frey Y., Schwendener S., Perreten V.;
RT    "Methicillin resistance in Staphylococcus fleurettii from bovine mastitis";
RL    Unpublished.
XX
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FH	Key	Location/Qualifiers
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FT		YGNIDRNVQFNFVKEDGMWKLWDHDSV IIPGMQKDQSIH IENLKSERGKILDRNNVELA
FT		NTGTAYEIGIVPKNVSKKDYKAI AKELSI SEDYIKQQMDQNWVQDDTFVPLKTVKKMDE
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FT		KNDYGSGETAIHPQTGELLALVSTPSYDVYPFMYGMSNEEYKNLTEDKKEPLLNFQITT
FT		SPGSTQKILTAMIGLNNKTLDDKTSYKIDGKGWQKDKSWGGYNVTRYEVVNGNIDLKQA
FT		IESSDNIFFARVALELGSKKFEKGMKKLGVGEDIPSDYPFYNAQISKNLNDNEILLADS
FT		GYGQGEILINPVQILSIYSALENNGNINAPHLLKDTKNKVWKKNIISKENINLLTDGMQ
FT		QVVKTHKKDIYRPIYANLIGKSGTAE LKMKQGETGRQIGWFI SYDKDNPNMMMAINVKD
FT		VQDKGMASYNAKISGKVYDELYENGKKYDIDE"

XX

SQ	Sequence 2164 BP; 916 A; 296 C; 366 G; 586 T; 0 other;	
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	ttaatagttg tagttgtcgg gtttggtata tatttttatg cttcaaaaaga taaagaaatt	180
	aataaaaacta ttgatgcaat tgaagataaa aattttcaaac aagttttataa agatagcagt	240
	tatatTTcca aaagcgataa tgggtgaagta gaaatgactg aacgtccgat aaaaatatat	300
	aaaagtttag gcgttaaaga tataaacatt caagaacgca aaataaaaaa aatatctaaa	360
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	ctgctcaaca agttccagat tacaacttca ccagggttcaa ctcaaaaaat attaacagca	1320
	atgattgggt taaataacaa aacattagac gataaaacaa gttataaaat cgatggtaaa	1380
	ggttggcaaa aagataaatc ttgggggtgggt tacaacgtta caagatatga agtggtaaat	1440
	ggtaatatcg acttaaaaca agcaatagaa tcatcagata acattttctt tgctagagta	1500
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***mecA* gene sequence of *S. fleurettii* strain M143/10 deposited into EMBL/Genbank/ DDBJ databases**

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ID    HE978795; SV 1; linear; genomic DNA; STD; PRO; 2007 BP.
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AC    HE978795;
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DT    16-AUG-2012 (Rel. 113, Created)
DT    16-AUG-2012 (Rel. 113, Last updated, Version 0)
XX
DE    Staphylococcus fleurettii mecA gene for penicillin binding protein 2',
DE    strain M143-10
XX
KW    .
XX
OS    Staphylococcus fleurettii
OC    Bacteria; Firmicutes; Bacillales; Staphylococcus.
XX
RN    [1]
RP    1-2007
RA    Perreten V.;
RT    ;
RL    Submitted (14-AUG-2012) to the INSDC.
RL    University of Berne, Institute of Veterinary Bacteriology, Laenggassstrasse
RL    122, CH - 3001 Berne, SWITZERLAND.
XX
RN    [2]
RA    Frey Y., Schwendener S., Perreten V.;
RT    "Methicillin resistance in Staphylococcus fleurettii from bovine mastitis";
RL    Unpublished.
XX
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SQ

Sequence 2007 BP; 857 A; 271 C; 342 G; 537 T; 0 other;

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***mecA* gene sequence of *S. fleurettii* strain M205/10 deposited into EMBL/Genbank/ DDBJ databases**

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AC    HE978794;
XX
DT    16-AUG-2012 (Rel. 113, Created)
DT    16-AUG-2012 (Rel. 113, Last updated, Version 0)
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DE    strain M205-10
XX
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XX
OS    Staphylococcus fleurettii
OC    Bacteria; Firmicutes; Bacillales; Staphylococcus.
XX
RN    [1]
RP    1-2007
RA    Perreten V.;
RT    ;
RL    Submitted (14-AUG-2012) to the INSDC.
RL    University of Berne, Institute of Veterinary Bacteriology, Laenggassstrasse
RL    122, CH - 3001 Berne, SWITZERLAND.
XX
RN    [2]
RA    Frey Y., Schwendener S., Perreten V.;
RT    "Methicillin resistance in Staphylococcus fleurettii from bovine mastitis";
RL    Unpublished.
XX
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SQ

Sequence 2007 BP; 857 A; 270 C; 342 G; 538 T; 0 other;

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ID HE978796; SV 1; linear; genomic DNA; STD; PRO; 2007 BP.
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XX
AC HE978796;
XX
DT 16-AUG-2012 (Rel. 113, Created)
DT 16-AUG-2012 (Rel. 113, Last updated, Version 0)
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DE Staphylococcus fleurettii mecA gene for penicillin binding protein 2',
DE strain M3783-09
XX
KW .
XX
OS Staphylococcus fleurettii
OC Bacteria; Firmicutes; Bacillales; Staphylococcus.
XX
RN [1]
RP 1-2007
RA Perreten V.;
RT ;
RL Submitted (14-AUG-2012) to the INSDC.
RL University of Berne, Institute of Veterinary Bacteriology, Laenggassstrasse
RL 122, CH - 3001 Berne, SWITZERLAND.
XX
RN [2]
RA Frey Y., Schwendener S., Perreten V.;
RT "Methicillin resistance in Staphylococcus fleurettii from bovine mastitis";
RL Unpublished.
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XX

SQ

Sequence 2007 BP; 856 A; 271 C; 343 G; 537 T; 0 other;

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***mecA* gene sequence of *S. epidermidis* strain M8/10 deposited into EMBL/Genbank/ DDBJ databases**

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ID    HE978797; SV 1; linear; genomic DNA; STD; PRO; 2007 BP.
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AC    HE978797;
XX
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DT    16-AUG-2012 (Rel. 113, Last updated, Version 0)
XX
DE    Staphylococcus epidermidis mecA gene for penicillin binding protein 2',
DE    strain M8-10
XX
KW    .
XX
OS    Staphylococcus epidermidis
OC    Bacteria; Firmicutes; Bacillales; Staphylococcus.
XX
RN    [1]
RP    1-2007
RA    Perreten V.;
RT    ;
RL    Submitted (14-AUG-2012) to the INSDC.
RL    University of Berne, Institute of Veterinary Bacteriology, Laenggassstrasse
RL    122, CH - 3001 Berne, SWITZERLAND.
XX
RN    [2]
RA    Frey Y., Schwendener S., Perreten V.;
RT    "Methicillin resistance in Staphylococcus epidermidis from bovine
RT    mastitis";
RL    Unpublished.
XX
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Sequence 2007 BP; 855 A; 272 C; 341 G; 539 T; 0 other;

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AC HE978798;
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DT 16-AUG-2012 (Rel. 113, Last updated, Version 0)
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DE Staphylococcus epidermidis mecA gene for penicillin binding protein 2',
DE strain M703-10
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KW .
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OS Staphylococcus epidermidis
OC Bacteria; Firmicutes; Bacillales; Staphylococcus.
XX
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RP 1-2007
RA Perreten V.;
RT ;
RL Submitted (14-AUG-2012) to the INSDC.
RL University of Berne, Institute of Veterinary Bacteriology, Laenggassstrasse
RL 122, CH - 3001 Berne, SWITZERLAND.
XX
RN [2]
RA Frey Y., Schwendener S., Perreten V.;
RT "Methicillin resistance in Staphylococcus epidermidis from bovine
RT mastitis";
RL Unpublished.
XX
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FT IESSDNIFARVALELGSKKFEKGMKKLGVGEDIPSDYPFYNAQISNKNLDNEILLADS
FT GYGQGEILINPVQILSIYSALENNGNINAPHLLKDTKNKVWKKNIISKENINLLTDGMQ
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SQ Sequence 2007 BP; 855 A; 272 C; 342 G; 538 T; 0 other;

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SQ	Sequence 2007 BP; 856 A; 272 C; 341 G; 538 T; 0 other;	
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***mecA* gene sequence of *S. xylosus* strain M1545/10 deposited into EMBL/Genbank/DDBJ databases**

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AC    HE978800;
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DT    16-AUG-2012 (Rel. 113, Last updated, Version 0)
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OC    Bacteria; Firmicutes; Bacillales; Staphylococcus.
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RP    1-2007
RA    Perreten V.;
RT    ;
RL    Submitted (14-AUG-2012) to the INSDC.
RL    University of Berne, Institute of Veterinary Bacteriology, Laenggassstrasse
RL    122, CH - 3001 Berne, SWITZERLAND.
XX
RN    [2]
RA    Frey Y., Schwendener S., Perreten V.;
RT    "Methicillin resistance in Staphylococcus haemolyticus and Staphylococcus
RT    xylosus from bovine mastitis";
RL    Unpublished.
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SQ Sequence 2007 BP; 856 A; 272 C; 341 G; 538 T; 0 other;

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//

CURRICULUM VITAE

Name, Vorname	Frey Yvonne
Geburtsdatum	17.11.1985
Geburtsort	Kantonsspital Aarau, in Aarau
Nationalität	Schweizerin
Heimatort	Küttigen, AG
Aug. 2001 – Jun. 2005	Gymnasium an der alten Kantonsschule Aarau in Aarau, Schweiz
30. Juni 2005	gymnasiale Maturität, Alte Kantonsschule Aarau in Aarau, Schweiz
Okt. 2006 – Aug. 2011	Studium der Veterinärmedizin, Vetsuisse Fakultät, Universität Bern, Bern, Schweiz
30. September 2011	Abschlussprüfung vet.med., Universität Bern, Bern, Schweiz
Sept. 2011 – Aug. 2012	Anfertigung der Dissertation unter der Leitung von Professor V. Perreten am Departement für Veterinärbakteriologie der Vetsuisse – Fakultät Universität Bern Direktor: Prof. Dr. Joachim Frey