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Spatio-temporal occurrence of *Culicoides* biting midges in the climatic regions of Switzerland, along with large scale species identification by MALDI-TOF mass spectrometry

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Abstract

Background: *Culicoides* biting midges are incriminated as biological vectors of a number of viruses, e.g. bluetongue virus. In order to define vector-free periods/areas and to assess the vectorial role of the various *Culicoides* species, a comprehensive knowledge on their spatio-temporal occurrence is required.

Methods: Biting midges were monitored on farm sites with livestock in the defined climatic regions, including high altitudes, of Switzerland by overnight trapping at 12 locations once a week over three years using UV-light traps. Based on morphological features, they were separated into three groups (i.e. *Obsoletus*, *Pulicaris*, other *Culicoides* spp.), and identification to the species level was achieved by protein profiling using MALDI-TOF mass spectrometry.

Results: Around 550,000 biting midges in total were collected, revealing a dominance (82 to 99%) of the *Obsoletus* group species up to an altitude of 1,200 m and of the *Pulicaris* group species above 1,500 m (85% at the highest trapping site at 2,130 m). The maximum number of midges collected in a summer night (756 to 19,682) as well as the total number of midges caught over three years (from 6,933 to 149,439) varied highly among the sites, whereas the annual variation in total midge abundance at the locations was statistically insignificant. MALDI-TOF MS of 100 randomly selected individual biting midges per trapping site yielded high quality spectra for 1,187 of the 1,200 (98.9%) specimens of which 1,173 could be assigned to one of the 15 *Culicoides* species for which biomarker mass sets are available in the reference database.

Conclusions: There are no biting midge-free zones in all of the agriculturally utilized areas (including alpine summer pastures) of Switzerland. Annual variations of midge numbers at the sampled locations were low, indicating that monitoring of midges should preferably be done by investigating a large number of sites for one season instead of few locations for extended periods of time. High throughput species identification of midges by MALDI-TOF MS is feasible, and this technique adds to other recently developed methods for the identification of midges (PCRs in various formats, interactive identification keys), facilitating epidemiological and biological in-depth studies of these important insects.

Keywords: *Culicoides*, Biting midge, Vectors, *Obsoletus* group, *Pulicaris* group, Species identification, MALDI-TOF MS, Monitoring, Abundance, Climatic region

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Background

The unexpected and explosive outbreak of the blue-tongue disease in Northern and Central Europe in 2006 triggered Europe-wide activities to monitor the biological vectors of the disease, biting midges of the genus *Culicoides* (Diptera, Ceratopogonidae), with the primary aim to define vector-free periods [1-5]. Thus, overviews of seasonally vector-free periods, determined for different years, are available for a number of countries [6].

Culicoides midges are incriminated as putative vectors of other orbiviruses of relevance for Europe at present (African horse sickness virus, epizootic haemorrhagic disease virus, Toggenburg virus; [7-11]). Very recently, an Orthobunya virus has emerged in Europe ('Schmallenberg virus'), causing fever, diarrhoea, malformed new-borns and abortion [12], and this virus is assumed to be transmitted by biting midges and/or mosquitoes in analogy to the known vectors of related viruses [13]. Interestingly, biting midges of another Ceratopogonidae genus (*Forcipomyia*, subgenus *Lasiohelea*), have recently been incriminated as vectors of protozoan parasites of the genus *Leishmania* in Australia [14]. In addition to their emerging role as vectors, biting midges are a well-known nuisance pest in many parts of the world, and they can cause insect bite hypersensitivity ('sweet itch'), particularly in equids, but the species that cause the clinical symptoms under field conditions are not known [15].

The identification of the tiny (1–3 mm) biting midges is primarily carried out using morphological features, and a recently developed interactive identification key will be of great practical value [16]. The wing patterns allow for a rapid classification of the insects into groups (Obsoletus group, Pulicaris group, other *Culicoides* [17] www.culicoides.net). Certain species can be identified with their wing pattern only, while others require time consuming microscopic analysis of slide-mounted insect preparations [18,19]. Therefore, morphological identification can be a time-consuming procedure and is known to be a very difficult task in many cases even for expert taxonomists [20], due to faint characteristics or intraspecific variability [21]. Furthermore, the existence of cryptic species, i.e. morphologically indistinguishable midges that are genetically distinct, has recently been described [21,22]. Hence, in large entomological surveys, trapped midges are for practical reasons first grossly separated to group level (Obsoletus, Pulicaris, other *Culicoides*) and further identified to species level depending on the available resources. Generally, species of the Obsoletus group dominated in European monitoring projects e.g. [2-5,23,24]. In a recent study in Austria, for example, a nation-wide monitoring at 54 locations at altitudes from 116 to 1,190 meters above sea level (m.a.s.l.) and run over 15 months revealed 90.2% of the biting midges as

belonging to the Obsoletus group, 5.3% to the Pulicaris group, and 4.5% to other *Culicoides* [3].

Alternative approaches to midge identification include PCR. Several such assays in various formats have been developed for the specific detection of a number of *Culicoides* species [summarized in 22], including a microarray format for the identification of Obsoletus group species [25]. Recently, mass spectrometry (matrix-assisted laser desorption/ionization time of flight mass spectrometry, MALDI-TOF MS) has been evaluated as a diagnostic tool [26], and a reference database of biomarker masses covering 15 *Culicoides* species, including an abundant cryptic species, was established [27].

The aims of the present study were to determine over three years the spatio-temporal dynamics of biting midges in the defined climatic regions, including high altitudes, of Switzerland, and to apply MALDI-TOF MS for the species identification of a large number of field-collected specimens.

Methods

Trapping locations

The 12 trap locations were chosen in the vicinity of Swiss national basic climatological network reference stations (though no meteorological data were evaluated in the work presented) in 11 of the 12 defined climatic regions of the country [28] (Table 1). From the three climatic regions along the northern slope of the Alps (east, central, west), only two were considered (east, central). Instead, an additional second trap at high altitude (municipality 'Juf', 2,130 m) was run in the alpine region 'North and Central Grisons'. The sites were distributed all over the country, covering low (270 m) and high (2,130 m) altitudes, areas with Atlantic climate and climate influenced by the Mediterranean Sea, and areas within and on both sites of the two mountain ranges (Alps, Jura).

Insect collection

Biting midges were caught on farms using the Onderstepoort blacklight suction traps [29] operated once per week, from approximately 2 h before dusk and until after dawn [17]. Criteria applied for the placement of the traps [17] were 1) presence of more than 10 head of ruminant livestock, i.e. cattle, sheep, goats or horses usually within 25 m, 2) forest and naturally created water pools, streams or swamps in neighbourhood, 3) electricity available, and 4) agreement of the farmer to operate the traps. The traps were placed outdoors at weatherproof sites, ideally on the stable wall at a height between 1.5 and 2.0 m above the ground, and with no strong light sources in the immediate vicinity. The insects were collected into 200 ml bottles containing 70% ethanol and which were sent to our institute every fortnight. The monitoring

Table 1 Features of the trapping locations and general entomological results of the 3 year monitoring

Location (municipality) Coordinates ¹ (N/E)	Altitude ¹	Climate region ² (specification)	No. analysable trappings ³	Total no. <i>Culicoides</i> / other insects	Overall <i>Culicoides</i> (%)		
					Obsoletus group	Pulicaris group	other <i>Culicoides</i>
Novazzano 45°50'30"/8°58'46"	270	South of the Alps ⁴ (border to Italy)	97	19,307 / 132,004	88.5	1.8	9.7
Dittingen 47°26'31"/7°29'46"	360	Eastern Jura mountain range (low altitude at northern slope)	109	149,439 / 113,677	98.6	1.0	0.4
Commugny 46°19'20"/6°10'24"	420	Western lowlands (shore of lake Geneva)	150	6,933 / 247,425	94.8	3.1	2.1
Granges 46°15'38"/7°27'52"	500	Wallis (low altitude inner alpine valley with east-west orientation)	140	11,329 / 33,972	91.8	6.6	1.6
Wädenswil 47°12'49"/8°39'53"	620	North-eastern lowlands	145	64,358 / 144,627	94.4	4.0	1.6
Mühlethurnen 46°48'50"/7°30'41"	670	Central lowlands	146	18,070 / 155,267	96.4	2.2	1.4
Bennau 47°8'55"/8°43'45"	810	Northern slope of the Alps, central	112	24,009 / 164,150	91.4	7.2	1.4
Château d'Oex 46°28'27"/7°7'48"	940	Northern slope of the Alps, west	148	32,990 / 119,144	83.4	12.5	4.1
Chaumont 47°1'38"/6°57'25"	1,110	Western Jura mountain range (mountain top)	155	80,906 / 64,977	81.8	17.6	0.6
Davos 46°47'59"/9°49'51"	1,560	North and Central Grisons (high altitude inner alpine valley with north-south orientation)	122	16,488 / 15,764	13.0	78.5	8.5
Sils 46°25'48"/9°45'48"	1,800	Engadin (high altitude inner alpine valley with east-west orientation)	95	57,858 / 19,531	16.3	65.9	17.8
Juf 46°26'43"/9°34'45"	2,130	North and central Grisons (highest permanently inhabited village in Europe)	155	65,715 / 68,564	2.0	85.8	12.2

Coordinates (latitude north/ longitude east) and altitudes (meters above sea level) according to Google Earth. Locations are listed in ascending order with regard to altitude. The six trapping locations with high sampling reliability (≥ 145 weekly catches) are in bold.

² As defined and described [28].

³ Onderstepoort blacklight suction traps operated once per week, from approximately 2 h before dusk and until after dawn, for 3 years (no. = weeks, maximum number of trappings = 156).

⁴ Climate influenced by the Mediterranean Sea.

was carried out from June 2008 to May 2011 (total 156 weeks).

Insect identification

Morphology

In a first step, *Culicoides* specimens were separated from the other insects using their characteristic features of wings, antennae, and legs. In a second step, females only were separated based on wing morphology into the Obsoletus group (*C. chiopterus*, *C. obsoletus*, *C. scoticus* and *C. dewulfi*, the latter phylogenetically not belonging to this group [30]), the Pulicaris group (*C. deltus*, *C. grisescens*, *C. impunctatus*, *C. lupicaris*, *C. newsteadi*, *C. pulicaris*, *C. punctatus*) and other *Culicoides* species. From catches that were estimated

to contain more than 500 *Culicoides*, subsamples were analysed as described [31].

MALDI-TOF MS

MALDI-TOF MS-based identification of biting midges was carried out as described [27]. Briefly, the abdomens of the *Culicoides* specimens were removed, the remaining parts air-dried for approx. 2–4 min and individually transferred to 1.5 ml Eppendorf tubes where they were triturated in 10 μ l of 25% formic acid using a manual homogenizer (Bio Vortexer, Fisher Scientific, Wohlen, Switzerland) with disposable pellet pestles. One μ l of the homogenate was spotted in duplicate onto a steel target plate, air dried at room temperature for approx. 15 min, and 1 μ l of SA matrix (saturated solution

of sinapic acid in 60% acetonitrile, 40% H₂O, 0.3% trifluoroacetic acid; all chemicals from Sigma-Aldrich, Buchs, Switzerland) was added directly onto the spots. After air-drying for 15 min, the plates were sent by overnight courier to a commercial company (Mabritec SA, Riehen, Switzerland) where the mass fingerprints were generated as described [27] and subjected to automated identification against >4,000 validated biomarker mass sets, including 15 *Culicoides* species-specific ones (Obsoletus group: *C. chiopterus*, *C. dewulfi*, *C. obsoletus*, *C. scoticus*; Pulicaris group: *C. deltus*, *C. griseescens* I, *C. griseescens* II {cryptic species [22]}, *C. lupicaris*, *C. pulicaris*, *C. punctatus*; and other *Culicoides*: *C. circumscriptus*, *C. festivipennis*, *C. imicola*, *C. nubeculosus*, *C. pallidicornis*) or, in case of no or low (<90%) identification value, to full spectra comparison against all available spectra (currently approx. 60,000 including all insect reference whole spectra).

PCR/sequencing

DNA was isolated from the retained abdomens with a kit (QIAamp DNA Mini Kit, Qiagen, Hilden, Germany) and the species determined by PCR/sequencing as described [22], sequencing by Synergene GmbH (Schlieren, Switzerland).

Statistics

Kruskal-Wallis tests (PASW Statistics 18 of SPSS) were used to detect temporal differences in the trapping success for all *Culicoides* and for the different *Culicoides* groups. Levels of significance ($p < 0.05$) are given without and with Bonferroni correction to correct for multiple testing [32].

Results

Culicoides biting midges were monitored at 12 locations in Switzerland over three years by overnight catching once a week. A total of nearly 550,000 biting midges were collected, together with around 1.3 million other insects as by-catch that entered the Onderstepoort black-light traps (Table 1). The biting midges were separated by morphology into Obsoletus group, Pulicaris group and other *Culicoides*, with the first group dominating the Swiss midge fauna up to an altitude of 1,200 m, i.e. accounting for more than 80% of the midges. Farther up, the species of the Pulicaris group prevailed (85% of the biting midges at the highest trapping site at 2,130 m). Notable abundances of other *Culicoides* species were only observed at the single trapping site south of the Alps as well as at the three sites of high (>1,500 m) altitudes (Table 1).

The maximum number of midges collected in a single summer night largely varied among the sites, ranging from 756 (Commugny, altitude 420 m, trapping date

4 June 2008) to 19,682 (Dittigen, 360 m, 11 June 2008). Interestingly, the two highest altitude sites ranked second ($n=18,129$, Sils, 1,800 m, 02 August 2010) and fourth ($n=9,633$, Juf, 2,130 m, 12 August 2009) (not shown). Also, the overall number of midges collected over the three years strongly varied at the different locations (from 6,933 to 149,439; Table 1).

For further analyses, only the six trapping locations with high sampling reliability (i.e. at least 145 of the maximal 156 weekly catches available) were considered (see farms highlighted in grey in Table 1). The seasonal dynamics of the biting midges at these six sites over the three investigated years (June to May) is shown in Figure 1, displaying the monthly averages of the catches. Significant activity (more than 10 midges per trap and night) was observed between April and November, and on average less than one midge was collected during the winter months (December to March) with the exception of the location 'Juf' where a low abundance of around 10 midges were observed during the second and third trapping year (Figure 1F).

The variability of midge abundance (total *Culicoides* as well as the three different *Culicoides* groups separately) at the sites between the 3 years was analysed with the catches from June to September, which account for more than 80% of the annual midge collection in the vast majority of cases (Table 2). Statistically significant annual variation in midge abundance was neither found for the total number of midges nor for the majority of analyses of the midges belonging to the three groups (Table 2). Statistically significant variability between the years was determined in three instances (sites 'Wädenswil' and 'Mühlethurnen' for the Pulicaris group midges, site 'Château d'Oex' for the other *Culicoides* group, Table 2), all relating to midge groups with low abundances at these locations. Furthermore, only the variation of the numerically marginal Pulicaris group from 'Mühlethurnen' over the 3 years remained significant after Bonferroni correction for multiple testing (Table 2).

One hundred randomly selected individual biting midges per trapping site, all collected in late summer 2010 (period of highest abundance) and stored in 70% ethanol for more than 1 year, were subjected to MALDI-TOF MS analyses (Table 3). Thus, high quality spectra were obtained for 1,187 of the 1,200 (98.9%) analysed specimens of which 1,173 could be assigned to one of the 15 *Culicoides* species for which biomarker mass sets were available in the reference database. PCR/sequencing was done with six randomly chosen (www.random.org) specimens per site, confirming in all cases the MALDI-TOF MS result (*C. obsoletus*, $N=38$; *C. griseescens* II, $N=13$; *C. scoticus*, $N=11$; *C. griseescens* I, $N=5$, *C. dewulfi*, $N=2$; 1 each *C. circumscriptus*, *C. deltus*, *C. lupicaris*, Table 3). Fourteen specimens yielded no MALDI-TOF

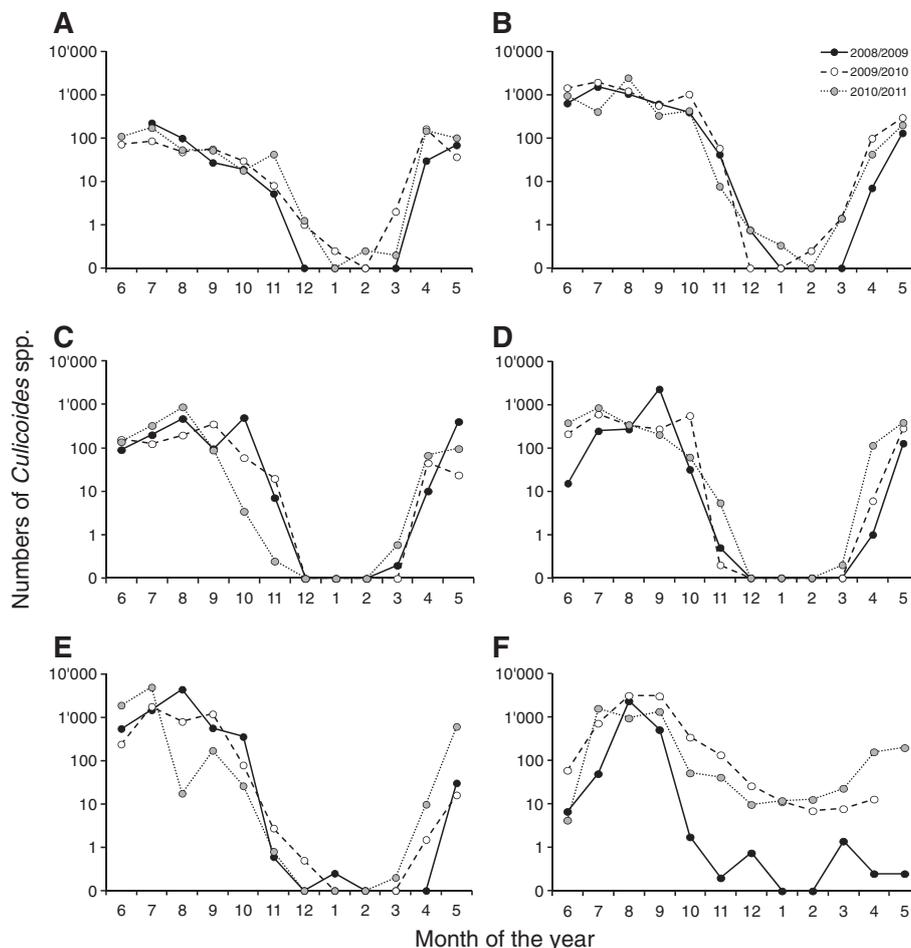


Figure 1 Seasonal dynamics of biting midges at six trapping sites. Data from the locations with high overall sampling reliability, i.e. at least 145 of the maximal 156 weekly catches available; see Table 1: **A**) Commugny (altitude 420 m); **B**) Wädenswil (620 m); **C**) Mühlethurnen (670 m); **D**) Chateau d'Oex (940 m); **E**) Chaumont (1,110 m); **F**) Juf (2,130 m). The monthly averages of total number of collected *Culicoides* biting midges are shown by black (season 2008/2009), open (2009/2010) and grey (2010/2011) circles. Note the logarithmic scale.

MS identification, i.e. no corresponding reference spectra have as yet been deposited in the database. These 14 insects were identified by PCR/sequencing as *C. fascipennis* (n = 5), *C. reconditus* (3), *C. kibunensis* (2), *C. segnis*-like (2) and 1 each of *C. newsteadi* and *C. segnis*. Finally, thirteen insects (1.1%) could not be identified by MALDI-TOF MS due to low quality spectra (low signal/noise ratio, few masses due to signal suppression).

The MALDI-TOF-MS-based identification corresponded well with the rough morphological classification into the three groups *Obsoletus*, *Pulicaris*, other *Culicoides*. For example, the corresponding percentage of insects belonging to these groups as identified morphologically or by mass spectrometry were 91.4/89, 1.9/3 and 6.7/8, respectively, for the sample from the location 'Novazzano' (Table 3). This confirms the reliability of morphological identification to the group level when done by experienced personnel.

Discussion

The spatio-temporal occurrence of *Culicoides* biting midges was determined in Switzerland over 3 years at 12 locations with one trap per site, at altitudes from 270 to 2,130 m. The six trapping locations (up to 670 m; Table 1) were part of a national monitoring system that included an additional 12 trapping sites in Switzerland and one in Liechtenstein up to a maximal altitude of 870 m. These 13 traps were only run from October until May to determine the vector-free period (i.e. less than five parous midges per trap night [33]) which is of high relevance when restriction measures for the movement of livestock are applied during an outbreak. Thus, for these two countries, the vector-free period lasts from week 47–49 to week 12–14, depending on weather conditions (for vector-free periods in other European countries see [6]). In order to also identify putatively vector-free areas, which are conceivable as retreat areas

Table 2 Statistical comparison¹ of the number of *Culicoides* collected at six locations²

Site (municipality)	Year (no. of trappings)	Total no. <i>Culicoides</i> (% ³) <i>p</i> -value	No. <i>Culicoides</i> of morphological group (% ³)		
			Obsoletus	Pulicaris	Other <i>Culicoides</i>
			<i>p</i> -value	<i>p</i> -value	<i>p</i> -value
Commugny	2008 (14)	1,440 (75.4)	1,234 (73.4)	159 (91.4)	47 (85.3)
	2009 (18)	1,130 (55.9)	1,079 (55.5)	15 (68.2)	36 (64.3)
	2010 (19)	1,751 (58.8)	1,704 (58.3)	10 (66.7)	37 (97.4)
		0.622	0.564	0.102	0.969
Wädenswil	2008 (19)	16,603 (88.7)	15,022 (88.1)	1,215 (94.3)	366 (100.0)
	2009 (17)	23,306 (91.7)	22,387 (91.8)	564 (87.3)	355 (98.6)
	2010 (19)	19,351 (96.1)	18,495 (96.1)	576 (93.8)	280 (97.2)
		0.588	0.574	0.040*	0.663
Mühlethurnen	2008 (18)	4,179 (58.3)	4,011 (59.5)	98 (30.3)	70 (73.7)
	2009 (17)	3,712 (87.9)	3,617 (88.3)	51 (82.3)	44 (66.7)
	2010 (19)	6,015 (90.0)	5,907 (89.9)	18 (81.8)	90 (96.8)
		0.889	0.857	<0.008*†	0.825
Château d'Oex	2008 (18)	13,516 (95.7)	11,991 (95.7)	1,350 (96.3)	175 (95.1)
	2009 (16)	7,177 (81.9)	5,269 (82.3)	1,390 (87.3)	518 (67.7)
	2010 (19)	7,655 (77.2)	6,333 (75.3)	937 (83.7)	385 (99.5)
		0.241	0.260	0.390	0.018*
Chaumont	2008 (19)	28,859 (94.9)	21,095 (93.5)	7,601 (99.0)	163 (99.4)
	2009 (18)	16,725 (98.8)	12,596 (98.5)	3,951 (99.8)	178 (100.0)
	2010 (19)	30,076 (92.3)	28,090 (93.5)	1,840 (77.0)	146 (100.0)
		0.763	0.709	0.311	0.931
Juf	2008 (16)	12,215 (99.9)	165 (99.4)	11,158 (99.9)	892 (99.9)
	2009 (19)	31,635 (94.4)	780 (90.6)	26,026 (93.7)	4,829 (98.8)
	2010 (18)	17,145 (89.7)	203 (83.2)	15,006 (89.8)	1,936 (90.0)
		0.336	0.270	0.466	0.069

¹ Kruskal-Wallis test; catches from the summer months (June to September) during the 3 years monitoring.

² Locations with high overall sampling reliability (i.e. at least 145 of the maximal 156 weekly catches available, see Table 1).

³ Percentage of total *Culicoides* caught over the whole collecting season (June – May).

* Significantly different using the Kruskal-Wallis test without Bonferroni correction ($p < 0.05$).

† Significantly different using the Kruskal-Wallis test with Bonferroni correction ($p < 0.008$).

for animals during outbreaks of midge-borne diseases, six traps were placed at locations not covered by the national monitoring, particularly at higher altitudes. Monitoring activities in other European countries focussed on locations below 1,400 m (highest altitudes investigated: 1,400 m in Sicily [34]; 1,190 m in Austria [3]; 1,184 m in Central Italy [35]). The highest record of *Culicoides* in Switzerland was 1,600 m (from a zoological study [36]). We determined high abundances at the high altitude locations (1,800, 2,130 m; Table 1). In accordance with earlier preliminary studies [37,38], midges of the Pulicaris group consistently prevailed at high altitudes, as is the case at high latitudes in Scandinavia [5,39]. Nevertheless, biting midges of the Pulicaris group occasionally (single farms, few catches) occurred in higher numbers than the largely dominating Obsoletus group species in Central Europe [40,41]. However, in only one

of our total 1,202 catches from the nine locations below 1,110 m altitude (Table 1) were the Pulicaris group species the most abundant (not shown). Species identification of 100 randomly selected midges per locations (by MALDI-TOF MS, see below) revealed a vast predominance of *C. obsoletus* (total 67.6%) and *C. scoticus* (24.8%) at the nine low altitude locations and of the cryptic species *C. grisescens* II (67.3%) at the three sites above 1,500 m. Highly interestingly, single specimens of *C. obsoletus* and *C. scoticus* were identified in the Alpine region and, vice versa, of *C. grisescens* II in the lowlands (Table 3), putatively due to the (albeit rare) long range dispersal of these tiny insects [42] and references cited therein. Further, whereas *C. obsoletus* was dominating in eight of the nine lower altitude locations, *C. scoticus* accounted for 95 of the 100 midges at the ninth location ('Dittingen', Table 3). As these analyses were done with

Table 3 MALDI-TOF MS based species identification of 100 randomly selected *Culicoides* specimens from the 12 trapping sites¹

Site (altitude ²)	<i>Culicoides</i> species (no.) {no. confirmed by PCR} ³ [% ⁴]				
	Morphological group			Unknown Genetic identification ⁴	No identification ⁵
	Obsoletus	Pulicaris	Other <i>Culicoides</i>		
Novazzano (270)	<i>C. obsoletus</i> (84) {5}	<i>C. grisescens</i> II (1)	<i>C. circumscriptus</i> (4) {1}	0	0
	<i>C. scoticus</i> (5)	<i>C. pulicaris</i> (1)	<i>C. pallidicornis</i> (4)		
	[91.4]	<i>C. punctatus</i> (1)			
Dittingen (360)	<i>C. obsoletus</i> (3)	<i>C. lupicaris</i> (2)	0	0	0
	<i>C. scoticus</i> (95) {6}				
	[99.9]	[0.1]	[0]		
Commugny (420)	<i>C. obsoletus</i> (91) {6}	0	0	<i>C. kibunensis</i> (2)	3
	<i>C. scoticus</i> (4)				
	[98.4]	[0]	[1.6]		
Granges (500)	<i>C. obsoletus</i> (88) {6}	<i>C. grisescens</i> II (1)	0	0	0
	<i>C. scoticus</i> (9)	<i>C. pulicaris</i> (2)			
	[100]	[0]	[0]		
Wädenswil (620)	<i>C. obsoletus</i> (91) {5}	<i>C. lupicaris</i> (5) {1}	<i>C. pallidicornis</i> (1)	0	0
	<i>C. scoticus</i> (2)	<i>C. pulicaris</i> (1)			
	[95]	[4.2]	[0.8]		
Mühlethurnen (670)	<i>C. obsoletus</i> (74) {4}	<i>C. lupicaris</i> (1)	0	<i>C. segnis</i> (1)	3
	<i>C. scoticus</i> (19) {1}				
	<i>C. dewulfi</i> (2) {1}				
	[99.0]	[1.0]	[0]		
Bennau (810)	<i>C. obsoletus</i> (74) {4}	<i>C. grisescens</i> II (1) {1}	<i>C. pallidicornis</i> (1)	0	4
	<i>C. scoticus</i> (11) {1}	<i>C. lupicaris</i> (9)			
	[92.1]	[5.6]	[2.2]		
Château d'Oex (940)	<i>C. obsoletus</i> (91) {6}	<i>C. lupicaris</i> (1)	0	<i>C. newsteadi</i> (1)	0
	<i>C. scoticus</i> (3)	<i>C. pulicaris</i> (3)			
		<i>C. punctatus</i> (1)			
	[90.6]	[4.7]	[4.7]		
Chaumont (1,110)	<i>C. obsoletus</i> (12) {2}	<i>C. lupicaris</i> (1)	0	0	3
	<i>C. scoticus</i> (75) {3}	<i>C. pulicaris</i> (1)			
	<i>C. chiopterus</i> (6)	<i>C. punctatus</i> (1)			
	<i>C. dewulfi</i> (1) {1}				
	[94.2]	[1.9]	[3.9]		
Davos (1,560)	<i>C. obsoletus</i> (5)	<i>C. grisescens</i> I (4) {1}	<i>C. pallidicornis</i> (1)	0	0
	<i>C. scoticus</i> (2)	<i>C. grisescens</i> II (87) {5}			
		<i>C. pulicaris</i> (1)			
	[4.2]	[95.0]	[0.8]		
Sils (1,800)	<i>C. obsoletus</i> (4)	<i>C. deltus</i> (6) {1}	0	<i>C. fascipennis</i> (2)	0
	<i>C. scoticus</i> (3)	<i>C. grisescens</i> I (5)		<i>C. reconditus</i> (1)	
		<i>C. grisescens</i> II (75) {5}			
		<i>C. pulicaris</i> (4)			
	[4.5]	[82.9]	[12.6]		

Table 3 MALDI-TOF MS based species identification of 100 randomly selected *Culicoides* specimens from the 12 trapping sites¹ (Continued)

Juf	<i>C. grisescens</i> I (51) {4}	0	<i>C. fascipennis</i> (3)	0
(2,130)	<i>C. grisescens</i> II (40) {2}		<i>C. reconditus</i> (2)	
	<i>C. pulicaris</i> (2)		<i>C. segnis</i> -like (2) ⁶	
	[1.5]	[94]	[4.5]	

¹ Specimens randomly selected from a single catch from the high abundance season (August/September 2010), stored in 70% ethanol for more than 1 year and analysed using a database containing validated biomarker mass sets of 15 *Culicoides* species [27].

² Meters above sea level.

³ PCR/sequencing [22] with 6 randomly chosen (www.random.org) specimens/site. Species identification by comparison with GenBank entries (>99% identity).

⁴ Percentage of *Culicoides* morphologically classified as belonging to Obsoletus group, Pulicaris group or other *Culicoides* of the catch from which the specimens were selected for the MALDI-TOF MS analyses.

⁴ Identified by PCR/sequencing [22].

⁵ No species identification achievable due to poor quality mass spectra.

⁶ Best match: 96% identity (403/422) with GenBank entry HQ824509 (*Ccxx segnis* from Switzerland).

insects from a single catch per location, obtained in summer, further analyses are required to understand the spatio-temporal population dynamics of the various species.

We observed a considerable range of the total number of collected biting midges between the locations. Such a variability between sites/farms is well known from many other studies e.g. [20,38,43-45]. In a previous study in Switzerland, for example, the total number of biting midges collected over one season at two farms only 4 km apart and located at the same altitude differed by the factor 24 [38]. Major factors influencing the abundance of midges are particularly topoclimate, land use and soil (as proxy for larval breeding sites) [35,46-48]. However, when comparing the number of midges we collected at the locations over three consecutive summers (June – September), statistically significant differences were neither observed for the total number nor for the vast majority of analyses of the three *Culicoides* groups (Table 2). Thus, collection of midges over the four summer months during one single year sufficed to obtain a representative picture of their abundances at a given location.

Our study was running over 3 years, adding up to a maximum total number of 156 weekly trappings per location. At six of the 12 locations, more than 145 catches were made (Table 1), and the data from these locations were used to depict the seasonal dynamics (Figure 1) and to analyse yearly differences in abundance (Table 2). The lower number of trappings at the other locations was due to technical problems (unnoticed failure of light bulbs/ventilator); missing/incorrect labelling of catches; independent decisions taken by farmers to skip trapping during adverse (mainly cold) weather conditions, and by simply forgetting to operate the traps by the farmers. A somewhat surprising and unique picture was obtained from the highest altitude trapping location (2,130 m) where, throughout two winters low midge activity was observed (Figure 1). We speculate that midges from the inside population escaped through shakes in the wooden

wall of this barn. Such indoor populations of biting midges can reach considerable sizes [49-52] albeit these populations are also strongly reduced in winter [50] and own unpublished results from another trapping location.

MALDI-TOF MS has come of age for high throughput, accurate and reproducible identification of medically relevant microorganisms (bacteria, yeasts, filamentous fungi) at low costs and minimal preparation time [53]. Only recently, this proteomic approach has become available for *Culicoides* identification, relying on a validated reference database of biomarker mass sets from 15 *Culicoides* species [27], all but one (*C. imicola*) being indigenous to Switzerland. Analyses of 1,200 biting midges with MALDI-TOF MS confirmed the method's reliability, as 98.9% of the specimens yielded high quality spectra, and 97.8% of the midges could be assigned to one of the species (Table 3) included in the database [27]. Obviously, the database covers the most abundant species of Central Europe. It is not clear how many indigenous *Culicoides* species exist in Switzerland. A compilation based on published data lists 35 established valid species [54]. In comparison, 51 species have been listed for north-eastern France, a region which has thoroughly been studied [18] (Delécolle, personal communication). However, several new species as well as specimens that could not unequivocally be identified by morphology have recently been reported from Switzerland [22,23,37,45]. In addition, cryptic species have been reported [21,22], and the genetic identification of two midges from the highest altitude trapping site as *C. segnis*-like (with 96% sequence identity to a *C. segnis* GenBank entry) indicates that the taxonomy of *Culicoides* midges remains an unfinished story.

No biomarker mass sets existed for 14 specimens, which therefore could not be assigned to a species. These 14 insects belonged to six species as identified by PCR/sequencing (Table 3), including five *C. fascipennis*. As the biomarker mass set for a species in general is derived from the reference spectra of at least five

genetically confirmed specimens [27], a *C. fascipennis*-specific biomarker mass set comprising 29 masses (not shown) could be derived and added to the database.

Poor quality mass spectra, yielding no information with regard to species or group affiliation, were obtained for 13 (1.1%) specimens. The main source for this failure is most probably insufficient homogenization, which was done by a hand-held homogenizer, and automated sample preparation is desirable. With the already very high rate of good quality spectra (98.9%) obtained using a 'quick and dirty' preparation, it seems doubtful whether the evaluation of laborious refinements of pre-analytical processes might be a worthwhile expedient approach to further increase the efficiency of MALDI-TOF MS analyses.

Thus, MALDI-TOF MS is a new tool available for high throughput *Culicoides* species identification. This method is particularly economic for approaches requiring detailed and quantitative information on the midge fauna (to gain a rapid overview on the species present; to follow their spatio-temporal occurrence; to identify morphologically similar or indistinguishable species, e.g. *C. obsoletus* and *C. scoticus*) whereas PCR performed on DNA from pools of midges remains the method of choice for tracking down one or a few species of interest.

The foundation for MALDI-TOF MS analyses is the availability of a database with reference biomarker masses from carefully confirmed reference specimens. As the creation and maintenance of such a database is a tedious task, a centralised structure seems to offer an efficient solution. The database we rely on was created in collaboration between our group and a private company (Mabritec SA, Riehen, Switzerland) and, as shown in this work, enlarges when being utilized. Thus, this database might be of value as the core of an eventual comprehensive *Culicoides* database.

Conclusion

There are no biting midge-free zones in the agriculturally utilized areas (including alpine summer pastures) of Switzerland, with midges of the *Obsoletus* group, which are considered the main vectors of bluetongue virus, dominating up to altitudes as high as 1,200 m. Above 1,500 m, the *Pulicaris* group whose species' vector competences are largely unknown prevails with high populations. Our three-year monitoring revealed highly different numbers of *Culicoides* spp. collected at the 12 sites, but annual variations at the various locations were statistically insignificant, indicating that monitoring of midges preferably is done by investigating large numbers of sites for one season instead of few locations during extended periods of time. High throughput species identification of midges by MALDI-TOF MS is feasible. This technique adds to other recently developed methods for the identification of midges (PCRs in various

formats, interactive identification keys), facilitating epidemiological and biological in-depth studies of these important insects.

Competing interests

The authors declare that they have no competing interests.

Authors' contributions

AM and FS conceived the study; FS supervised the morphological and AM the genetical identification; CK and IS supervised and carried out the field and laboratory work; CK and DH analysed the data and interpreted the results; AM wrote the first draft of the paper, and all authors contributed to the final manuscript which they approve.

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References

1. Meiswinkel R, Baldet T, de Deken R, Takken W, Delécolle J-C, Mellor PS: **The 2006 outbreak of bluetongue in northern Europe - the entomological perspective.** *Prev Vet Med* 2008, **87**:55–63.
2. Hoffmann B, Bauer B, Bauer C, Bätza H-J, Beer M, Clausen P-H, Geier M, Gethmann JM, Kiel E, Liebisch G, Liebisch A, Mehlhorn H, Schaub GA, Werner D, Conraths FJ: **Monitoring of putative vectors of bluetongue virus serotype 8, Germany.** *Emerg Infect Dis* 2009, **15**:1481–1484.
3. Silbermayr K, Hackländer K, Doscher C, Koefer J, Fuchs K: **A spatial assessment of *Culicoides* spp. distribution and bluetongue disease risk areas in Austria.** *Berl Munch Tierarztl* 2011, **124**:10–17.
4. Berisha B, Goga I, Hulaj B, Caushi D, Sherifi K, Wilsmore AJ, Taylor WP: **Entomological research on the vectors of bluetongue disease and the monitoring of activity of *Culicoides* in the Prishtine region of Kosova.** *Vet Ital* 2010, **46**:431–437.
5. Ander M, Meiswinkel R, Chirico J: **Seasonal dynamics of biting midges (Diptera: Ceratopogonidae: *Culicoides*), the potential vectors of bluetongue virus, in Sweden.** *Vet Parasitol* 2012, **184**:59–67.
6. Anonymous: *Bluetongue*. European Commission Health and Consumers; [http://ec.europa.eu/food/animal/diseases/controlmeasures/bluetongue_en.htm].
7. Zimmerli U, Herholz C, Schwermer H, Hofmann M, Griot C: **Afrikanische Pferdepest und equine Encephalosis: Muss sich die Schweiz vorbereiten?** *Schweiz Arch Tierh* 2010, **152**:165–175.
8. Mellor PS, Hamblin C: **African horse sickness.** *Vet Res* 2004, **35**:445–466.
9. Chagnat V, Worwa G, Scherrer N, Hilbe M, Ehrensperger F, Batten C, Cortyen M, Hofmann M, Thuer B: **Toggenburg Orbivirus, a new bluetongue virus: initial detection, first observations in field and experimental infection of goats and sheep.** *Vet Microbiol* 2009, **138**:11–19.
10. Anonymous: **Scientific opinion on epizootic hemorrhagic disease.** *EFSA J* 2009, **7**:1418.
11. Yadin H, Brenner J, Bumbrov V, Oved Z, Stram Y, Klement E, Perl S, Anthony S, Maan S, Batten C, Mertens PP: **Epizootic haemorrhagic disease virus type 7 infection in cattle in Israel.** *Vet Rec* 2008, **162**:53–56.
12. Hoffmann B, Scheuch M, Höper D, Jungblut R, Holsteg M, Schirmer H, Eschbaumer M, Goller KV, Wernike K, Fischer M, Breithaupt A, Mettenleiter TC: **Novel orthobunyavirus in cattle, Europe, 2011.** *Emerg Infect Dis* 2011, **18**:469–472.
13. Lehmann K, Werner D, Hoffmann B, Kampen H: **PCR identification of culicoid biting midges (Diptera, Ceratopogonidae) of the *Obsoletus* complex including putative vectors of bluetongue and Schmallenberg viruses.** *Parasit Vectors* 2012, **5**:213.
14. Dougall AM, Alexander B, Holt DC, Harris T, Sultan AH, Bates PA, Rose K, Walton SF: **Evidence incriminating midges (Diptera: Ceratopogonidae) as potential vectors of *Leishmania* in Australia.** *Int J Parasitol* 2011, **41**:571–579.
15. Van Oldruitenborgh-Oosterbaan MM S, Van Poppel M, De Raat IJ, Van den Boom R, Savelkoul HF: **Intradermal testing of horses with and without**

- insect bite hypersensitivity in The Netherlands using an extract of native *Culicoides* species. *Vet Dermatol* 2009, **20**:607–614.
16. Mathieu B, Cêtre-Sossah C, Garros C, Chavernac D, Balenghien T, Carpenter S, Setier-Rio M-L, Vignes-Lebbe R, Ung U, Candolfi E, Delécolle JC: Development and validation of ILC: an interactive identification key for *Culicoides* (Diptera: Ceratopogonidae) females from the Western Palaearctic region. *Parasit Vectors* 2012, **5**:137.
 17. Goffredo M, Meiswinkel R: Entomological surveillance of bluetongue in Italy: methods of capture, catch analysis and identification of *Culicoides* biting midges. *Vet Ital* 2004, **40**:260–265.
 18. Delécolle J-C: *Nouvelle contribution à l'étude systématique et iconographique des espèces du genre Culicoides (Diptera: Ceratopogonidae) du nord-est de la France*. PhD thesis. Strasbourg: Université Louis Pasteur; 1985.
 19. Campbell JA, Pelham-Clinton EC: A taxonomic review of the British species of *Culicoides* Latreille (Diptera, Ceratopogonidae). *Proc Roy Soc Edinb B* 1960, **67**:181–302.
 20. Meiswinkel R, Goffredo M, Leijts P, Conte A: The *Culicoides* 'snapshot': a novel approach used to assess vector densities widely and rapidly during the 2006 outbreak of bluetongue (BT) in The Netherlands. *Prev Vet Med* 2008, **87**:98–118.
 21. Pages N, Munoz-Munoz F, Talavera S, Sarto V, Lorca C, Nunez JI: Identification of cryptic species of *Culicoides* (Diptera: Ceratopogonidae) in the subgenus *Culicoides* and development of species-specific PCR assays based on barcode regions. *Vet Parasitol* 2009, **165**:298–310.
 22. Wenk C, Kaufmann C, Schaffner F, Mathis A: Molecular characterisation of Swiss Ceratopogonidae (Diptera) and evaluation of real-time PCR assays for the identification of *Culicoides* biting midges. *Vet Parasitol* 2012, **184**:258–266.
 23. Casati S, Raclou V, Delécolle JC, Kuhn M, Mathis A, Griot C, Stark KDC, Vanzetti T: An investigation on the *Culicoides* species composition at seven sites in southern Switzerland. *Med Vet Entomol* 2009, **23**:93–98.
 24. Venail R, Balenghien T, Guis H, Tran A, Setier-Rio M-L, Delécolle JC, Mathieu B, Cêtre-Sossah C, Martinez D, Languille J, Baldet T, Garros C: Assessing diversity and abundance of vector populations at a national scale: example of *Culicoides* surveillance in France after bluetongue virus emergence. In *Arthropods as vectors of emerging diseases*. Edited by Mehlhorn H. Berlin: Springer; 2012:77–102.
 25. Deblauwe I, de Witte JC, de Deken G, De Deken R, Madder M, Van Erk S, Hoza FA, Lathouwers D, Geysen D: A new tool for the molecular identification of *Culicoides* species of the *Obsoletus* group: the glass slide microarray approach. *Med Vet Entomol* 2012, **26**:83–91.
 26. Kaufmann C, Ziegler D, Schaffner F, Carpenter S, Pflüger V, Mathis A: Evaluation of matrix-assisted laser desorption/ionization time of flight mass spectrometry for characterization of *Culicoides nubeculosus* biting midges. *Med Vet Entomol* 2011, **25**:32–38.
 27. Kaufmann C, Schaffner F, Ziegler D, Pflüger V, Mathis A: Identification of field-caught *Culicoides* biting midges using matrix-assisted laser desorption/ionization time of flight mass spectrometry. *Parasitology* 2012, **139**:248–258.
 28. Begert M, Seiz G, Foppa N, Schlegel T, Appenzeller C, Müller G: Die Überführung der klimatologischen Referenzstationen der Schweiz in das Swiss National Basic Climatological Network (Swiss NBCN). In *Arbeitsberichte MeteoSchweiz*, Volume 215. Zürich: MeteoSchweiz; 2007:1–43.
 29. Venter GJ, Meiswinkel R: The virtual absence of *Culicoides imicola* (Diptera: Ceratopogonidae) in a light-trap survey of the colder, high-lying area of the eastern Orange Free State, South Africa, and implications for the transmission of arboviruses. *Onderstepoort J Vet Res* 1994, **61**:327–340.
 30. Schwenkenbecher JM, Mordue AJ, Piertney SB: Phylogenetic analysis indicates that *Culicoides dewulfi* should not be considered part of the *Culicoides obsoletus* complex. *Bull Entomol Res* 2009, **99**:371–375.
 31. Van Ark H, Meiswinkel R: Subsampling of large light trap catches of *Culicoides* (Diptera: Ceratopogonidae). *Onderstepoort J Vet Res* 1992, **59**:183–189.
 32. Bland JM, Altman DG: Multiple significance tests: the Bonferroni method. *Br Med J* 1973, **310**:170.
 33. Anonymous: Scientific opinion of the Scientific Panel on Animal Health and Welfare on the EFSA selfmandate on bluetongue origin and occurrence. *EFSA J* 2007, **480**:1–20.
 34. Torina A, Caraccappa S, Mellor PS, Baylis M, Purse BV: Spatial distribution of bluetongue virus and its *Culicoides* vectors in Sicily. *Med Vet Entomol* 2004, **18**:81–89.
 35. De Liberato C, Farina F, Magliano A, Rombola P, Scholl F, Spallucci V, Scaramozzino P: Biotic and abiotic factors influencing distribution and abundance of *Culicoides obsoletus* group (Diptera: Ceratopogonidae) in central Italy. *J Med Entomol* 2010, **47**:313–318.
 36. Kremer M, Callot J: Sur quelques *Culicoides* (Diptera: Ceratopogonidae) du Haut-Valais. *Acta Trop* 1961, **18**:254–256.
 37. Kaufmann C, Schaffner F, Mathis A: Monitoring of biting midges (*Culicoides* spp.), the potential vectors of the bluetongue virus, in the 12 climatic regions of Switzerland. *Schweiz Arch Tierh* 2009, **151**:205–213.
 38. Tschuor AC, Kaufmann C, Schaffner F, Mathis A: Occurrence of biting midges (*Culicoides* spp.) at three different altitudes in an alpine region of Switzerland. *Schweiz Arch Tierh* 2009, **151**:215–221.
 39. Nielsen SA, Callot J, Chirico J: Monitoring of biting midges (Diptera: Ceratopogonidae: *Culicoides* Latreille) on farms in Sweden during the emergence of the 2008 epidemic of bluetongue. *Parasitol Res* 2010, **106**:1197–1203.
 40. Mehlhorn H, Walldorf V, Klimpel S, Schaub G, Kiel E, Focke R, Liebisch G, Liebisch A, Werner D, Bauer C, Clausen H, Bauer B, Geier M, Horbrand T, Batza HJ, Conraths FJ, Hoffmann B, Beer M: Bluetongue disease in Germany (2007–2008): monitoring of entomological aspects. *Parasitol Res* 2009, **105**:313–319.
 41. Kiel E, Liebisch G, Focke R, Liebisch A, Werner D: Monitoring of *Culicoides* at 20 locations in northwest Germany. *Parasitol Res* 2009, **105**:351–357.
 42. Sanders CJ, Selby R, Carpenter S, Reynolds DR: High-altitude flight of *Culicoides* biting midges. *Vet Rec* 2011, **169**:208.
 43. Hörbrand T, Geier M: Monitoring of *Culicoides* at nine locations in Southern Germany (2007–2008). *Parasitol Res* 2009, **105**:387–392.
 44. Takken W, Verhulst N, Scholte E-J, Jacobs F, Jongema Y, van Lammeren R: The phenology and population dynamics of *Culicoides* spp. in different ecosystems in The Netherlands. *Prev Vet Med* 2008, **87**:41–54.
 45. Cagienard A, Griot C, Mellor PS, Denison E, Stark KDC: Bluetongue vector species of *Culicoides* in Switzerland. *Med Vet Entomol* 2006, **20**:239–247.
 46. Acevedo P, Ruiz-Fons F, Estrada R, Marquez AL, Miranda MA, Gortazar C, Lucientes J: A broad assessment of factors determining *Culicoides imicola* abundance: modelling the present and forecasting its future in climate change scenarios. *PLoS One* 2010, **5**:e14236.
 47. Conte A, Goffredo M, Ippoliti C, Meiswinkel R: Influence of biotic and abiotic factors on the distribution and abundance of *Culicoides imicola* and the *Obsoletus* complex in Italy. *Vet Parasitol* 2007, **150**:333–344.
 48. Conte A, Ippoliti C, Savini G, Goffredo M, Meiswinkel R: Novel environmental factors influencing the distribution and abundance of *Culicoides imicola* and the *Obsoletus* complex in Italy. *Vet Ital* 2007, **43**:571–580.
 49. Baldet T, Delécolle JC, Cêtre-Sossah C, Mathieu B, Meiswinkel R, Gerbier G: Indoor activity of *Culicoides* associated with livestock in the bluetongue virus (BTV) affected region of northern France during autumn 2006. *Prev Vet Med* 2008, **87**:84–97.
 50. Baylis M, Parkin H, Kreppel K, Carpenter S, Mellor PS, McIntyre KM: Evaluation of housing as a means to protect cattle from *Culicoides* biting midges, the vectors of bluetongue virus. *Med Vet Entomol* 2010, **24**:38–45.
 51. Meiswinkel R, Goffredo M, Dijkstra EGM, van der Ven IJK, Baldet T, Elbers A: Endophily in *Culicoides* associated with BTV-infected cattle in the province of Limburg, south-eastern Netherlands, 2006. *Prev Vet Med* 2008, **87**:182–195.
 52. Clausen P-H, Stephan A, Bartsch S, Jandowsky A, Hoffmann-Kohler P, Schein E, Mehlitz D, Bauer B: Seasonal dynamics of biting midges (Diptera: Ceratopogonidae, *Culicoides* spp.) on dairy farms of Central Germany during the 2007/2008 epidemic of bluetongue. *Parasitol Res* 2009, **105**:381–386.
 53. Croxatto A, Prod'homme G, Greub G: Applications of MALDI-TOF mass spectrometry in clinical diagnostic microbiology. *FEMS Microbiol Rev* 2012, **36**:380–407.
 54. Merz B, Bächli G, Haenni J-P: Zweiter Nachtrag zur Checkliste der Diptera der Schweiz. *Mitt Schweiz Entomol Gesell* 2006, **56**:135–165.

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